

## Exercise-induced bronchoconstriction update—2016

John M. Weiler, MD,\* John D. Brannan, PhD,\*‡ Christopher C. Randolph, MD,\*§ Teal S. Hallstrand, MD,‡ Jonathan Parsons, MD,‡ William Silvers, MD,‡ William Storms, MD,‡ Joanna Zeiger, MS, PhD,‡ David I. Bernstein, MD,§ Joann Blessing-Moore, MD,§ Matthew Greenhawt, MD, MBA, MSc,§ David Khan, MD,§ David Lang, MD,§ Richard A. Nicklas, MD,§ John Oppenheimer, MD,§ Jay M. Portnoy, MD,§ Diane E. Schuller, MD,§ Stephen A. Tilles, MD,§ and Dana Wallace, MD§

The first practice parameter on exercise-induced bronchoconstriction (EIB) was published in 2010. This updated practice parameter was prepared 5 years later. In the ensuing years, there has been increased understanding of the pathogenesis of EIB and improved diagnosis of this disorder by using objective testing. At the time of this publication, observations included the following: dry powder mannitol for inhalation as a bronchial provocation test is FDA approved however not currently available in the United States; if baseline pulmonary function test results are normal to near normal (before and after bronchodilator) in a person with suspected EIB, then further testing should be performed by using standardized exercise challenge or eucapnic voluntary hyperpnea (EVH); and the efficacy of nonpharmaceutical interventions (omega-3 fatty acids) has been challenged. The workgroup preparing this practice parameter updated contemporary practice guidelines based on a current systematic literature review. The group obtained supplementary literature and consensus expert opinions when the published literature was insufficient. A search of the medical literature on PubMed was conducted, and search terms

included pathogenesis, diagnosis, differential diagnosis, and therapy (both pharmaceutical and nonpharmaceutical) of exercise-induced bronchoconstriction or exercise-induced asthma (which is no longer a preferred term); asthma; and exercise and asthma. References assessed as relevant to the topic were evaluated to search for additional relevant references. Published clinical studies were appraised by category of evidence and used to document the strength of the recommendation. The parameter was then evaluated by Joint Task Force reviewers and then by reviewers assigned by the parent organizations, as well as the general membership. Based on this process, the parameter can be characterized as an evidence- and consensus-based document. (*J Allergy Clin Immunol* 2016;■■■■:■■■■-■■■■.)

**Key words:** *Exercise-induced bronchoconstriction, exercise-induced bronchospasm, exercise-induced asthma, exercise-induced bronchoconstriction pathogenesis, diagnosis, differential diagnosis and therapy, nonpharmacologic, pharmacologic*

\*Chief Editor.

‡Workgroup Contributor.

§Task Force Reviewer.

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scientific advisory council for the National Peanut Board; has consultant arrangements with Adamis Pharmaceutical, Canadian Transportation Agency, Nutricia, Nestle/ Gerber, and Aimmune; is an Associate Editor for the *Annals of Allergy, Asthma, and Immunology*; and has received payment for lectures from the American College of Allergy, Asthma, and Immunology, Reach MD, Thermo Fisher Scientific, California Society for Allergy and Immunology, the Allergy and Asthma Network, New England Society for Allergy, UCLA/Harbor Heiner Lectureship, Medscape, Western Michigan School of Medicine, Canadian Society of Allergy and Clinical Immunology, and the Pennsylvania Society for Allergy and Immunology. D. Khan has consultant arrangements with Aimmune; has received grants from the NIH, has received payment for lectures from Genentech, and has received royalties from UpToDate. D. Lang has consultant arrangements with Genentech/Novartis, Adamis, Merck, Meda, GlaxoSmithKline, and AstraZeneca; has received grants from Genentech/Novartis and Merck; and has received payment for lectures from Genentech/Novartis. J. Oppenheimer has consultant arrangements with GlaxoSmithKline, Mylan, and Meda; has received fees for participation in review activities from Quintiles and PRA; has received money from UpToDate and *Annals of Allergy*; is a member of the American Board of Allergy and Immunology; and is employed by the Pulmonary & Allergy Associates Atlantic Health System. J. M. Portnoy has received payment for lectures from Mylan and Thermo Fisher. D. Schuller declares that she has no relevant conflicts of interest. S. Tilles received grant support from Merck, Genentech, Novartis, Teva, Mylan, NIAID, Circassia, Astellas, and AstraZeneca. D. Wallace has consultant arrangements with Neohealth, Sanofi, Allergan, and Kaleo and has received payment for lectures from Mylan and MEDA. The rest of the authors declare that they have no relevant conflicts of interest.

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These parameters were developed by the Joint Task Force on Practice Parameters (JTFPP), representing the American Academy of Allergy, Asthma & Immunology (AAAAI); the American College of Allergy, Asthma & Immunology (ACAAI); and the Joint Council of Allergy, Asthma & Immunology.

The AAAAI and ACAAI have jointly accepted responsibility for establishing “Exercise-induced bronchoconstriction update—2016.” This is a complete and comprehensive document at the current time. The medical environment is a changing environment, and not all recommendations will be appropriate for all patients. Because this document incorporated the efforts of many participants, no single individual, including those who served on the JTFPP, is authorized to provide an official AAAAI or ACAAI interpretation of these practice parameters. Any request for information about or an interpretation of these practice parameters by the AAAAI or ACAAI should be directed to the Executive Offices of the AAAAI or the ACAAI.

The JTFPP understands that the cost of diagnostic tests and therapeutic agents is an important concern that can appropriately influence the workup and treatment chosen for a given patient. The JTFPP recognizes that the emphasis of our primary recommendations regarding a medication can vary, for example, depending on third-party payer issues and product patent expiration dates. However, because a given test or agent’s cost is so widely variable and there is a paucity of pharmacoeconomic data, the JTFPP generally does not consider cost when formulating practice parameter recommendations. In extraordinary circumstances, when the cost benefit of an intervention is prohibitive, as supported by pharmacoeconomic data, commentary can be provided. These parameters are not designed for use by pharmaceutical companies in drug promotion.

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## EIB EXECUTIVE SUMMARY

The first practice parameter on exercise-induced bronchoconstriction (EIB) was published in 2010. This update is required by the National Clearinghouse and JTF consistent with the requirement of an update every 5 years.

In the ensuing years, since the first publication of the EIB practice parameter, there has been increased understanding of the pathogenesis of EIB and improved diagnosis of this disorder by using objective pulmonary function tests. At the time of this publication, dry powder mannitol for inhalation is no longer available in the United States but is available in many other countries.

If baseline pulmonary function test results are normal to near normal (before and after bronchodilator) in a person with suspected EIB, then further testing should be performed by using a standardized exercise challenge or eucapnic voluntary hyperpnea (EVH).

Since 2010, the efficacy of nonpharmaceutical interventions, such as omega-3 fatty acids, has been challenged and needs validation.

This updated 2016 practice parameter was commissioned by the JTF to capture recent advances in the field of EIB, as elucidated in the most recent literature.

The chair of this workgroup, Dr John Weiler, convened workgroup members who are recognized as experts in the field of EIB. The members have been reviewed for conflicts of interest by the JTF, and conflicts of interest have been listed by the JTF on the JTF Web site at <http://www.allergyparameters.org>.

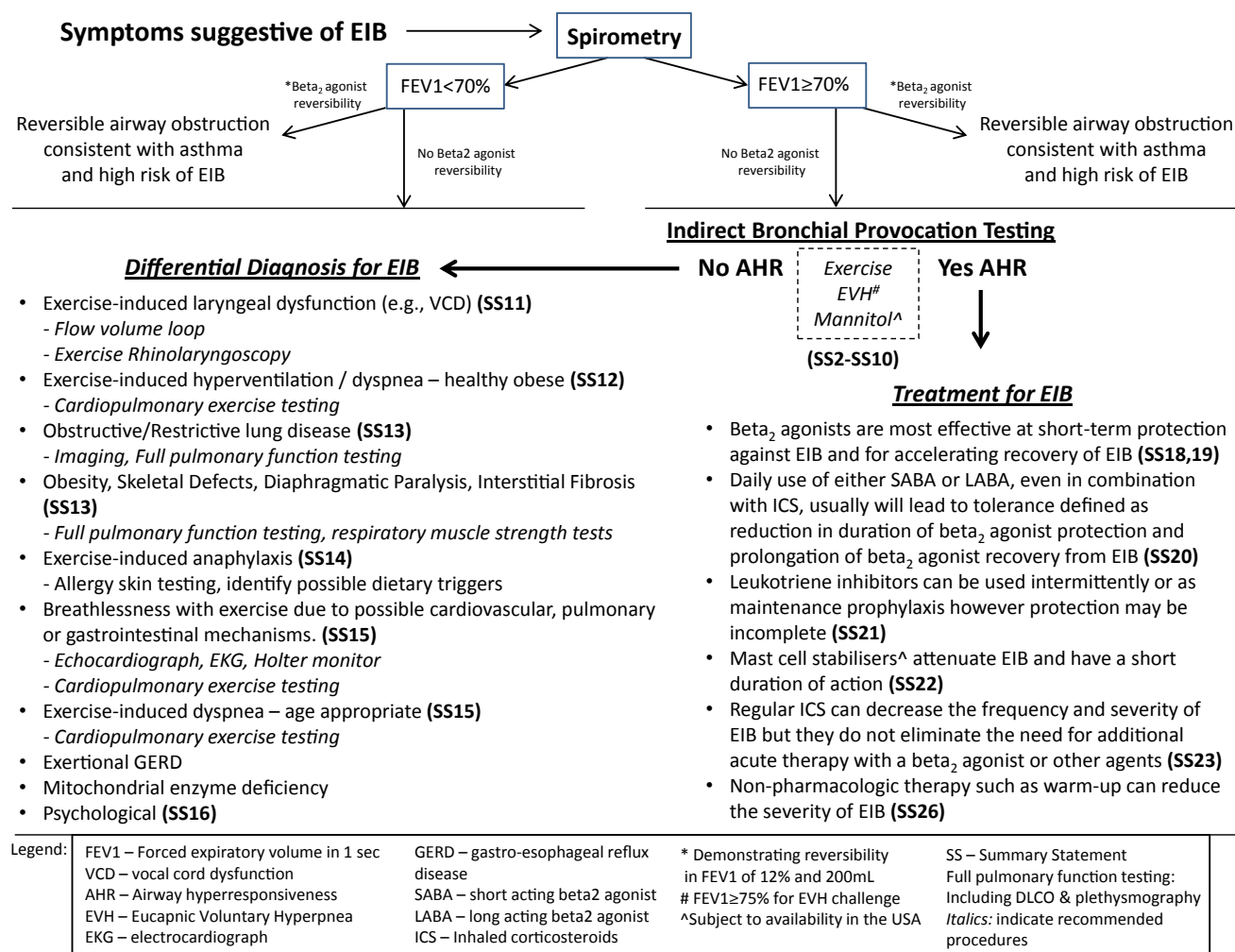
During the development of this practice parameter, at the request of the JTF, the workgroup also recruited a patient advocate to provide a dimension from the patient’s perspective.

The workgroup was asked to update contemporary practice guidelines based on a current systematic literature review. The workgroup obtained supplementary literature, and consensus expert opinions were used when published literature was insufficient.

A search of the medical literature on PubMed was conducted, and all reference categories were included. Search terms included pathogenesis, diagnosis, differential diagnosis, and therapy (both pharmaceutical and nonpharmaceutical) of exercise-induced bronchoconstriction, or exercise-induced asthma (which is no longer a preferred term); asthma; and exercise and asthma.

References assessed as relevant to the topic were evaluated to search for other relevant references. Published clinical studies were appraised by category of evidence and used to document the strength of the recommendation (see category of evidence and strength of recommendation ratings). The parameter was then evaluated by JTF reviewers and then by reviewers assigned by the AAAAI and ACAAI, as well as the general memberships of the AAAAI and ACAAI. Based on this process, the parameter can be characterized as an evidence- and consensus-based document.

The pathophysiology of EIB has been elucidated in the last 2 decades. Strenuous exercise is known to create a hyperosmolar environment by introducing dry air in the airway with compensatory water loss, leading to transient osmotic change on the airway surface. The hyperosmolar environment leads to mast cell degranulation with release of mediators, predominately leukotrienes, but also including histamine, tryptase, and prostaglandins. In addition, eosinophils can also be activated, producing further mediators, including leukotrienes. In turn, this might lead to bronchoconstriction and inflammation of the airway, as well as stimulation of sensory nerves, with neurokinin release stimulating release of the gel-forming mucin MUC5AC. The water content of the inspired air, the level of ventilation achieved and maintained during exercise, or both are the major determinants of EIB. The major trigger for bronchoconstriction in



**FIG 1.** Algorithm for the diagnosis, treatment, and differential diagnosis of EIB.

a vulnerable subject is either the loss of water during periods of high ventilation or the addition of an osmotically active agent. Alterations in airway temperature develop during exercise, but thermal factors are thought to have only a minor effect on the amount of bronchoconstriction that occurs.

Exercise itself is not needed to cause bronchoconstriction, just the creation of a hyperosmolar environment. Diagnosis of EIB is made by using exercise or hyperosmolar surrogate challenges, such as EVH or mannitol. If pulmonary function test (PFT) results are normal, then exercise challenge or surrogate hyperosmolar challenge, such as with mannitol or EVH, should be performed.

Management of EIB is based on the understanding that EIB susceptibility varies widely among asthmatic patients, as well as those who do not have other features of asthma. Therefore EIB can occur in the presence or absence of asthma. Vulnerable subjects have characteristics of both airway inflammation with infiltration of the airways by mast cells and eosinophils and airway smooth muscle with hyperresponsiveness. These observations indicate that treatment should be based on the awareness that exercise causes release of mediators, including predominantly leukotrienes but also tryptase, prostaglandins, and histamine, to act on smooth muscle, leading to bronchoconstriction after exercise.

Therefore therapeutic interventions include short-acting  $\beta_2$ -agonists to provide bronchodilation and bronchoprotection. Additionally, anti-inflammatory medications, including inhaled steroids and leukotriene receptor antagonists (LTRAs) or combination therapy (with inhaled corticosteroids and long-acting  $\beta_2$ -agonists [LABAs]), are recommended for inflammation. Combination therapy that includes a LABA should not be used in persons with normal or near-normal baseline lung function (ie, FEV<sub>1</sub> >80% of predicted value) because regular use of short-acting  $\beta_2$ -agonists and LABAs can cause tolerance, limiting their ability to provide bronchoprotection and bronchodilation.

Use of face masks might promote humidification and prevent water loss, attenuating EIB.

The prevalence (Summary Statements 1-4) of EIB is poorly defined because there is no gold standard for diagnosis. EIB is frequently documented with asthma and reflects insufficient control of underlying asthma. Elite athletes have a higher prevalence of EIB than seen in the general population, varying with the intensity of exercise and the environment. EIB should be diagnosed by means of objective testing, preferably by using standardized bronchoprovocation challenge, because the prevalence of EIB varies with the type of challenge and climatic

conditions of relative humidity and temperature. It is important to reiterate that there is no firm consensus for a positive response or the conditions under which exercise should be performed.

Diagnosis (Summary Statements 5-10) of EIB relies on performing a standardized bronchoprovocation challenge in a subject who has been shown to have normal to near-normal PFT results both before and after bronchodilator (Fig 1). Self-reported symptoms and therapeutic trials without a diagnosis are not diagnostic. In a subject who has no history of current clinical asthma, normal PFT results, and no response to bronchodilator, an exercise challenge with a treadmill or cycle or in the sport venue or a surrogate challenge, such as EVH, can be indicated. With exercise challenge, the patient should achieve a heart rate at least 85% of maximum value (95% in children) for 6 minutes after 2 to 4 minutes of ramping up.

If EIB is to be investigated in a patient with known asthma, a graded challenge with inhaled mannitol, if available, might be preferable for reasons of safety to diagnose EIB. If there is no response to a graded challenge and EIB is still suspected, then consider an ungraded challenge.

Differential diagnosis (Summary Statements 11-16) of EIB requires distinguishing inspiratory stridor alone from inspiratory stridor with or without expiratory wheezing. This is essential to differentiate EIB from exercise-induced laryngeal dysfunction. Diagnosis requires performance of an appropriate exercise challenge, direct or indirect surrogate challenge, and flexible laryngoscopy. Providers should determine whether exercise-induced dyspnea and hyperventilation are masquerading as asthma. Furthermore, it is essential to perform spirometry and a focused detailed physical examination if shortness of breath with exercise is associated with underlying conditions, such as chronic obstructive pulmonary disease (COPD) or restrictive lung conditions. Providers should differentiate between exercise-induced anaphylaxis and EIB based on history of shortness of breath accompanied by pruritus, urticaria, and low blood pressure. Appropriate cardiopulmonary testing and referral to an appropriate specialist might be required when breathlessness with exercise with or without chest pain is caused by these mechanisms in the absence of EIB. A psychological evaluation can also be performed when history is suggestive of a psychiatric disorder (Fig 1).

Therapy (Summary Statements 17-28) for EIB requires re-evaluation of patients with frequent EIB, which suggests poor asthma control, and those who do not have appropriate management. Providers should recognize that there is inpatient and outpatient variability in the effectiveness of pharmacotherapeutic agents on an individual basis. Patients should be scheduled to have regular follow-up of their therapy to determine

the effectiveness of the medication. Medications can differ in effectiveness over time because of the variability of asthma, environmental conditions, intensity of exercise, and tolerance to  $\beta_2$ -agonists, as well as patient compliance (Fig 1).

Inhaled  $\beta_2$ -agonist monotherapy should be used only for short-term prophylaxis against EIB. Providers should only use a single dose of short-acting  $\beta_2$ -agonist (SABA) and/or LABA on an intermittent basis because this might protect against or attenuate EIB. SABAs are effective for 2 to 4 hours and LABAs for up to 12 hours. Caution is recommended in daily use of  $\beta_2$ -agonists alone or in combination with inhaled corticosteroids (ICSs) because this can lead to tolerance. Tolerance can manifest as a reduction in duration and magnitude of protection against EIB and a prolongation of recovery in response to SABAs after exercise.

Leukotriene modifiers can be used daily or intermittently to prevent EIB and do not lead to tolerance. However, they can provide incomplete protection and cannot reverse existing airway obstruction. Mast cell stabilizers, such as cromolyn and nedocromil, can be given shortly before exercise to attenuate EIB but have a short duration of action either alone or as added therapy with other drugs for EIB. These agents are not currently available in the United States.

ICSs taken alone or in combination with other therapies can decrease the frequency and severity of EIB. However, ICSs do not eliminate EIB in all subjects, and ICS therapy might not prevent occurrence of tolerance from daily LABA therapy.

Anticholinergic agents provide inconsistent results in attenuating EIB. Methylxanthines and antihistamines should be used cautiously or selectively because they have inconsistent results.

Nonpharmacologic therapy is recommended by using pre-exercise warm-up to prevent EIB and partially reduce the severity of EIB. Dietary supplementation with fish oil (ie, omega-3 fatty acids) and ascorbic acid and measures to reduce sodium intake are inconclusive in reducing the severity of EIB.

Competitive and elite athletes can have EIB alone, which might have different characteristics to those seen in patients with EIB with asthma in relation to pathogenesis, presentation, diagnosis, management, and requirements by governing bodies for permission to use pharmaceutical agents. However, recent studies indicate that both recreational and elite athletes with EIB with asthma can be treated in a similar manner. EIB alone, without underlying asthma, although not extensively studied in athletes, responds to similar treatment as with asthma. The presence of EIB reflects active asthma. Good control of EIB can be attained with the management discussed above, leading to a healthy lifestyle, including regular exercise and pursuit of the chosen sport.



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**Chief Editors:** John M. Weiler, MD, John D. Brannan, PhD, and Christopher C. Randolph, MD

**Workgroup Contributors:** John D. Brannan, PhD, Teal S. Hallstrand, MD, Jonathan Parsons, MD, William Silvers, MD, William Storms, MD, and Joanna Zeiger, MS, PhD

**Task Force Reviewers:** David I. Bernstein, MD, Joann Blessing-Moore, MD, Matthew Greenhawt, MD, MBA, MSc, David Khan, MD, David Lang, MD, Richard A. Nicklas, MD, John Oppenheimer, MD, Jay M. Portnoy, MD, Christopher R. Randolph, MD, Diane E. Schuller, MD, Stephen A. Tilles, MD, and Dana Wallace, MD

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## TABLE OF CONTENTS

- I. Classification and recommendation of evidence
- II. Glossary
- III. Preface

- IV. Executive summary
- V. Summary statements
- VI. Pathophysiology
- VII. Prevalence
- VIII. Diagnosis
- IX. Differential diagnosis
- X. Therapy

## CONTRIBUTORS

The JTF has made a concerted effort to acknowledge all contributors to this parameter. If any contributors have been excluded inadvertently, the task force will ensure that appropriate recognition of such contributions is made subsequently.

## CHIEF EDITORS

### John M. Weiler, MD, MBA

Division of Immunology  
Department of Medicine  
Carver College of Medicine  
University of Iowa  
Iowa City, Iowa  
CompleWare Corporation  
Iowa City, Iowa

### John D. Brannan, PhD

Department of Respiratory and Sleep Medicine  
John Hunter Hospital  
New Lambton, Australia

### Christopher Randolph, MD

Pediatrics, Allergy, Immunology Division  
Center for Allergy, Asthma and Immunology  
Yale University Hospital  
Regional Hospitals  
Waterbury, Connecticut

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## WORKGROUP CONTRIBUTORS

### John D. Brannan, PhD

Department of Respiratory and Sleep Medicine  
John Hunter Hospital  
New Lambton, Australia

### Teal S. Hallstrand, MD, MPH

Division of Pulmonary & Critical Care  
Center for Lung Biology  
University of Washington  
Seattle, Washington

### Jonathan Parsons, MD, MSc, FCCP

Ohio State University Asthma Center  
OSU Multidisciplinary Cough Program  
Ohio State University Wexner Medical Center  
Division of Pulmonary, Allergy, Critical Care &  
Sleep Medicine  
Columbus, Ohio

**William Silvers, MD**

Division of Allergy & Clinical Immunology  
University of Colorado Denver School of Medicine  
Denver, Colorado

**William Storms, MD**

University of Colorado Health Sciences Center  
Denver, Colorado

**Joanna Zeiger, MS, PhD**

Patient Advocate  
Denver, Colorado

**TASK FORCE REVIEWERS****David I. Bernstein, MD**

Division of Immunology, Allergy, and Rheumatology  
University of Cincinnati College of Medicine  
Cincinnati, Ohio

**Joann Blessing-Moore, MD**

Consulting Clinical Faculty  
Department of Immunology  
Stanford University Medical Center  
Palo Alto, California

**Matthew Greenhawt, MD, MBA, MSc**

Allergy Section, Children's Hospital Colorado  
University of Colorado Denver School of Medicine  
Aurora, Colo

**David A. Khan, MD**

Division of Allergy & Immunology  
University of Texas Southwestern Medical Center  
Dallas, Texas

**David M. Lang, MD**

Allergy/Immunology Section  
Respiratory Institute  
Cleveland Clinic Foundation  
Cleveland, Ohio

**Richard A. Nicklas, MD**

Department of Medicine  
George Washington Medical Center  
Washington, DC

**John Oppenheimer, MD**

Department of Internal Medicine  
New Jersey Medical School  
Morristown, NJ

**Jay M. Portnoy, MD**

Section of Allergy, Asthma & Immunology  
Children's Mercy Hospital  
University of Missouri–Kansas City School of  
Medicine  
Kansas City, Missouri

**Christopher C. Randolph, MD**

Center for Allergy, Asthma and Immunology  
Yale Hospital

Waterbury, Connecticut

**Diane E. Schuller, MD**

Chief of Allergy and Immunology  
Pennsylvania State University  
Hershey Medical College  
Hershey, Pennsylvania

**Stephen A. Tilles, MD**

University of Washington School of Medicine  
Seattle, Washington

**Dana Wallace, MD**

Department of Medicine  
Nova Southeastern University  
Davie, Florida

**SUMMARY OF CONFLICT OF INTEREST  
DISCLOSURES**

The following is a summary of interests disclosed on workgroup members' conflict of interest disclosure statements (not including information concerning family member interests). Completed conflict of interest disclosure statements are available on request.

Workgroup member	Disclosures
John Weiler, MD	<i>Employment and stockholder:</i> CompleWare Corporation <i>Stockholder:</i> Iowa Clinical Research Corporation
Christopher Randolph, MD	<i>Consultant:</i> GlaxoSmithKline, Astra, TEVA, and Sanofi <i>Advisory boards:</i> Astra, TEVA, and Sanofi <i>Speaker:</i> GlaxoSmithKline, Astra, TEVA, Sanofi, and Shire <i>Grants:</i> GlaxoSmithKline, Astra, Amgen, Genentech, and Merck
John D. Brannan, PhD	Receives a 10% share of royalties for Aridol/Osmohale given to Royal Prince Alfred Hospital, Sydney, Australia provided by Pharmaxis, Australia <i>Stockholder:</i> Pharmaxis At the time of writing, Aridol was not available in the United States but is US Food and Drug Administration approved
Teal Hallstrand, MD	No conflicts
Jonathan Parsons, MD	No conflicts
William Silvers, MD	No conflicts
William Storms, MD	<i>Grants:</i> Amgen, Genentech/Novartis, GlaxoSmithKline, Circassia, Meda, Mylan, Sanofi, Sunovion, and TEVA <i>Consultant:</i> Amgen, Astra, Bosch & Lomb, Merck, Sunovion, and TEVA <i>Speaker:</i> Astra, Genentech, Merck, Sanofi, and TEVA
Joanna Zeiger	No conflicts

**Resolution of potential conflicts of interest**

The JTF recognizes that experts in a field are likely to have interests that could come into conflict with development of a completely unbiased and objective practice parameter. A process

has been developed to prevent potential conflicts from influencing the final document in a negative way to take advantage of that expertise.

At the workgroup level, members who have a potential conflict of interest either do not participate in discussions concerning topics related to the potential conflict, or if they do write a section on that topic, the workgroup completely rewrites it without their involvement to remove potential bias. In addition, the entire document is reviewed by the JTF, and any apparent bias is removed at that level. Finally, the practice parameter is sent for review both by invited reviewers and by anyone with an interest in the topic by posting the document on the Web sites of the ACAAI and the AAAAI.

Statement	Definition	Implication
Strong recommendation (StrRec)	A strong recommendation means the benefits of the recommended approach clearly exceed the harms (or that the harms clearly exceed the benefits in the case of a strong negative recommendation) and that the quality of the supporting evidence is excellent (Grade A or B). <sup>*</sup> In some clearly identified circumstances, strong recommendations might be made based on lesser evidence when high-quality evidence is impossible to obtain and the anticipated benefits strongly outweigh the harms.	Clinicians should follow a strong recommendation unless a clear and compelling rationale for an alternative approach is present.
Moderate (Mod)	A recommendation means the benefits exceed the harms (or that the harms exceed the benefits in the case of a negative recommendation), but the quality of evidence is not as strong (grade B or C). <sup>*</sup> In some clearly identified circumstances, recommendations might be made based on lesser evidence when high-quality evidence is impossible to obtain and the anticipated benefits outweigh the harms.	Clinicians should also generally follow a recommendation but should remain alert to new information and sensitive to patient preferences.
Weak (Weak)	An option means that either the quality of evidence that exists is suspect (grade D) <sup>*</sup> or that well-done studies (grade A, B, or C) <sup>*</sup> show little clear advantage to one approach versus another.	Clinicians should be flexible in their decision making regarding appropriate practice, although they might set bounds on alternatives; patient preference should have a substantial influencing role.

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Statement	Definition	Implication
No recommendation (NoRec)	No recommendation means there is both a lack of pertinent evidence (grade D) <sup>*</sup> and an unclear balance between benefits and harms.	Clinicians should feel little constraint in their decision making and be alert to new published evidence that clarifies the balance of benefit versus harm; patient preference should have a substantial influencing role.

## I. CLASSIFICATION OF RECOMMENDATIONS AND EVIDENCE

### Recommendation rating scale

#### Category of evidence

- Ia Evidence from meta-analysis of randomized controlled trials
- Ib Evidence from at least 1 randomized controlled trial
- IIa Evidence from at least 1 controlled study without randomization
- IIb Evidence from at least 1 other type of quasiexperimental study
- III Evidence from nonexperimental descriptive studies, such as comparative studies
- IV Evidence from expert committee reports or opinions, clinical experience of respected authorities, or both

#### Strength of recommendation<sup>\*</sup>

- A Directly based on category I evidence
- B Directly based on category II evidence or extrapolated recommendation from category I evidence
- C Directly based on category III evidence or extrapolated recommendation from category I or II evidence
- D Directly based on category IV evidence or extrapolated recommendation from category I, II, or III evidence
- LB Laboratory based
- NR Not rated

## HOW THIS PRACTICE PARAMETER WAS DEVELOPED

### The Joint Taskforce on Practice Parameters

The JTFPP is a 12-member task force consisting of representatives assigned by the AAAAI and the ACAAI. This task force oversees the development of practice parameters; selects the workgroup chair or chairs; and reviews drafts of the parameters for accuracy, practicality, clarity, and broad utility of the recommendations for clinical practice.

### The Exercise-induced Bronchoconstriction Workgroup

The Exercise-induced Bronchoconstriction Practice Parameter Workgroup was commissioned by the JTF to develop and update a practice parameters that address the pathogenesis, diagnosis and differential diagnosis, epidemiology, management, and treatment, both pharmaceutical and nonpharmaceutical, of EIB.



The chair (John Weiler, MD) invited workgroup members who are considered experts in the field of EIB to participate in the parameter update development. Workgroup members have been vetted for financial conflicts of interest by the JTF, and their conflicts of interest have been listed in this document and are posted on the JTF Web site at <http://www.allergyparameters.org>. Where a potential conflict of interest is present, the potentially conflicted workgroup member was excluded from discussing relevant issues.

The charge to the workgroup was to use a systematic literature review in conjunction with consensus expert opinion and workgroup-identified supplementary documents to develop practice parameters that provide a comprehensive approach for understanding the pathogenesis, diagnosis and differential diagnosis, epidemiology, management, and treatment, both pharmaceutical and nonpharmaceutical, of EIB.

The authors note that this document contains an update of the earlier practice parameter<sup>E1</sup> and other documents published by various entities (the American Thoracic Society [ATS], AAAAI, GALEN, National Athletic Trainers Association, and Agency for Healthcare Research & Quality).

### Protocol for finding evidence

Search terms and programs encompassed PubMed review or meta-analysis (2010-2014) for exercise-induced asthma (60 references), exercise-induced bronchospasm (93 references), exercised-induced bronchoconstriction (55 references), clinical or randomized controlled trial exercise-induced asthma (114 references), exercise-induced bronchospasm (174 references), and exercise-induced bronchoconstriction (99 references) in the last 5 years; National Institute for Health and Care Excellence (NICE) evidence search for exercise-induced asthma systematic review in the last 3 years (76 references), exercise-induced bronchoconstriction (36 references), and exercise induced-bronchospasm (26 references) in the last 3 years; NICE primary research for exercise-induced bronchoconstriction (3 references) and exercise-induced asthma (10 references); the Trip Database for exercise from 2010 systematic review for exercise-induced asthma (7 references), exercise-induced bronchoconstriction (13 references), exercise-induced bronchospasm (20 references), controlled clinical trial exercise-induced bronchospasm (11 references), exercise-induced bronchoconstriction (56 references), and exercise-induced asthma (77 references); the Health Services/Technology Assessment Texts (HSTAT) collection for exercise-induced asthma (59 references); [clinicaltrials.gov](http://clinicaltrials.gov) for exercise-induced asthma (78 references), exercise-induced bronchoconstriction (35 references), and exercise-induced bronchospasm (19 references); Cumulative Index of Nursing and Allied Health Literature (CINAHL) meta-analysis systematic review (2010) for exercise-induced bronchoconstriction and exercise bronchospasm (21 references), randomized control or clinical trial (77 references); Exerpta Medica database (EMBASE) for exercise bronchospasm (107 references), exercise-induced bronchoconstriction (232 references), exercise or exercise and induced asthma 2010-2014 (813 references), same search terms without year limit 4543 references, clinical or randomized control trial (38 references), clinical trial and controlled study (190 references); Sport Discus for exercise-induced asthma, exercise-induced asthma, bronchospasm, exercise-induced bronchoconstriction 2010 on (76 references); and [ahrq.gov](http://ahrq.gov) for exercise-induced bronchoconstriction,

exercise-induced asthma, and exercise-induced bronchospasm (1 reference).

References identified as being relevant were evaluated and searched for additional references, which were searched also for citable references. In addition, members of the workgroup were asked to identify references of which they were aware that were missed by this initial search.

## II. GLOSSARY

*Exercise-induced bronchoconstriction* (EIB) is defined as a transient narrowing of the lower airway after exercise in the presence or absence of clinically recognized asthma. The term *exercise-induced asthma* (EIA) is not used in this document because it might imply incorrectly that exercise causes rather than exacerbates or triggers an asthma attack.

*Bronchial hyperresponsiveness* (BHR) or *airway hyperresponsiveness* is an increase in sensitivity to an agent and is expressed as the dose or concentration of a substance that produces a specific decrease in FEV<sub>1</sub> (eg, PD<sub>20</sub> or PC<sub>20</sub>, respectively).

*Bronchial reactivity* is the rate of change in FEV<sub>1</sub> in relation to the dose or stimulus (eg, response/dose ratio with mannitol is the percentage decrease divided by the dose that achieves that decrease or the percentage decrease in exercise in response to the optimal stimulus).

*Competitive athletes* are persons who engage in strenuous aerobic activity at any level from grade school age and older.

*Conditioning* is defined as preparing the body for physical exercise and, in particular, sports performance. It is also a term used in relation to the heat and humidity conditions of the inspired air whereby water and heat are transferred to the air so that they match lower airway conditions.

*Direct challenges* are those in which a single pharmacologic agent, such as methacholine or histamine, is the provoking substance administered exogenously that acts directly through receptors on airway smooth muscle to cause contraction.

*Elite athletes* are highly competitive persons who train and compete consistently at higher levels (eg, Olympics or professional aerobic sports).

*Eucapnic voluntary hyperpnea* (EVH) describes a type of indirect challenge in which a subject inhales a eucapnic gas mixture (5% CO<sub>2</sub>, 21% O<sub>2</sub>, and balance N<sub>2</sub>) for about 6 minutes and then performs spirometry.

*Graded challenge* is a challenge test in which an agent is administered by means of inhalation at increasing doses or concentration to cause a decrease in FEV<sub>1</sub>. This permits the construction of a dose-response curve to determine the degree of airway sensitivity and is expressed as a provoking dose or provoking concentration.

*Indirect challenges* are those in which exercise or a surrogate, such as EVH; inhaled osmotic agents, such as mannitol or hypertonic saline; or inhalation of AMP is the provoking agent that in turn triggers endogenous mediator release that acts to cause airway smooth muscle contraction.

*Tolerance* is a decrease in the degree, duration, or both of response to an agent when used continuously instead of intermittently. Tolerance ordinarily refers to inhibition of bronchoconstriction and in some cases bronchodilation to  $\beta_2$ -adrenergic agents.

*Ungraded challenges* are challenges in which a single episode of hyperpnea (dose) is administered to cause a specific decrease in

FEV<sub>1</sub>. FEV<sub>1</sub> is measured regularly over more than 1 time point at the conclusion of the challenge. No dose-response curve can be constructed, but the severity of the response is based on the decrease in FEV<sub>1</sub> (eg, laboratory or field exercise and EVH).

*Exercise challenge* in the assessment of asthma requires effort that is ramped up rapidly so that maximum heart rate should be achieved within 2 to 3 minutes. This should be considered for both laboratory-based exercise challenge tests with a treadmill or cycle ergometer and when performing field-based exercise challenge. This exercise protocol is more intensive in the initial stages of exercise than a cardiopulmonary exercise test, and it is for this reason that cardiopulmonary exercise testing is not recommended for investigating EIB.

### III. PREFACE

The first practice parameter on EIB was published in 2010. This update is required by the National Clearinghouse and JTF consistent with the requirement of an update every 5 years.

In the ensuing years since the first publication of the EIB practice parameter, there has been increased understanding of the pathogenesis of EIB and improved diagnosis of this disorder by means of objective pulmonary function testing. At the time of this publication, dry powder mannitol for inhalation is not currently available in the United States but is available in many other countries.

If baseline pulmonary function test results are normal to near normal (before and after bronchodilator) in a person with suspected EIB, then further testing should be performed by using a standardized exercise challenge or EVH.

Since 2010, the efficacy of nonpharmaceutical interventions, such as omega-3 fatty acids, has been challenged and needs validation.

This updated 2016 practice parameter was commissioned by the JTF to capture recent advances in the field of EIB, as elucidated in the most recent literature.

The chair of this workgroup, Dr John Weiler, convened workgroup members who are recognized as experts in the field of EIB. The members have been reviewed for conflicts of interest by the JTF, and conflicts of interest have been listed by the JTF on the JTF Web site at <http://www.allergyparameters.org>.

During the development of this practice parameter, at the request of the JTF, the workgroup also recruited a patient advocate to provide a dimension from the patient's perspective.

The workgroup was asked to update contemporary practice guidelines based on a current systematic literature review. The workgroup obtained supplementary literature, and consensus expert opinions were used when published literature was insufficient.

A search of the medical literature on PubMed was conducted, and all reference categories were included. Search terms included pathogenesis, diagnosis, differential diagnosis, and therapy (both pharmaceutical and nonpharmaceutical) of exercise-induced bronchoconstriction or exercise-induced asthma (which is no longer a preferred term); asthma; and exercise and asthma.

References assessed as relevant to the topic were evaluated to search for other relevant references. Published clinical studies were appraised by category of evidence and used to document the strength of the recommendation (see the Category of evidence and Strength of recommendation ratings sections). The parameter was then evaluated by JTF reviewers and then by reviewers assigned by the AAAAI and ACAAI, as well as the general

memberships of the AAAAI and ACAAI. Based on this process, the parameter can be characterized as an evidence- and consensus-based document.

### IV. EXECUTIVE SUMMARY

The pathophysiology of EIB has been elucidated in the last 2 decades. Strenuous exercise is known to create a hyperosmolar environment by introducing dry air into the airway with compensatory water loss, leading to transient osmotic change on the airway surface. The hyperosmolar environment leads to mast cell degranulation with release of mediators, predominately leukotrienes, but also including histamine, tryptase, and prostaglandins. In addition, eosinophils can also be activated, producing further mediators, including leukotrienes. In turn, this can lead to bronchoconstriction and inflammation of the airway, as well as stimulation of sensory nerves with neurokinin release, stimulating the release of the gel-forming mucin MUC5AC. The water content of the inspired air, the level achieved and maintained during exercise, or both are the major determinants of EIB in subjects. The major trigger for bronchoconstriction in a vulnerable subject is either water loss during periods of high ventilation or the addition of an osmotically active agent. Alterations in airway temperature develop during exercise, but thermal factors are thought to have only a minor effect on the amount of bronchoconstriction that occurs.

Exercise itself is not needed to cause bronchoconstriction, just the creation of a hyperosmolar environment. Diagnosis of EIB is made by using exercise or hyperosmolar surrogate challenges, such as EVH or mannitol. If pulmonary function test (PFT) results are normal, then exercise challenge or surrogate hyperosmolar challenge, such as with mannitol or EVH, should be performed.

Management of EIB is based on the understanding that EIB susceptibility varies widely among asthmatic patients, as well as those who do not have other features of asthma. Therefore EIB can occur in the presence or absence of asthma. Vulnerable subjects have characteristics of both airway inflammation with infiltration of the airways by mast cells and eosinophils and airway smooth muscle with hyperresponsiveness. These observations indicate that treatment should be based on the awareness that exercise causes release of mediators, including predominantly leukotrienes, but also tryptase, prostaglandins, and histamine, to act on smooth muscle, leading to bronchoconstriction after exercise.

Therefore therapeutic interventions include short-acting  $\beta_2$ -agonists (SABAs) to provide bronchodilation and broncho-protection. Additionally, anti-inflammatory medications, including inhaled corticosteroids (ICSs) and leukotriene receptor antagonists (LTRAs), or combination therapy (with ICSs and long-acting  $\beta_2$ -agonists [LABAs]) are recommended for inflammation. Combination therapy that includes a LABA should not be used in persons with normal or near-normal baseline lung function (ie, FEV<sub>1</sub> >80% of predicted value) because regular use of SABAs and LABAs can cause tolerance, limiting their ability to provide bronchoprotection and bronchodilation.

Use of face masks can promote humidification and prevent water loss, attenuating EIB.

The prevalence (Summary Statements 1-4) of EIB is poorly defined because there is no gold standard for diagnosis. EIB is frequently documented with asthma and reflects insufficient control of underlying asthma. Elite athletes have a higher prevalence of EIB than seen in the general population, varying

with the intensity of exercise and the environment. EIB should be diagnosed by means of objective testing, preferably standardized bronchoprovocation challenge, because the prevalence of EIB varies with type of challenge and climatic conditions of relative humidity and temperature. It is important to reiterate that there is no firm consensus for a positive response or the conditions under which exercise should be performed.

Diagnosis (Summary Statements 5-10) of EIB relies on performing a standardized bronchoprovocation challenge in a subject who has been shown to have normal to near-normal PFT results both before and after bronchodilator (Fig E1). Self-reported symptoms and therapeutic trials without a diagnosis are not diagnostic. In a subject who has no history of current clinical asthma, normal PFT results, and no response to bronchodilator, exercise challenge with a treadmill or cycle or in the sport venue or a surrogate challenge, such as EVH, might be indicated. With exercise challenge, the patient should achieve a heart rate at least 85% of maximum (95% in children) for 6 minutes after 2 to 4 minutes of ramping up.

If EIB is to be investigated in a patient with known asthma, a graded challenge with inhaled mannitol, if available, might be preferable for reasons of safety to diagnose EIB. If there is no response to a graded challenge and EIB is still suspected, then consider an ungraded challenge.

Differential diagnosis (Summary Statements 11-16) of EIB requires distinguishing inspiratory stridor alone from inspiratory stridor with or without expiratory wheezing. This is essential to differentiate EIB from exercise-induced laryngeal dysfunction. Diagnosis requires performance of appropriate exercise challenge, direct or indirect surrogate challenge, and flexible laryngoscopy. Providers should determine whether exercise-induced dyspnea and hyperventilation are masquerading as asthma. Furthermore, it is essential to perform spirometry and a focused detailed physical examination if shortness of breath with exercise is associated with underlying conditions, such as chronic obstructive pulmonary disease (COPD) or a restrictive lung condition. Providers should differentiate between exercise-induced anaphylaxis and EIB based on a history of shortness of breath accompanied by pruritus, urticaria, and low blood pressure. Appropriate cardiopulmonary exercise testing and referral to an appropriate specialist might be required when breathlessness with exercise with or without chest pain is caused by these mechanisms in the absence of EIB. A psychological evaluation can also be performed when history is suggestive of psychiatric disorder (Fig E1).

Therapy (Summary Statements 17-28) for EIB requires reevaluation of patients with frequent EIB, which suggests poor asthma control, and those in whom appropriate management fails. Providers should recognize that there is inpatient and interpatient variability in the effectiveness of pharmacotherapeutic agents on an individual basis. Patients should be scheduled to have regular follow-up of their therapy to determine the effectiveness of the medication. Medications can differ in effectiveness over time because of variability of asthma, environmental conditions, intensity of exercise, and tolerance to  $\beta_2$ -agonists, as well as patient compliance (Fig E1).

Inhaled  $\beta_2$ -agonist monotherapy should be used only for short-term prophylaxis against EIB. Providers should only use a single dose of SABA, LABA, or both on an intermittent basis because this can protect against or attenuate EIB. SABAs are effective for 2 to 4 hours and LABAs for up to 12 hours. Caution is

recommended in daily use of  $\beta_2$ -agonists alone or in combination with ICSs because this can lead to tolerance. Tolerance can be manifested as a reduction in the duration and magnitude of protection against EIB and a prolongation of recovery in response to SABAs after exercise.

Leukotriene modifiers can be used daily or intermittently to prevent EIB and do not lead to tolerance. However, they can provide incomplete protection and cannot reverse existing airway obstruction. Mast cell stabilizers, such as cromolyn and nedocromil, can be administered shortly before exercise to attenuate EIB but have a short duration of action either alone or as added therapy with other drugs for EIB. These agents are not currently available in the United States.

ICSs taken alone or in combination with other therapies can decrease the frequency and severity of EIB. However, ICSs do not eliminate EIB in all subjects, and ICS therapy might not prevent occurrence of tolerance from daily LABA therapy.

Anticholinergic agents provide inconsistent results in attenuating EIB. Methylxanthines and antihistamines should be used cautiously or selectively because they have inconsistent results.

Nonpharmacologic therapy is recommended by using pre-exercise warm-up to prevent EIB and partially reduce the severity of EIB. Dietary supplementation with fish oil (ie, omega-3 fatty acids) and ascorbic acid and measures to reduce sodium intake are inconclusive in reducing the severity of EIB.

Competitive and elite athletes might have EIB alone, which can have different characteristics than are seen in patients with EIB with asthma in relation to pathogenesis, presentation, diagnosis, management, and requirements by governing bodies for permission to use pharmaceutical agents. However, recent studies indicate that both recreational and elite athletes with EIB with asthma can be treated in a similar manner. EIB alone, without underlying asthma, although not extensively studied in athletes, responds to similar treatment as with asthma. The presence of EIB reflects active asthma. Good control of EIB can be attained with the management discussed above, leading to a healthy lifestyle, including regular exercise and pursuit of the chosen sport.

## V. SUMMARY STATEMENTS

*Summary Statement 1:* In asthmatic patients EIB can indicate lack of control of the underlying asthma. Therefore treat the uncontrolled asthma to get control of EIB. [Strength of Recommendation: Strong; Evidence: D]

*Summary Statement 2:* A diagnosis of EIB should be confirmed by demonstration of airways reversibility or challenge in association with a history consistent with EIB because self-reported symptoms are not adequate. [Strength of Recommendation: Strong; Evidence: B]

*Summary Statement 3:* Evaluate EIB in elite athletes by using objective testing. [Strength of Recommendation: Strong; Evidence: B]

*Summary Statement 4:* Perform a standardized bronchoprovocation (exercise or a surrogate) challenge to diagnose EIB because the prevalence of EIB will vary with the type of challenge and the conditions under which the challenge is performed. [Strength of Recommendation: Strong; Evidence: A]

*Summary Statement 5:* In subjects with no current clinical history of asthma, use an indirect ungraded challenge (eg, exercise challenge or surrogate testing, such as with EVH) for assessing EIB in the recreational or elite athlete who has normal lung function. [Strength of Recommendation: Strong; Evidence: D]

*Summary Statement 6:* Use an indirect graded challenge (eg, mannitol, if available) for assessing EIB in recreational or elite athletes who have normal to near-normal lung function and who might currently require treatment for the prevention of EIB or asthma. [Strength of Recommendation: Strong; Evidence: D]

*Summary Statement 7:* Perform an indirect challenge (eg, exercise challenge or surrogate testing, such as with EVH or mannitol, where available) instead of a direct challenge (eg, methacholine) for assessing EIB, recognizing that an indirect challenge is more sensitive for detection of EIB than a direct (eg, methacholine) challenge. [Strength of Recommendation: Strong; Evidence: B]

*Summary Statement 8:* Ensure the ventilation reached and sustained during exercise challenge testing is at least 60% of the maximum voluntary ventilation by using dry medical grade air to achieve an adequate challenge. If ventilation cannot be measured, ensure the heart rate as a percentage of maximum heart rate (HRmax) that is reached and sustained is at least 85% in adults and 95% in children and elite athletes. [Strength of Recommendation: Strong; Evidence: B]

*Summary Statement 9:* Perform EVH as the preferred surrogate challenge for the athlete without a current history of asthma participating in competitive sports in whom the diagnosis of EIB is suspected. [Strength of Recommendation: Strong; Evidence: D]

*Summary Statement 10:* If an indirect graded challenge (eg, mannitol) result is negative and EIB is still suspected, an ungraded challenge should be considered. [Strength of Recommendation: Weak; Evidence: B]

*Summary Statement 11:* To differentiate between EIB and exercise-induced laryngeal dysfunction (EILD), perform appropriate challenge tests (eg, exercise, EVH, and mannitol for EIB) and potentially flexible laryngoscopy during exercise for diagnosis of EILD. [Strength of Recommendation: Strong; Evidence: B]

*Summary Statement 12:* To determine whether exercise-induced dyspnea and hyperventilation are masquerading as asthma, especially in children and adolescents, perform cardiopulmonary exercise testing. [Strength of Recommendation: Moderate; Evidence: C]

*Summary Statement 13:* Perform spirometry, as well as detailed pulmonary examination, to determine whether shortness of breath with exercise is associated with underlying conditions, such as COPD, or restrictive lung conditions, such as obesity, skeletal defects (eg, pectus excavatum), diaphragmatic paralysis, or interstitial fibrosis, rather than EIB. [Strength of Recommendation: Moderate; Evidence: C]

*Summary Statement 14:* Consider a diagnosis of exercise-induced anaphylaxis (EIA<sub>na</sub>) instead of EIB based on a history of shortness of breath or other respiratory tract symptoms accompanied by systemic symptoms (eg, pruritis, urticaria, and hypotension). [Strength of Recommendation: Moderate; Evidence: C]

*Summary Statement 15:* Refer to appropriate specialists (eg, cardiologist or pulmonologist) to perform cardiopulmonary testing when breathlessness with exercise, with or without chest pain, might be caused by heart disease or other conditions in the absence of EIB. [Strength of Recommendation: Moderate; Evidence: C]

*Summary Statement 16:* Refer patients for psychological evaluation when the symptoms (eg, hyperventilation and anxiety disorders) are in the differential diagnosis of EIB. [Strength of Recommendation: Weak; Evidence: D]

*Summary Statement 17:* Schedule regular office visits with patients because medications can differ in effectiveness over time because of variability of asthma, environmental conditions, intensity of the exercise stimulus, and tachyphylaxis. [Strength of Recommendation: Strong; Evidence: A]

### **β<sub>2</sub>-Adrenergic receptor agonists**

*Summary Statement 18:* Prescribe inhaled short-acting β<sub>2</sub>-adrenergic receptor agonists for protection against EIB and for accelerating recovery of pulmonary function when given after a decrease in pulmonary function after exercise. [Strength of Recommendation: Strong; Evidence: A]<sup>E2</sup>

*Summary Statement 19:* Prescribe a single dose of SABA, LABA, or both on an intermittent basis (ie, <4 times per week) before exercise because this might protect against or attenuate EIB. [Strength of Recommendation: Strong; Evidence: A]

*Summary Statement 20:* Be cautious in daily use of β<sub>2</sub>-adrenergic agents alone or in combination with ICSs because this can lead to tolerance manifested as a reduction in duration, magnitude, or both of protection against EIB and a prolongation of recovery in response to SABAs after exercise. [Strength of Recommendation: Strong; Evidence: A]<sup>E2</sup>

### **Leukotriene inhibitors**

*Summary Statement 21:* Consider prescribing daily therapy with leukotriene inhibitors because this does not lead to tolerance and has been shown to attenuate EIB in 50% of patients. It can also be used for intermittent or maintenance prophylaxis; however, it provides incomplete protection and is not effective for reversing airway obstruction. [Strength of Recommendation: Strong; Evidence: A]<sup>E2</sup>

### **Mast cell stabilizers**

*Summary Statement 22:* Consider prescribing inhaled cromolyn sodium and nedocromil sodium (currently not available in the United States as a metered-dose inhaler or dry powder inhaler) shortly before exercise; this attenuates EIB but can have a short duration of action. There is no bronchodilator activity. They might be effective alone or as added therapy with other drugs for EIB. [Strength of Recommendation: Strong; Evidence: A]<sup>E2</sup>

### **ICSs**

*Summary Statement 23:* Consider prescribing ICSs in combination with other therapies because ICSs can decrease the frequency and severity of EIB but not necessarily eliminate it. [Strength of Recommendation: Strong; Evidence: A]<sup>E2</sup>

*Summary Statement 24:* Do not prescribe daily LABAs with ICS therapy to treat EIB unless needed to treat moderate-to-severe persistent asthma. The ICS might not prevent the occurrence of tolerance from daily β<sub>2</sub>-agonist use. [Strength of Recommendation: Strong; Evidence: A]

### **Anticholinergic agents**

*Summary Statement 25:* Consider prescribing inhaled ipratropium bromide for patients who have not responded to other agents; however, its ability to attenuate EIB is inconsistent. [Strength of Recommendation: Weak; Evidence: A]<sup>E2</sup>



## Nonpharmacologic therapy

*Summary Statement 26:* Prescribe pre-exercise warm-up for EIB because it can be helpful in reducing the severity of EIB. [Strength of Recommendation: Strong; Evidence: A]<sup>E2</sup>

*Summary Statement 27:* Consider with caution the recommendation of reduction of sodium intake and ingestion of fish oil and ascorbic acid supplementation; results are questionable in reducing the severity of EIB. [Strength of Recommendation: Weak; Evidence: B]<sup>E2</sup>

## Competitive and elite athletes

*Summary Statement 28:* Treat athletes with EIB alone in a similar manner to those with EIB and asthma by using the recommended general treatments for asthma. This might require additional consideration in athletes in whom some governing bodies might have requirements for obtaining permission to receive pharmaceutical agents for competition. [Strength of Recommendation Strong; Evidence: A]

## VI. PATHOPHYSIOLOGY

### Definition and overview

A period of high ventilation causes respiratory water loss with cooling of the airways and a temporary increase in the osmolarity of the airway surface liquid (ASL) because of a loss of ASL volume. Changes in osmolarity occur only transiently and are rapidly resolved by the movement of water from the osmotically sensitive epithelium into the lumen. The responses of the airways to thermal gradients and the response to water loss and transient hyperosmolarity can act as independent stimuli for the airways to narrow. Cooling is a mechanical stimulus that could induce reactive hyperemia of the bronchial vasculature, whereas the response of the epithelium and other cells to changes in ASL volume and osmolarity is the most likely trigger for mediator release that serves as the primary stimulus for sustained bronchoconstriction.<sup>E3,E4</sup>

When air of subfreezing temperature is inspired during exercise, airway cooling causes vasoconstriction of the bronchial vasculature.<sup>E5</sup> On cessation of exercise, when ventilation decreases and the airways rewarm, a reactive hyperemia with vascular engorgement and edema of the airway wall occurs.<sup>E4</sup> This is known as the thermal theory of EIB. It is generally accepted that the thermal theory is inadequate to explain many of the events that occur in the airways after exercise challenge.<sup>E6,E7</sup> In a canine model ligation of the bronchial circulation did not attenuate hyperpnea-induced bronchoconstriction (HIB), indicating that the bronchial vasculature is not the primary mechanism of bronchoconstriction.<sup>E6</sup> In human subjects inspiring warm air after a challenge test with cold air has a modest effect on the amount of bronchoconstriction experienced 5 to 15 minutes after exercise.<sup>E4</sup>

The osmotic theory of EIB developed as it became apparent that cooling of the airways was not a prerequisite for EIB,<sup>E8</sup> changes in ASL osmolarity could be demonstrated with direct delivery of dry air in the peripheral airways,<sup>E9</sup> and the airways of asthmatic patients were sensitive to inhalation of osmotically active substances.<sup>E10-E13</sup> Dehydration of the ASL causes a transient increase in ion content and osmolarity when water from the ASL is evaporated faster than it is returned by means of condensation or from the epithelium or submucosa.<sup>E14,E15</sup> Dehydration of the ASL also results in reduction in ASL volume.

Such a reduction in ASL volume reduces mucociliary clearance.<sup>E16</sup> These events have been demonstrated *in vivo* by a marked reduction in mucociliary clearance during dry air breathing both in asthmatic patients and healthy subjects.<sup>E17</sup>

The precise mechanism by which water loss and transient osmotic gradients lead to leukocyte activation is not certain. It is well known that mast cells and eosinophils release mediators in response to shifts in osmolarity.<sup>E18-E20</sup> Changes in ASL volume and osmolarity also trigger cellular signaling events in epithelial cells and the release of regulatory proteins from the epithelium that could be involved in leukocyte activation.<sup>E3</sup>

The osmotic and vascular theories of EIB can operate together under conditions of breathing cold dry air when vascular effects that can result in airway edema amplify the contractile effect of mediator release. As the temperature of the inspired air increases toward body temperature, the osmotic effects of water loss are more important than cooling.<sup>E21-E23</sup>

Exercise itself is not necessary to cause bronchoconstriction; voluntary hyperpnea of dry air induces bronchoconstriction similar to exercise in susceptible subjects.<sup>E24,E25</sup> Thus EVH of dry air containing 4.9% to 5% carbon dioxide is often used as a surrogate for exercise in the diagnosis of EIB, particularly in athletes.<sup>E26-E29</sup>

The EVH test for EIB was developed to evaluate military recruits for EIB.<sup>E30</sup> EVH is recommended by the European Respiratory Society/European Academy of Allergy and Clinical Immunology Task Force<sup>E31</sup> to identify EIB in athletes and is included in the World Anti-Doping Agency assessment of asthma ([www.wada-ama.org/Documents/Science\\_Medicine/](http://www.wada-ama.org/Documents/Science_Medicine/)). EVH should not be performed on a patient who has an FEV<sub>1</sub> of less than 70% of predicted value and only with caution if FEV<sub>1</sub> is less than 80% of predicted value because of the risk of a major decrease in FEV<sub>1</sub>.

Most studies indicate that the subjects who are susceptible to EIB have increased cellular inflammation. The fraction of exhaled nitric oxide is increased among asthmatic patients who are susceptible to EIB,<sup>E32</sup> especially atopic subjects.<sup>E33</sup> Inflammatory lipid mediators are implicated in EIB. In particular, concentrations of the cysteinyl leukotrienes (CysLTs) leukotriene (LT) C<sub>4</sub>, LTD<sub>4</sub>, and LTE<sub>4</sub> is increased in induced sputum of adults with EIB<sup>E34</sup> and in exhaled breath condensate (EBC) of children with EIB.<sup>E35</sup> Levels of nonenzymatic products of phospholipid oxidation, 8-isoprostanes, are increased in the EBC of patients who have asthma with EIB. Reduction in the formation of protective lipid mediators in the airways, such as lipoxin A<sub>4</sub>, can also be involved in the pathogenesis of EIB.<sup>E36</sup> Prostaglandin E<sub>2</sub> (PGE<sub>2</sub>) is a key regulatory eicosanoid that inhibits EIB when administered by means of inhalation.<sup>E37</sup> PGE<sub>2</sub> production relative to CysLTs is reduced in patients with EIB.<sup>E38</sup>

The intensity of cellular inflammation in the airways might be an important factor in the susceptibility to EIB because the formation of inflammatory eicosanoids, such as CysLTs and PGD<sub>2</sub>, is largely restricted to myeloid cells.<sup>E39</sup> Several studies have associated the degree of sputum eosinophilia with the severity of EIB, especially in steroid-naïve asthmatic patients.<sup>E38,E40</sup> A reduction in EIB severity after treatment with an ICS is also accompanied by a reduction in the percentage of eosinophils in sputum.<sup>E40</sup>

Increased expression of mast cell genes has been identified in patients with EIB by using genome-wide methods in asthmatic patients based on induced sputum<sup>E38</sup> and, subsequently, epithelial

brushings.<sup>E41</sup> In epithelial brushings the findings of increased expression of tryptase and carboxypeptidase A3 but relatively low chymase expression mirror the findings in patients with T<sub>H</sub>2-high asthma.<sup>E42,E43</sup> The density of intraepithelial mast cells per volume of the airway epithelium measured by using quantitative morphometry in endobronchial tissue of asthmatic patients is markedly increased in subjects who are susceptible to EIB and suggests that mast cell infiltration of the airways is a defining feature of EIB.<sup>E44</sup>

The cellular mechanism leading to leukocyte activation either directly through the movement of water or through a signal from the epithelium in response to this water movement is not known in detail. There is strong evidence that leukocyte-derived eicosanoids, including CysLTs and PGD<sub>2</sub>, are released into the airways after an exercise challenge.<sup>E45-E49</sup> CysLT levels are also increased in exhaled breath condensate after exercise challenge and correlate with the severity of EIB.<sup>E47</sup> CysLTs, such as LTD<sub>4</sub>, mediate airway narrowing at a lower concentration than other mediators, such as PGD<sub>2</sub>, histamine, or methacholine (Fig E2).

Mast cells and eosinophils are the leukocytes most strongly implicated as a source of mediators in patients with EIB.<sup>E44,E48,E49</sup> Mast cells generate PGD<sub>2</sub>, CysLTs, and histamine, whereas eosinophils are also a major source of CysLTs and eosinophilic cationic protein. These leukocyte-derived products are known to mediate airway smooth muscle contraction, sensory nerve activation, and mucus release and increase microvascular permeability (MVP), leading to airway edema.<sup>E50</sup> Although early studies found only small increases in histamine in arterial plasma in response to exercise,<sup>E51,E52</sup> recent studies with induced sputum have confirmed that mast cell degranulation with the release of histamine and tryptase occurs during EIB.<sup>E45,E53,E54</sup> Pharmacologic inhibitors indicate that histamine is responsible for bronchoconstriction early after mannitol challenge, whereas CysLT release is responsible for sustained bronchoconstriction.<sup>E55</sup> After EVH challenge, levels of 9α,11β-PGF<sub>2</sub>, the metabolite of PGD<sub>2</sub>, are increased in urine, and PGD<sub>2</sub> release can be inhibited by pretreatment with a cromone or a high dose of ICS.<sup>E56,E57</sup>

There is strong evidence that CysLTs play a causative role in EIB through pharmacologic studies with CysLT<sub>1</sub> antagonists and 5-lipoxygenase inhibitors.<sup>E58,E59</sup> CysLT<sub>1</sub> receptor antagonists modify the maximum decrease in FEV<sub>1</sub> and the time of recovery after EIB.<sup>E58,E59</sup> The 5-lipoxygenase inhibitor zileuton administered 4 times daily for 2 days reduced the decrease in FEV<sub>1</sub> after exercise challenge by approximately one half.<sup>E60</sup> These results clearly demonstrate a role for CysLTs in the pathogenesis of EIB but also indicate that protection from EIB is incomplete, suggesting that other mediators might play a role. Histamine antagonists have been observed to have inconsistent activity on EIB.<sup>E45,E61-E63</sup>

Many mediators increase MVP,<sup>E64</sup> an event that contributes to bronchoconstriction through leak of proteinaceous material into the airways and edema of the airway wall. An increase in MVP after exercise has been demonstrated by a change in sputum-serum ratio of albumin.<sup>E65</sup> The increase in this ratio has been shown to relate to the severity of EIB. Vascular endothelial growth factor and angiopoietin 2, which alter MVP, have been related to the severity of EIB when measured in the airways after exercise challenge.<sup>E65,E66</sup> An increase in MVP could act to amplify airway narrowing caused by ASM contraction in the context of EIB. There is increasing evidence in athletes of hyperpnea leading to mechanical stress on the airway epithelium. Increases in both

serum and urinary concentrations of pneumoprotein club cell (Clara cell) CC16 levels have been observed after swimming in elite swimmers.<sup>E67,E68</sup> In symptomatic recreational summer athletes increases in CC16 levels were observed in urine after EVH and mannitol,<sup>E69</sup> suggesting it is the osmotic stress that disrupts the epithelium in these subjects in contrast to that observed in elite swimmers.<sup>E67</sup>

The airways contain abundant sensory nerve endings within the epithelium that can be activated directly by changes in osmolarity or in response to other mediators in the airways in a process that involves neurokinin release. Although sensory nerves send signals from the airways to the central nervous system, they can also act locally through a process called retrograde axonal transmission, leading to bronchoconstriction and mucus release. Eicosanoids, such as CysLTs, can either directly activate or alter the activation threshold of sensory nerves.<sup>E70</sup> In dog and guinea pig models of HIB, leukotriene antagonists inhibit both the release of neurokinins and HIB, whereas neurokinin receptor antagonists inhibit the development of HIB without altering neurokinin levels consistent with leukotriene-mediated bronchoconstriction that occurs through sensory nerve activation.<sup>E71,E72</sup> In human subjects the effects of neurokinin 1 antagonists have been mixed in exercise and hypertonic saline models,<sup>E73,E74</sup> possibly because of the predominance of the neurokinin 2 receptor in human subjects that binds to neurokinin A.<sup>E75</sup> In human subjects release of the major gel-forming mucin MUC5AC after exercise challenge is associated with CysLT levels in the airways, and CysLT and neurokinin A levels in the airways are correlated after exercise challenge, supporting a role for sensory nerve activation in this process.<sup>E76</sup>

In approximately half of patients with EIB, there is an interval of refractoriness lasting approximately 1 to 3 hours during which additional exercise produces less bronchoconstriction.<sup>E77-E79</sup> This bronchoprotective effect might be additive to the protective effect of pretreatment with a SABA.<sup>E77</sup> The precise mechanism of the refractory period is not fully understood; however, recent findings with mannitol challenge suggest that the refractory period could be explained by tolerance to the effect of mediator release.<sup>E80,E81</sup> A probable explanation for the refractory period is that it induces the generation of protective prostaglandins because the administration of a nonsteroidal anti-inflammatory drug that inhibits the COX pathway reduces the refractoriness to both exercise and LTD<sub>4</sub> challenge.<sup>E82,E83</sup>

## VII. PREVALENCE

EIB is reported frequently in asthmatic patients but also occurs in the absence of chronic asthma. EIB is very common in athletes. The prevalence of EIB is significantly influenced by the criteria used for diagnosis. The use of self-reported symptoms to make the diagnosis of EIB will result in significant rates of both false-positive and false-negative diagnoses of EIB.<sup>E2,E28,E84,E85</sup> Self-reported symptoms should not be relied on solely for the diagnosis of EIB without the concomitant use of spirometry and bronchoprovocation challenge to confirm the diagnosis.<sup>E2,E28,E84,E86</sup> The prevalence of EIB can be influenced by age, sex, ethnicity/race, and environmental conditions (eg, air temperature and humidity, allergen content, and pollution) in which exercise is performed.<sup>E2,E85,E87,E88</sup>

*Summary Statement 1:* In asthmatic patients EIB can indicate lack of control of the underlying asthma. Therefore treat the

uncontrolled asthma to obtain control of EIB. [Strength of Recommendation: Strong; Evidence: D]

*Summary Statement 2:* A diagnosis of EIB should be confirmed by demonstration of airways reversibility or challenge in association with a history consistent with EIB because self-reported symptoms are not diagnostic. [Strength of Recommendation: Strong; Evidence: B]

The occurrence of bronchoconstriction, especially with symptoms during or after exercise, is one of the common characteristics of asthma, but it also occurs in the absence of other clinical features of asthma. In asthmatic patients EIB in itself is a marker of poor control and suggests the need to initiate or step up therapy.<sup>E89</sup> The prevalence of EIB varies considerably, depending on a multitude of factors, including whether chronic asthma is present, the severity and control of asthma if it is present, environmental conditions, type of testing, and demographics factors.<sup>E2,E89</sup>

EIB is a common disorder in children.<sup>E90,E91</sup> The prevalence of EIB has been reported to be 10% in schoolchildren.<sup>E92</sup> However, estimates of up to 20% have been reported.<sup>E92</sup> Overall, there is a paucity of data from cohort studies in children examining changes in the prevalence of EIB over time and virtually no data in adults.

Researchers<sup>E93</sup> exercised 15,241 children using a 6-minute run and a decrease of 15% for peak flow as an indicator of EIB. In this cohort girls (8.5%) were more likely than boys (6.4%) and those from urban settings (8.9%) were more likely than those from rural environments (7%) to have a positive challenge result. As expected in all populations, symptoms were a poor predictor of positive challenge results.

A study using EVH reported a presumed prevalence of EIB of 19.4% in 212 adults without a history of asthma.<sup>E94</sup> Similar findings were documented in another cohort of 136 recreationally active subjects in whom EIB was documented in 13% of the cohort by using EVH.<sup>E95</sup> Subjects with a family history of asthma might have a higher prevalence of EIB.<sup>E96</sup> EIB is also more frequent in atopic subjects,<sup>E97,E98</sup> including those with allergic rhinitis,<sup>E99</sup> and after viral respiratory tract infections and other respiratory diseases.<sup>E100</sup> It is unclear what the relationship is between the natural history of EIB when not associated with chronic asthma and the subsequent development of chronic asthma.

The frequency and severity of asthma can vary by sex, with male subjects having greater frequency during childhood but female subjects having more severe asthma during adulthood.<sup>E101-102</sup> In contrast to the above findings,<sup>E93</sup> another study<sup>E103</sup> did not demonstrate sex differences in patients with EIB. However, these investigators found that with increasing age, the frequency of EIB decreased.

The frequency of asthma and EIB can also vary by sex in elite athletes. In winter sports female subjects appear to exceed male subjects in the prevalence of EIB. In US Olympic winter sports the prevalence of EIB by using an exercise challenge test was 26% in female and 18% in male athletes, with a combined percentage of 23%.<sup>E104</sup> When using EVH as a surrogate challenge for EIB, other studies failed to find such a difference in prevalence between the sexes.<sup>E30,E105</sup> Using questionnaires and methacholine challenges, investigators<sup>E106</sup> found that the prevalence of both exercise-associated asthma symptoms and BHR was higher in female than male athletes.

Data suggest there can be racial/ethnic differences in the prevalence of EIB. A standardized free running test with peak flow monitoring demonstrated that African Americans had a

higher prevalence of EIB than white subjects (13% and 2%, respectively).<sup>E107</sup> When assessing 9-year-old children with cycle ergometry in Great Britain, ethnic differences in EIB were also evident, with Asian children (originating from the Indian subcontinent) having a prevalence 3.6 times higher than that of white inner-city children.<sup>E108</sup>

*Summary Statement 3:* Evaluate EIB in elite athletes by using objective testing. [Strength of Recommendation: Strong; Evidence: B]

No test for EIB is fully sensitive or specific, and thus no test is sufficient to substantiate or exclude the diagnosis of EIB in all athletes. Additional variability can be caused by seasonal and environmental influences, which might affect the ability to detect BHR.

Reports of the occurrence of asthma symptoms in elite athletes have varied from none to 61%,<sup>E109,E110</sup> depending on the sport and environment.<sup>E84,E104,E109-E119</sup> In fact, higher prevalence rates are reported in certain populations, such as elite endurance athletes, and in unique environments, such as competitive skaters and cross-country skiers.<sup>E119-E121</sup> Some investigators suggested that endurance athletes, whether summer or winter, had considerably more symptoms than athletes participating in less aerobic sports.<sup>E109,E110</sup> Overall, this study suggested that, based on symptoms, as many as 1 in 4 athletes had EIB. By using a questionnaire in recreational athletes, a similar prevalence of asthma symptoms was reported.<sup>E122</sup> However, these data do not suggest that the subjects who report asthma are the same as those who have a positive exercise challenge test result. However, a similar prevalence of EIB in Winter Olympics athletes has been reported based on objective data by using an exercise challenge.<sup>E104</sup> Certain populations have a higher than expected prevalence based on unique circumstances, such as the high prevalence of EIB in skaters (20% to 35%) that has been attributed to high emission pollution from ice-cleaning equipment and cold dry air.<sup>E113,E114</sup> A similar example is the extremes of cold dry air, such as that to which cross-country skiers are exposed, which might increase the prevalence of EIB to 30% to 50%.<sup>E123</sup> Similarly, another study found as many as 78% of elite cross-country skiers have symptoms, BHR, or both.<sup>E124</sup>

Athletes who participated in the 1996 Summer Olympics also had variations in EIB prevalence, which might have depended on the sport in which they participate. For example, long-distance runners were found in one study to have a prevalence of 17%, whereas speed runners had a prevalence of 8%.<sup>E125</sup> Whether these differences are significant might depend on how the test was performed rather than on a difference in the sports for athletes who expend a similar amount of work. By survey, none of the US Olympic divers and weightlifters had symptoms, whereas 45% of mountain bikers experienced symptoms, which is consistent with the hypothesis that endurance sports have a higher prevalence of associated EIB during sport participation.<sup>E110</sup>

Poor air quality can also be associated with a high prevalence of EIB in athletes.<sup>E126</sup> In swimmers chloramines above the water can trigger EIB. Interestingly, swimmers with longer duration of exposure (>100 hours of chlorinated pool exposure) tend to have a higher prevalence of EIB.<sup>E127</sup> Discontinuation of swimming resulted in a decreased incidence of EIB.<sup>E128</sup> As noted previously, the high prevalence of EIB in skaters (20% to 35%) has been attributed to high emission pollution from ice-cleaning equipment and cold dry air.<sup>E129,E130</sup>



Athletic fields and school playgrounds in an urban environment might present a major health concern. Daily measurements (62 days) of air pollutants at a university soccer field in proximity to major highway traffic showed extremely high levels of airborne particulate matter that were related to significant decreases in lung function in soccer players.<sup>E131</sup> High levels of ambient ozone, as well as emissions and particulate matter from vehicular traffic, have been shown to enhance the EIB response in asthmatic patients.<sup>E132</sup>

Seasonal variation of EIB is also described in Olympic athletes<sup>E97</sup> and the general population.<sup>E133,E134</sup> For example, when using a 6.5% decrease in FEV<sub>1</sub> with running, 28% of runners had probable EIB. Of these runners, 22% had EIB that occurred only in the winter, and 7% had EIB only during the pollen season.<sup>E97</sup> A seasonal difference was also demonstrated in another investigation,<sup>E135</sup> which found that 35% of runners training in the cold reported an increased prevalence of EIB compared with summer, when the prevalence was less. Data collected from the Summer Olympics between the years 1992 and 2008 have documented an increasing percentage of elite athletes reporting asthma. This trend follows a similar increase noted in the general population.<sup>E136</sup>

**Summary Statement 4:** Perform a standardized bronchoprovocation (exercise or a surrogate) challenge to diagnose EIB because the prevalence of EIB will vary with the type of challenge and the conditions under which the challenge is performed. [Strength of Recommendation: Strong; Evidence: A]

By examining responses to history questions on the intake forms of athletes participating in the 1996 Summer Olympic Games, as required by the US Olympic Committee, investigators<sup>E110</sup> found 0% to 45% of summer athletes, depending on the sport, answered questions compatible with having EIB. The prevalence varied significantly among different sports, with nonendurance sports having minimal levels and endurance sports having higher prevalence rates. By using the same data extraction method, the same researcher<sup>E109</sup> found that up to 60.7% of athletes participating in Nordic skiing events responded to questions that suggested they had EIB.

The use of self-reported symptoms to make the diagnosis of EIB will likely misdiagnose asthma in patients who do not have EIB and miss persons with the condition. A limitation of determining prevalence by survey is evident in multiple studies<sup>E28,E84,E86</sup> in which results showed participants who had symptoms did not necessarily have a positive challenge result and those who had a positive challenge result did not necessarily have symptoms.

It has also been demonstrated that symptoms are neither sensitive nor specific to suggest a positive EVH test result as evidence for EIB.<sup>E28</sup> This study found that 36% of college athletes without symptoms had a comparable decrease of 10% in FEV<sub>1</sub>, and a similar number (35%) of those with symptoms had such a decrease with EVH.<sup>E28</sup>

Another study used different techniques in an attempt to clarify the prevalence of EIB. These investigators challenged 50 elite athletes with and without a history of asthma documented by questionnaire with methacholine provocation and EVH.<sup>E137</sup> The results showed that of the 42 athletes who reported respiratory symptoms, 9 had a positive methacholine test result, and 25 had a positive EVH test result. Methacholine had an excellent negative predictive value but only a 36% sensitivity for identifying those with a positive EVH test result. These findings are

consistent with the observations found in 2 more studies, which demonstrated that EIB and asthma symptoms do not correlate well with exhaled nitric oxide levels, results of bronchoalveolar lavage, or challenges with AMP or histamine.<sup>E117,E118</sup>

Elite athletes can have a high prevalence of EIB, which can be associated with extreme atmospheric conditions, such as high levels of pollen, pollution, dry air, and chemicals, particularly in the training environment. EIB can be demonstrated in persons without symptoms, but symptoms are not a sensitive predictor of EIB.<sup>E138</sup> The prevalence of EIB also can be affected by age, sex, ethnicity, urbanization, and, most significantly, the diagnostic method used to detect it.

## VIII. DIAGNOSIS

Symptoms of EIB primarily include wheeze, chest tightness and shortness of breath (dyspnea), and cough; however, they can also include chest pain (primarily in children), excessive mucus production, or feeling out of shape when the patient is actually in good physical condition.<sup>E28,E84,E85,E139,E140</sup> Because these symptoms also occur with other conditions, a diagnosis of EIB based only on symptoms lacks any reasonable diagnostic sensitivity or specificity to predict a positive exercise challenge result in adults or children.<sup>E84,E93,E141-E143</sup> Thus the diagnosis of EIB should never be made based on symptoms alone when unaccompanied by data from an objective exercise or surrogate challenge (Fig E1).<sup>E28,E84,E85,E139,E144-E146</sup>

**Summary Statement 5:** In subjects with no current clinical history of asthma, use an indirect ungraded challenge (eg, exercise challenge or surrogate testing, such as with EVH) for assessing EIB in the recreational or elite athlete who has normal lung function. [Strength of Recommendation: Strong; Evidence: D]

**Summary Statement 6:** Use an indirect graded challenge (eg, mannitol, if available) for assessing EIB in recreational or elite athletes who have normal to near-normal lung function and who might currently require treatment for the prevention of EIB or asthma. [Strength of Recommendation: Strong; Evidence: D]

Diagnostic challenges used to identify airway hyperresponsiveness are of 2 types classified based on mechanism of action: (1) direct challenges in which a single pharmacologic agent, such as methacholine or histamine, is the provoking substance administered exogenously that acts directly through receptors on airway smooth muscle to cause contraction and (2) indirect challenges in which exercise or a surrogate, such as EVH; inhaled osmotic agents, such as mannitol or hypertonic saline; or inhalation of AMP is the provoking agent that in turn triggers endogenous mediator release that acts to cause airway smooth muscle contraction. These mediators act on specific receptors on bronchial smooth muscle to cause bronchoconstriction. Indirect challenges are more specific in reflecting BHR caused by the presence of airway inflammation and are preferred as a way to confirm underlying asthma and potentially the need for ICSs.<sup>E28,E84,E85,E139,E144-E146</sup> In addition, indirect challenges are recommended for monitoring asthma therapy because BHR is most often associated with inflammation,<sup>E28,E84,E139,E144-E146</sup> which is diminished by ICS therapy.<sup>E85,E146-E149</sup>

**Summary Statement 7:** Perform an indirect challenge (eg, exercise challenge or surrogate testing, such as with EVH or mannitol, where available) instead of a direct challenge (eg, methacholine) for assessing EIB, recognizing that an indirect challenge is more sensitive for detection of EIB than a direct



(eg, methacholine) challenge. [Strength of Recommendation: Strong; Evidence: B]

Direct challenges with methacholine, an approved agonist, can be performed in an office setting by trained personnel. The challenge, as described in a consensus statement by the ATS,<sup>E145</sup> requires administering increasing concentrations of methacholine by means of inhalation and measuring FEV<sub>1</sub> levels after each dose. Although the direct challenge is used as a screening test for chronic asthma, especially to rule out asthma, it is not useful to detect EIB. This is because it has low specificity for EIB as a result of reflecting the effect of only a single agonist (Fig E3).<sup>E85,E144-E146,E150,E151</sup>

**Summary Statement 8:** Ensure the ventilation reached and sustained during exercise challenge testing is at least 60% of the maximum voluntary ventilation by using dry medical grade air to achieve an adequate challenge. If ventilation cannot be measured, ensure the heart rate as a percentage of HR<sub>max</sub> that is reached and sustained is at least 85% in adults and 95% in children and elite athletes. [Strength of Recommendation: Strong; Evidence: B]

Indirect challenges should also be conducted only by trained personnel using standardized protocols. For example, laboratory-based exercise should be performed as described in the consensus statement published by the ATS.<sup>E2,E145</sup> Such a laboratory challenge controls minute ventilation and water content of inhaled air.<sup>E2,E85,E144,E145</sup> Exercise ramp-up should be brisk within 2 to 3 minutes to reach a heart rate of 85% of maximum and an exercise duration of no more than 8 minutes, of which 6 minutes is maximum exercise, while a maximum heart rate of 95% for children with a preferred exercise duration of 6 minutes. It is preferable to use a source of dry air (medical grade) at 20°C to 25°C to achieve more than 40% of the patient's calculated maximum voluntary ventilation.<sup>E2,E85,E144,E145</sup> Medical air can be supplied directly from a compressed air tank with a demand valve that delivers air at high flow rates or alternatively supplied to a balloon reservoir bag (eg, Douglas bag) fitted with a 2-way nonrebreathing valve before being attached to a mouthpiece or facemask.<sup>E152,E153</sup> Measurement of ventilation should be encouraged because it is the level of ventilation reached and sustained, which is key to providing a maximal stimulus.<sup>E154</sup> This can be measured by using a spirometer that measures minute ventilation of expired air in real time (eg, Bi-directional Universal Ventilation Meter, VacuMed, Ventura, Calif). In the absence of this, maximal heart rate can be used alternatively and is estimated by using the following formula:

$$220 - \text{Age (in years)}^{E2};$$

however, a more accurate equation, which was published recently, to predict HR<sub>max</sub> is as follows:

$$208 - 0.7 \times \text{Age}^{E155}$$

Ideally, the exercise ventilation should be greater than 60% of predicted maximum (ie, >21 times FEV<sub>1</sub>)<sup>E2,E144,E145</sup>; very well-conditioned subjects might require the exercise intensity to be greater than 90% HR<sub>max</sub>. There might be a need to reach a higher target HR<sub>max</sub> of 95% for adolescent children because one study in patients 9 to 17 years of age demonstrated the decrease in FEV<sub>1</sub> was 25.1% at 95% HR<sub>max</sub> but 8.8% when the children reached only 85% HR<sub>max</sub> (Fig E4).<sup>E139</sup>

Spirometry should be performed at baseline according to the ATS standards of reproducibility before exercise challenge and at predetermined time points after exercise, usually at 5, 10, 15,

30 minutes and occasionally 45 to 60 minutes after exercise. A pre-exercise value is obtained by performing a full forced vital capacity (FVC) maneuver at baseline.<sup>E2,E85,E144,E145</sup> The International Olympic Committee Medical Commission Independent Panel on Asthma recommends that FEV<sub>1</sub> should be recorded beginning as soon as 3 minutes after completion of the challenge to overcome the problem of posttest respiratory fatigue. Reproducibility of FEV<sub>1</sub> after exercise is desirable because at times moderate-to-severe decreases have occurred. A measurement at 1 and or 3 minutes after exercise for reasons of safety might be warranted in persons who are suspected of having large decreases in FEV<sub>1</sub>. FEV<sub>1</sub> is often performed without having the patient perform full FVC maneuvers to avoid causing the patient to become tired because of the spirometric efforts (eg, 2-4 seconds of expiration). The highest FEV<sub>1</sub> at each time point is used to calculate the percentage decrease from baseline. A 10% or greater decrease in FEV<sub>1</sub> from the pre-exercise value at any 2 consecutive time points within 30 minutes of ceasing exercise can be considered diagnostic of EIB.<sup>E2,E85,E144,E145,E154</sup> Reproducibility of FEV<sub>1</sub> after exercise becomes essential in cases in which borderline decreases in FEV<sub>1</sub> have resulted. If a greater decrease in FEV<sub>1</sub> is required, such as a decrease of 20% in FEV<sub>1</sub>, as in some pharmaceutical studies, then only 1 time point might be necessary to be diagnostic of EIB.

**Summary Statement 9:** Perform EVH as the preferred surrogate challenge for the athlete without a current history of asthma participating in competitive sports in whom the diagnosis of EIB is suspected. [Strength of Recommendation: Strong; Evidence: D]

**Summary Statement 10:** If an indirect graded challenge (eg, mannitol) result is negative and EIB is still suspected, an ungraded challenge should be considered. [Strength of Recommendation: Weak; Evidence: B]

The profile of the decrease in FEV<sub>1</sub> after an exercise or EVH challenge should be examined to determine whether the decrease is sustained and not the product of a single measurement that might represent an artifact because of inadequate spirometric effort at 1 or more time points. There might be variability in the airway response to exercise when more than 1 test is performed, particularly in those with milder airway responses, and thus repeat testing might need to be considered in some cases in which EIB is strongly suspected.<sup>E141,E156</sup>

However, there is no single test that will identify all patients with EIB.<sup>E84</sup> Decreases in FEV<sub>1</sub> consistent with EIB can occur in subjects who are subsequently found to have other conditions.<sup>E85</sup> A flat or "truncated" inspiratory flow-volume loop on the flow-volume curve suggests an upper airway dysfunction rather than EIB.<sup>E85</sup> EILD can occur independently or coexist with EIB.

Exercise challenge by treadmill is most easily standardized for office practice or a hospital laboratory. Alternative exercise challenges using cycle ergometry can be more difficult to perform and might provide a suboptimal exercise stimulus compared with the treadmill challenge.<sup>E154</sup> Furthermore, field challenge and free running are challenge tests that are more difficult to standardize.<sup>E2,E85,E142,E144,E145</sup>

Although sport governing bodies require specific cutoff values to diagnose EIB, there is no specific decrease in FEV<sub>1</sub>, and there is no single absolute cutoff for a decrease in FEV<sub>1</sub> or change in some other spirometric measure that clearly and unequivocally distinguishes between the presence and absence of EIB.<sup>E85</sup> The ATS has suggested that the postexercise decrease in FEV<sub>1</sub> required

to make the diagnosis must be 10%, whereas other groups have suggested a decrease of 13% to 15% is necessary to make the diagnosis.<sup>E2,E144,E145</sup> A decrease in FEV<sub>1</sub> of 15% after a “field” challenge and a decrease of 6% to 10% in the laboratory have also been recommended.<sup>E2,E85,E144,E145</sup>

Surrogate challenges for exercise in which a hyperosmolar agent, mannitol (graded challenge), or EVH (ungraded challenge) are used are increasingly being recommended by organizations that regulate drug use by elite athletes. EVH should only be performed by highly trained specialists, and all safety precautions should be observed. EVH can cause substantial decreases in FEV<sub>1</sub> in a patient with reduced lung function caused by airway inflammation. The EVH test should be performed with caution, especially in patients with an FEV<sub>1</sub> of less than 80% of predicted value. The EVH test should not be performed on patients in whom FEV<sub>1</sub> is less than 75% of predicted value.<sup>E2,E85,E144,E145</sup> For all these challenge tests, treatments that are effective at attenuating or inhibiting airway hyperresponsiveness should be withheld for an appropriate time before testing to ensure sufficient washout of the drug, so that it does not influence the airway response (Table E1).<sup>E158-E170</sup>

## IX. DIFFERENTIAL DIAGNOSIS

*Summary Statement 11:* To differentiate between EIB and EILD, perform appropriate challenge tests (eg, exercise, EVH, and mannitol for EIB) and potentially flexible laryngoscopy during exercise for diagnosis of EILD. [Strength of Recommendation: Strong; Evidence: B]

EILD, primarily vocal cord dysfunction (VCD) and also other glottic abnormalities, can be elicited by exercise and mimic EIB. Inspiratory stridor is a differentiating hallmark sign with EILD and not with EIB. However, the presence of inspiratory symptoms does not necessarily differentiate athletes with and without EILD.<sup>E171,E172</sup> Flattening of the inspiratory curve on spirometric maneuvers can be seen concomitant with symptoms (Fig E1). EILD can occur alone or with EIB. Failure to respond to asthma management is a key historical feature suggesting EILD.

Since the initial description of VCD as a functional disorder that mimicked attacks of asthma,<sup>E173</sup> VCD and glottis structural abnormalities elicited with exercise have been increasingly recognized. These functional and structural disorders can be grouped as EILD, including (1) paradoxical VCD, (2) exercise-induced laryngeal prolapse,<sup>E174</sup> (3) exercise-induced laryngomalacia,<sup>E175</sup> and (4) variants, including arytenoid collapse while the vocal cords move normally.<sup>E176</sup> EILD occurs in all age groups, especially among young adult female elite athletes.<sup>E177</sup> VCD is more common in middle school- to high school-aged athletes than college-aged athletes.<sup>E178</sup> There is a question as to whether VCD and exercise-induced laryngeal malacia in children and adolescents are separate clinical entities.<sup>E179</sup>

Bronchial provocation challenge results with methacholine, exercise, and EVH can be negative in patients with EILD who do not otherwise have BHR. The onset of breathing difficulties occurs and peaks during exercise with EILD, rather than peaking after exercise with EIB. Medications used to treat asthma, such as  $\beta_2$ -agonists, are ineffective to prevent or reverse EILD. EILD can be suspected based on bronchial provocation challenges with EVH, methacholine, and/or exercise, demonstrating variable extrathoracic airway obstruction.<sup>E180</sup>

Inspiratory stridor with throat tightness during maximal exercise resolves within approximately 5 minutes of discontinuation of exercise in patients with EILD. Inspiratory stridor with EILD contrasts with EIB, in which case dyspnea generally occurs after exercise, peaks 5 to 20 minutes after stopping, and involves expiration rather than inspiration. There can be variations in the timing of the manifestations of EILD symptoms, depending on such factors as the duration and intensity of the exercise.

In patients with VCD, direct observation of vocal cord adduction by means of laryngoscopy and flattening or truncation of the inspiratory portion of the spirometric flow-volume loop are the hallmarks for diagnosis (Fig E1). These findings can be seen only during symptomatic periods. Methacholine challenge can be used to elicit VCD.<sup>E181,E182</sup> Additional evidence of VCD can be suggested by examining a video of the patient recorded while exercising in the natural setting at the time that inspiratory stridor is heard.<sup>E183</sup> Diagnosis can be made directly by using continuous laryngoscopy during exercise challenge.<sup>E184</sup> Spirometry and laryngoscopy with sound recording can be performed during exercise, detecting minor and major aryepiglottic and vocal cord abnormalities.

Exercise-induced laryngeal prolapse has been seen in otherwise healthy athletes and can present with subtotal occlusion of the larynx. This condition can result from mucosal edema from the aryepiglottic folds being drawn into the endolarynx (laryngomalacia).<sup>E174</sup> Laryngoscopic evaluation at rest can be normal, and various laryngeal abnormalities can be elicited only with exercise challenge.<sup>E185</sup>

Laryngomalacia is associated with diminished laryngeal tone, resulting in supraglottic collapse, and is usually a congenital condition.<sup>E186</sup> Laryngomalacia is the most common cause of inspiratory stridor in infants<sup>E187</sup> but might not manifest until later childhood with participation in competitive sports.<sup>E175,E186-E190</sup> It has been questioned clinically whether exercise-induced VCD and exercise-induced laryngomalacia in children and adolescents present as the same clinical syndrome.<sup>E179</sup> Although the typical anatomic features of congenital laryngomalacia (shortened aryepiglottic folds or retroflexed epiglottis) might not be seen, other presentations, such as profound arytenoid redundancy and prolapse, can be seen during nasolaryngeal endoscopy. As in infants with laryngomalacia, supraglottoplasty can improve late-onset disease.<sup>E186,E187</sup> Laryngomalacia can also be seen in adults.<sup>E191</sup>

Concurrent laryngeal abnormalities can be seen in patients with VCD. Laryngoscopy can identify findings suggestive of gastroesophageal reflux disease (GERD), chronic laryngitis, laryngomalacia, vocal cord motion impairment, nodules, and subglottic stenosis, especially in patients in whom exercise induces symptoms.<sup>E192</sup> EILD can coexist with EIB. Inspiratory stridor is the signature clinical feature suggesting EILD rather than EIB.

Gastroesophageal reflux anatomic findings can be seen on laryngoscopy in children and adults with EILD, but whether they are causative or concomitant is difficult to establish. Empiric pharmacologic treatment of GERD in juveniles with VCD has been recommended because posterior laryngeal changes associated with GERD are common in these patients.<sup>E193</sup>

Although laryngopharyngeal reflux can be a contributing factor in many patients with EILD, there is very little supporting objective evidence. The sensitivity and specificity of laryngoscopic examination to diagnose laryngopharyngeal reflux are also controversial.<sup>E194</sup> Although there might be a clinical suspicion of

laryngopharyngeal reflux, there is an absence of an objective gold standard to establish this diagnosis. Although great attention has been given to EILD and other dysfunctional breathing disorders in the differential diagnosis, therapeutic strategies might require a multidisciplinary approach, including speech therapy and addressing possible psychophysiological stress.<sup>E195</sup>

**Summary Statement 12:** To determine whether exercise-induced dyspnea and hyperventilation are masquerading as asthma, especially in children and adolescents, perform cardiopulmonary exercise testing. [Strength of Recommendation: Moderate; Evidence: C]

Exercise-induced respiratory symptoms have been described as an “epidemic” among adolescents.<sup>E196</sup> EILD and exercise-induced hyperventilation are common, and the prevalence is uncertain, as described primarily in uncontrolled case reports.<sup>E197,E198</sup> Chest discomfort perceived as dyspnea during vigorous exercise can be associated with hypocapnia from hyperventilation without bronchoconstriction, especially in children and young adolescents previously given a diagnosis of and having been treated for EIB.<sup>E140,E199-E203</sup>

It has been demonstrated that exercise-induced lactic acidosis is causally involved in hyperventilation. However, lactic acidosis does not represent the only additional stimulus of ventilation during intense exercise. Sensory input from exercising muscles, such as muscle afferents, can also trigger hyperventilation.<sup>E204,E205</sup>

Idiopathic hyperventilation is a poorly understood condition in which patients have sustained hyperventilation, hypocapnia, and dyspneic drive.<sup>E206</sup>

Perhaps the most common reason for exercise-induced dyspnea in children is physiologic (poorly conditioned) limitation without bronchospasm or underlying disease.<sup>E199</sup> Limits in exercise performance and respiratory system oxygen transport can occur in highly fit adults.<sup>E207</sup> This might be due to flow limitation in the intrathoracic airways because of narrowed hyperactive airways or secondary to excessive ventilatory demands superimposed on a normal maximum flow-volume envelope. In addition, exercise-induced arterial hypoxemia occurs as a result of an excessively widened alveolar-arterial oxygen pressure difference. This inefficient gas exchange might be attributable in part to small intracardiac or intrapulmonary shunts of deoxygenated mixed venous blood during exercise. Finally, fatigue of the respiratory muscles resulting from sustained high-intensity exercise and the resultant vasoconstrictor effects on lung muscle vasculature will also compromise oxygen transport and performance. Exercise in the hypoxic environment of even moderately high altitudes will greatly exacerbate the negative influences of these respiratory system limitations to exercise performance, especially in highly fit subjects.

Dyspnea on exertion is present in obese patients. This dyspnea has been strongly associated with an increased oxygen cost of breathing without bronchoconstriction in otherwise healthy obese women.<sup>E208</sup> Exercise capacity has been variously reported as unchanged in obese female subjects to being reduced at or near maximal effort.<sup>E209</sup>

Cardiopulmonary exercise testing should be performed with close observation to assess the clinical presentation (Fig E1).

**Summary Statement 13:** Perform spirometry, as well as detailed pulmonary examination, to determine whether shortness of breath with exercise is associated with underlying conditions, such as COPD, or restrictive lung conditions, such as obesity, skeletal

defects (eg, pectus excavatum), diaphragmatic paralysis, or interstitial fibrosis, rather than EIB. [Strength of Recommendation: Moderate; Evidence: C]

Dyspnea with exertion in some obese patients might not be a manifestation of EIB.<sup>E210</sup> Idiopathic pectus excavatum can be associated with exercise symptoms, including chest pain, dyspnea, or impaired endurance. Even in the absence of clinical symptoms, restrictive lung defects and lower airway obstruction are common (Fig E1).<sup>E211,E212</sup>

Scoliosis has been associated with decreased exercise tolerance. Patients with mild scoliosis can be asymptomatic at rest but with exercise can have decreased tidal volume, as well as hypercapnia and hypoxia.<sup>E213</sup>

Although diaphragmatic paralysis has a predictable effect on lung function, the symptoms depend on pre-existing heart-lung diseases and can mimic various cardiorespiratory processes, including EIB.<sup>E214,E215</sup>

Patients with interstitial lung disease frequently have dyspnea with exercise. These patients’ exercise limitations appear to be related to arterial hypoxemia and not respiratory mechanics. Their dyspnea is often fixed and reproducible.<sup>E216,E217</sup>

**Summary Statement 14:** Consider a diagnosis of EIAAna instead of EIB based on a history of shortness of breath or other lower respiratory tract symptoms accompanied by systemic symptoms (eg, pruritis, urticaria, and hypotension). [Strength of Recommendation: Moderate; Evidence: C]

EIAAna is characterized by the exertion-related onset of cutaneous pruritus and warmth, generalized urticaria, and the appearance of such additional manifestations as shortness of breath, upper respiratory tract distress, vascular collapse, and gastrointestinal tract symptoms. This must be differentiated from asthma, cholinergic urticaria, angioedema, and cardiac arrhythmias, which are recognized as exertion-related phenomena in predisposed patients but are distinct from EIAAna.<sup>E218</sup> There is variability in the reproducibility of EIAAna symptoms given similar testing conditions.

Episodes of food-dependent exercise-induced anaphylaxis (FDEIAAna) might or might not be dependent on ingestion of identifiable foods.<sup>E219,E220</sup> The cumulative effect of exercise and food ingestion can trigger the mediator release and anaphylaxis, whereas this is not the case for each of these triggers independently.

Foods reported as predisposing factors range from shellfish, eaten 4 to 24 hours before EIAAna, to seemingly benign foods, such as celery, eaten before or after exercise.<sup>E219,E220</sup> Skin testing with foods might be helpful in eliciting the trigger when history taking does not.<sup>E221</sup> Serum tryptase measurements might help in confirming the diagnosis of EIAAna.<sup>E221</sup> FDEIAAna occurs in both children and adults.<sup>E218-E223</sup>

Wheat gliadin has been identified as the cause of FDEIAAna caused by wheat.<sup>E224</sup> It has been further determined that crosslinking between tissue transglutaminase and omega-5 gliadin-derived peptides increases IgE binding. The tissue transglutaminase becomes activated in the patients’ intestinal mucosa, and large allergen complexes become capable of eliciting anaphylaxis.<sup>E225</sup> Exercise and aspirin have been shown to increase the levels of circulating gliadin peptides in patients with wheat FDEIAAna, suggesting facilitated allergen absorption from the gastrointestinal tract.<sup>E226</sup>

Although skin testing to the specific foods, commercial or fresh food extracts, or crude gliadin is most used, measurement of IgE

levels specific to epitope peptides of omega-5 gliadin or recombinant omega-5 gliadin might be useful as an *in vitro* diagnostic method (Fig E1).<sup>E226,E227</sup>

Oral challenge with gluten alone or along with aspirin and alcohol is a sensitive and specific test for the diagnosis of wheat-dependent EIA. Exercise is not an essential trigger for the onset of symptoms in these patients.<sup>E228</sup>

FDEIA can have a delayed onset for an unpredictable number of hours. Therefore it has been suggested that in such patients exercise should be avoided 4 to 6 hours after specific food ingestion. In patients with wheat gliadin-associated EIA, a gluten-free diet is recommended.<sup>E224,E229</sup>

Susceptible patients might be advised to take an antihistamine before exercise and should carry self-injectable epinephrine, which is the primary treatment for anaphylaxis.<sup>E229</sup>

**Summary Statement 15:** Refer to appropriate specialists (eg, cardiologist or pulmonologist) to perform cardiopulmonary testing when breathlessness with exercise, with or without chest pain, might be caused by heart disease or other conditions in the absence of EIB. [Strength of Recommendation: Moderate; Evidence: C]

Although the incidence of cardiac-related dyspnea with exercise in young healthy patients is minimal, it remains an important differential in patients with EIB (Fig E1). Idiopathic pulmonary arterial hypertension can occur in both adults and children. Patients with primary pulmonary hypertension can demonstrate peripheral airway obstruction, poor oxygenation, and early physiologic aerobic limits restricting exertion and, in children, documentation of significant reversibility of lower airway obstruction.<sup>E230-E232</sup>

A case report documents how idiopathic pulmonary arterial hypertension can masquerade as asthma. Two adult nonsmokers presenting with wheezing, chronic cough, and irreversible obstructive lung disease were given a diagnosis of adult-onset severe refractory asthma but actually had dilation of the central pulmonary arteries, compressing the mainstream bronchi.<sup>E233</sup>

“Cardiac asthma” can be considered one presentation of cardiac dyspnea caused by cardiogenic pulmonary edema. The pathogenesis might be reflex bronchoconstriction as a manifestation of pulmonary venous hypertension. In distinguishing cardiac from pulmonary dyspnea, the most useful studies include B-natriuretic peptide measurement, echocardiography, and, if needed, a cardiopulmonary exercise test.<sup>E198</sup> Congestive heart failure can present with dyspnea on exertion. Hyperpnea with exercise can occur without lung function impairment. Ventilation-perfusion mismatch in exercise can be enhanced by increased treatment of heart failure.<sup>E234</sup>

Hypertrophic cardiomyopathy is well known to cause sudden death in young athletes, with an annual 1% mortality rate.<sup>E235</sup> Patients can have dyspnea and chest pain that improve with  $\beta$ -blockers.<sup>E236</sup>

Cardiac dysrhythmias can also cause dyspnea with exercise. Supraventricular tachycardia can cause EIB in children.<sup>E199</sup> Young adults with complete heart block can have shortness of breath, dyspnea on exertion, syncope, dizziness, or fatigue.<sup>E237</sup>

Vascular rings of the aorta are rare but can present as asthma. Spirometry in these patients reveals decreased peak expiratory flow and truncation of the expiratory flow-volume loop with normal FVC, FEV<sub>1</sub>, and FEV<sub>1</sub>/FVC ratio values. Chest radiographs are significant for a right aortic arch.<sup>E238</sup>

Pulmonary arteriovenous malformations and disorders with right-to-left shunts can cause exercise-induced dyspnea because of hypoxemia, without associated bronchoconstriction. Hereditary hemorrhagic telangiectasia, atrial septal defects, ventricular septal defects, and Osler-Rendu-Weber syndrome are among the primary causes. Cardiopulmonary exercise testing, as differentiated from pulmonary function testing for EIB, is an appropriate noninvasive tool to begin and guide the evaluation of these patients presenting with undiagnosed dyspnea.

The evaluation might require procedures, such as cardiac catheterization, to further delineate the right-to-left shunt.<sup>E239,E240</sup> Exertional dyspnea in symptomatic patients with COPD might be due to the combined deleterious effects of higher ventilatory demand and abnormal ventilatory dynamics but not temporally attributable to bronchoconstriction.<sup>E241</sup> Patients with COPD might have evidence of small-airway dysfunction with increased ventilatory requirements during exercise, likely on the basis of greater ventilation and perfusion abnormalities. These abnormalities also involve changes in end-expiratory lung volume and breathing patterns that are more shallow and rapid than in a comparatively healthy cohort.

Although there are reports of exertional gastroesophageal reflux in healthy subjects, most studies have demonstrated no significant correlations between GERD and EIB.<sup>E242-E244</sup>

Although acid reflux can be common in patients with EIB, many patients with exercise-related respiratory symptoms can receive a misdiagnosis of asthma when they truly have exercise-onset GERD (Fig E1).<sup>E245</sup> Some controversies exist in the treatment of GERD and EIB. One study demonstrated that symptoms of acid reflux related to running were relieved by a proton pump inhibitor, but the respiratory symptoms of EIB were not relieved by proton pump inhibitors.<sup>E246</sup> In contrast, other investigators have reported improvements in exercise-related breathing symptoms when patients were treated with proton pump inhibitors.<sup>E245</sup>

Impaired oxidative phosphorylation in working muscle disrupts the normal regulation of cardiac output and ventilation relative to muscle metabolic rate in exercise.<sup>E247</sup> Deficiencies of mitochondrial enzymes cause a number of severe neurologic syndromes in pediatric patients. Isolated myopathies secondary to enzymatic deficiency have been recognized in adults and might be more prevalent than reported previously (Fig E1).<sup>E248,E249</sup>

**Summary Statement 16:** Refer patients for psychological evaluation when the symptoms (eg, hyperventilation and anxiety disorders) are in the differential diagnosis of EIB. [Strength of Recommendation: Weak; Evidence: D]

Psychological factors can obfuscate the diagnosis in patients with apparent exercise intolerance. Such scenarios, such as subjects, particularly young women, complaining of shortness of breath while running without having stridor, wheezing, or relief with trial bronchodilators, are not uncommon but might be vexing to patients, their parents, and their physicians.

Although VCD and exercise-induced hyperventilation can have functional triggers, differentiating EIB requires subjective and objective assessment.<sup>E250</sup> Mental stress might be one trigger factor in which hyperventilation is seen in patients with asthma-like symptoms with negative asthma test results.<sup>E251</sup> If objective testing does not reveal any bronchoconstriction or other physiologic explanations, then a psychological cause should be considered and addressed with the patient, which might involve a recommendation for psychological consultation (Fig E1).



## X. THERAPY

EIB is a reflection of BHR and, in asthmatic children and adults, ordinarily is due to underlying inflammation. EIB in these subjects, most of whom are not elite athletes, might represent inadequacy of overall asthma control.<sup>E252,E253</sup>

The goal of therapy for EIB is to prevent symptoms induced by exercise, to enhance overall control of asthma, and to ameliorate symptoms rapidly when they occur. Pharmacotherapeutic agents that are effective in controlling chronic asthma generally have bronchoprotective activity for EIB. If asthma is otherwise well controlled, bronchoprotective therapy is administered only as needed. This therapy can be delivered by means of inhalation or oral administration minutes to hours before exercise, respectively. Nonpharmacologic therapies can also be helpful in preventing EIB when used alone or in combination with pharmacotherapy; these are described in the “Nonpharmacologic therapy” section.

Pharmacotherapeutic agents act to prevent or attenuate EIB through various mechanisms with different degrees of effectiveness. None of the available therapies completely eliminates EIB. Pharmacotherapy shifts the dose-response relationship to a more favorable position after exercise.<sup>E254,E255</sup> The efficacy of a given agent in protecting against EIB can vary at different times and among different subjects.

*Summary Statement 17:* Schedule regular office visits with patients because medications can differ in effectiveness over time because of variability of asthma, environmental conditions, intensity of the exercise stimulus, and tachyphylaxis. [Strength of Recommendation: Strong; Evidence: A]

The variability of effectiveness within a subject might be due to changes in airway responsiveness over time, environmental conditions, and intensity of the exercise stimulus.<sup>E256</sup> The variability among subjects might result from differences in baseline airway responsiveness and susceptibility to tachyphylaxis and perhaps genetic differences.<sup>E85</sup> Pharmacotherapeutic studies supported by pharmaceutical sponsors generally have used parallel groups or crossover designs to compare active drugs with placebo or to compare 2 (or more) active drugs.<sup>E256</sup> The primary end point is most commonly the maximum percentage decrease in FEV<sub>1</sub>, especially for studies submitted to the US Food and Drug Administration (FDA)<sup>E256</sup> in support of a bronchoprotective end point. Peak expiratory flow has also been used as an end point in some studies but not as a primary end point, and it is used less commonly than FEV<sub>1</sub>.

In addition to the maximum absolute decrease in FEV<sub>1</sub> expressed as a percentage of baseline, the results might indicate FEV<sub>1</sub> before and after therapy.<sup>E257</sup> Baseline lung function can also be compared before and after therapy if a bronchodilator response is also being investigated.<sup>E59,E145,E254,E258-E263</sup> Some studies have examined the percentage of subjects protected from EIB after therapy (responder analysis).

The maximum decrease in FEV<sub>1</sub> required to produce a positive test result varies with the situation in which the test is performed. In a clinical setting the decrease in FEV<sub>1</sub> from baseline required to diagnose EIB is usually 10%<sup>E145</sup> or perhaps 13%<sup>E264</sup> or 15%.<sup>E265</sup> As an inclusion criterion in a pharmaceutical trial, a 20% decrease in FEV<sub>1</sub> is usually required to define a positive challenge result (as described in FDA guidance).<sup>E256</sup> In a clinical setting it is desirable to produce as complete protection as possible so that there is no decrease in FEV<sub>1</sub> after exercise with treatment. In a pharmaceutical trial protection might be defined as a less than 10% decrease

in FEV<sub>1</sub> after exercise or 50% protection compared with placebo<sup>E266</sup> for patients who are required to have a 20% decrease at the screening visit. When other end points are used (eg, area under the curve [AUC] or time to recovery), the percentage of protection must also be adjusted for the situation in which the test is performed. Protection has been defined in some studies based only on statistically significant differences between responses after pretreatment with active drug compared with placebo. Attempts to define protection that have clinical relevancy have resulted in concepts such as “complete protection” and “clinical protection.”<sup>E258,E267</sup> Complete protection can be suggested by predefined decreases in FEV<sub>1</sub> percentage within an accepted reference range (eg, <10% for FEV<sub>1</sub>). Clinical protection has been defined as a 50% inhibition of the placebo response to exercise by drug pretreatment.<sup>E263,E266</sup> The 10% decrease is based on the mean plus 2 SDs of the decrease in healthy subjects,<sup>E268</sup> and the 50% protection is based on the coefficient of variation for repeated tests.<sup>E266</sup>

Numerous studies have been assessed in developing the evidence-based recommendations for therapy in this document, but there is little information on the consistency, presence, or absence of drug effect from recurrent testing. Also, most published evidence is from EIB studies in patients with clinically diagnosed (and usually atopic) asthma, and recommendations made herein are based on these data.<sup>E2,E85,E253</sup> Information suggesting possible differences in the pathogenesis of EIB and response to pharmacotherapies in apparently nonasthmatic elite athletes is summarized elsewhere in separate sections (“Pathophysiology” and “Competitive and elite athletes”). It is recommended that this section on therapy be used largely as a starting point on which to base a trial of best therapy for a patient with EIB tempered by the responsiveness and needs of that individual patient over time. Failure to demonstrate inhibition or significant attenuation of apparent EIB by using effective bronchoprotective agents should indicate the need for re-evaluation of the diagnosis.

### β<sub>2</sub>-Adrenergic receptor agonists

*Summary Statement 18:* Prescribe inhaled short-acting β<sub>2</sub>-adrenergic receptor agonists for protection against EIB and for accelerating recovery of pulmonary function when given after a decrease in pulmonary function after exercise. [Strength of Recommendation: Strong; Evidence: A]<sup>E2</sup>

β<sub>2</sub>-Adrenergic receptor agonists are the single most effective therapeutic group of agents for acute prevention of intermittent EIB (Fig E1).<sup>E259</sup> They attenuate or protect against EIB in most patients.<sup>E152,E259,E269-E273</sup> Their effectiveness might be due to their action to enhance recovery of FEV<sub>1</sub> to baseline values when given after a decrease after exercise.<sup>E260,E261</sup>

*Summary Statement 19:* Prescribe a single dose of SABA, LABA, or both on an intermittent basis (ie, <4 times per week) before exercise because this might protect against or attenuate EIB. [Strength of Recommendation: Strong; Evidence: A]

Early investigations of β<sub>2</sub>-adrenergic drugs developed for asthma showed that these agents were highly effective in protecting against EIB when inhaled 5 to 20 minutes before exercise.<sup>E262,E269,E274,E275</sup> Protection was found to last from 2 to 4 hours after inhalation, with most studies showing a duration at the lower end of this interval (Table E1).<sup>E163,E270</sup> There appear to be no substantial differences among SABAs currently in

use.<sup>E163,E270</sup> Mast cell stabilizers, as described below, have been used as add-on therapy to supplement SABAs in increasing the degree of bronchoprotection.<sup>E259,E276</sup>

Multiple LABAs are currently in use and differ in their actions, mainly in their onset of effect (Table E1). Formoterol has a more rapid onset of bronchodilator and bronchoprotective action; in contrast, salmeterol requires 15 to 30 minutes.<sup>E271,E272</sup>

**Summary Statement 20:** Be cautious in daily use of  $\beta_2$ -adrenergic agents alone or in combination with ICSs because this can lead to tolerance manifested as a reduction in duration, magnitude, or both of protection against EIB and a prolongation of recovery in response to SABAs after exercise. [Strength of Recommendation: Strong; Evidence: A]<sup>E2</sup>

Prolonged duration of a bronchoprotective effect for as long as 12 hours has been shown for these drugs after the first dose in  $\beta_2$ -agonist-naïve patients.<sup>E157,E270,E273,E277-E279</sup> However, many patients are not protected for this entire dosing interval, and the optimal dosing interval for bronchoprotection for EIB might be closer to 6 hours on average.<sup>E157,E270,E277,E278</sup>

Prolonged protection with intermittent use of LABAs is sustained,<sup>E157,E277,E280-E282</sup> but daily maintenance use of LABAs (and SABAs) often results in some loss of bronchoprotection (“tolerance”) with cross-tolerance to other  $\beta_2$ -agonists.<sup>E278,E283-E289</sup> Moreover, daily use of LABAs and SABAs might actually increase the severity of EIB.<sup>E288,E289</sup> Of additional concern, the degree of tolerance can increase with increasing bronchoconstriction, potentially putting patients experiencing severe asthma attacks at risk of less bronchodilator responsiveness at the time of greatest need.<sup>E290</sup> Therefore only intermittent use of adrenergic agonists is recommended for bronchoprotection.<sup>E2,E85</sup>

Although some subjects might have a greater propensity than others to develop tolerance, only a small number of patients are required to demonstrate tolerance,<sup>E285,E288-E293</sup> suggesting that tolerance occurs in most patients. Studies addressing this in subjects with and without the Arg16Gly  $\beta_2$ -receptor polymorphism, which has previously suggested susceptibility to  $\beta_2$ -agonist tolerance, demonstrate that these polymorphisms do not influence tolerance to loss of bronchoprotection to  $\beta_2$ -agonists with EIB.<sup>E294</sup> Importantly, tolerance occurs even when patients are also receiving ICSs (Fig E1).<sup>E153,E284</sup>

The onset of tolerance can be rapid. By means of extrapolation of the effects on methacholine-induced bronchoconstriction<sup>E285</sup> and other challenges, such as AMP,<sup>E295</sup> it can occur within 12 to 24 hours after a first dose.<sup>E285,E295-E297</sup> The degree appears to increase with constant  $\beta_2$ -agonist use before it reaches a plateau.<sup>E285</sup> By means of similar extrapolation,  $\beta_2$ -antagonist sensitivity can recover within 72 hours after the last dose of  $\beta_2$ -agonist.<sup>E166,E285</sup>

Tolerance might not develop when  $\beta$ -agonist use is limited to an interval of 48 to 72 hours.<sup>E166</sup> However, a longer period for recovery might be required for other stimuli, such as allergen challenges.<sup>E298</sup> Tolerance is manifested most strikingly by a decrease in the effectiveness of SABAs<sup>E299</sup> and by a shortening of duration of LABA effects,<sup>E153,E273,E278,E280,E284</sup> with one study demonstrating this in less than 3 hours.<sup>E300</sup> There is also evidence this is manifested by prolongation of recovery from bronchoconstriction.<sup>E285,E288</sup> The presence of tolerance is often missed clinically because a patient is rarely challenged at the point of care; consequently, the shorter duration of protection and the prolonged recovery time are not revealed. Importantly, prescribing additional doses of  $\beta_2$ -agonist aerosol immediately

before exercise might unintentionally contribute to further generation of tolerance.

The mechanism or mechanisms by which long-term (daily) use of  $\beta_2$ -agonists lead to tolerance is unclear. A number of observations have led to suggestions for possible mechanisms involved in the development of “tolerance.” Long-term exposure of  $\beta$ -receptors to  $\beta_2$ -agonists results in uncoupling and internalization or sequestration in the cells in which they are degraded.<sup>E301</sup> This net loss in the number of available functional  $\beta_2$ -receptors<sup>E302</sup> results in “downregulation” of responsiveness to  $\beta_2$ -agonists, which manifests as a lack of clinical protection to bronchoconstrictive stimuli. Restoration of sensitivity requires resynthesis of the receptor to the active state. This resynthesis is observed clinically within 72 hours of cessation of exposure to a  $\beta_2$ -agonist.<sup>E166,E285</sup>

Stimulation of mast cell  $\beta$ -receptors normally inhibits mediator release. The process of  $\beta$ -receptor desensitization varies markedly among different cell types; bronchial mast cells are more easily desensitized than bronchial smooth muscle cells.<sup>E301,E303</sup> Downregulation appears to occur more readily in mast cells, as can occur with therapeutic administration of  $\beta_2$ -agonists.<sup>E304,E305</sup>

For this reason, the clinical effects of downregulation are evident more rapidly on mast cells, with an effect on bronchoprotection rather than on smooth muscle and bronchodilation.<sup>E306</sup> The downregulation of mast cell  $\beta$ -receptors not only enhances mediator release but potentially enhances bronchoconstriction as well.<sup>E288,E304,E305,E307,E308</sup>

This  $\beta_2$ -receptor downregulation or tolerance is demonstrated clinically as a reduction in duration of  $\beta_2$ -agonist bronchoprotection to stimuli, such as exercise, which depends on mast cell mediator release for bronchoconstriction.<sup>E309</sup> Tolerance to bronchodilation after EIB is demonstrated by prolongation of the time of recovery from bronchoconstriction in response to usual  $\beta_2$ -agonist doses.<sup>E285,E288,E289</sup>

Downregulation of the  $\beta_2$ -receptor is accompanied by augmentation of pathways mediated through the LT, histamine, and thromboxane receptors. Activation of these receptors has the added potential to enhance bronchoconstriction.<sup>E310-E312</sup> Also, BHR can be induced by non-mast cell mediator mechanisms involving cholinergic agonists, for example, independently or with mast cell mediator mechanisms.<sup>E313-E315</sup>

Use of LABAs as daily monotherapy to provide overall asthma control is not recommended.<sup>E252</sup> When ICSs alone are not adequate in controlling chronic asthma, LABAs are often combined with ICSs to provide effective maintenance therapy; however, there is no convincing clinical evidence that this combination diminishes tolerance to the bronchoprotective effect of LABAs in asthma or EIB with asthma.<sup>E153,E284,E316</sup> LABAs alone used intermittently, up to 3 times a week, do not appear to be associated with tolerance and can be prescribed for EIB.<sup>E166,E317</sup>

## LT inhibitors

**Summary Statement 21:** Consider prescribing daily therapy with leukotriene inhibitors because this does not lead to tolerance and has been shown to attenuate EIB in 50% of patients. It can also be used for intermittent or maintenance prophylaxis; however, it provides incomplete protection and is not effective for reversing airway obstruction. [Strength of Recommendation: Strong; Evidence: A]<sup>E2</sup>

The role of LTs in patients with EIB is to sustain the bronchoconstrictive and inflammatory response, although their role appears to vary significantly among patients. Correspondingly, inhibitors of the LT pathway (LTRAs and lipoxygenase inhibitors) are effective in reducing the severity of the decrease in FEV<sub>1</sub>, as well as enhancing recovery of airway narrowing. Furthermore, there is much variability in their effectiveness, from completely blocking EIB in some asthmatic patients to blocking EIB less so or not at all in others. There is a 30% to 80% attenuation of EIB, with approximately 50% of patients being responders.<sup>E257,E318,E319</sup> These percentages can vary, depending in part on the FEV<sub>1</sub> decrease required to make a diagnosis of EIB (>10%, >15%, or >20%) or used to define protection. Most patients do not experience complete protection.<sup>E258</sup> This is not surprising given that other mediators (eg, PGD<sub>2</sub> and histamine)<sup>E48,E320</sup> are involved in EIB.

Various LTRAs have been found to be effective in attenuating EIB.<sup>E58,E321-E324</sup> Most studies have examined specific LTD<sub>4</sub> receptor antagonists, particularly montelukast. Montelukast is approved by the FDA for treatment of EIB in adolescents and adults. Montelukast acts within 1 to 2 hours of oral administration<sup>E62,E324-E326</sup> and has a bronchoprotective activity of 24 hours (Table E1).<sup>E58,E59,E257,E326-E329</sup> Maximum protection might not be retained in some subjects toward the end of this period.<sup>E329</sup> LTRAs also accelerate the time to recovery from EIB.<sup>E58,E299</sup> Although LTRAs are not as effective overall in attenuating EIB as  $\beta$ -agonists,<sup>E258</sup> tolerance does not develop with long-term use.<sup>E58,E286,E287,E330</sup> The variability in effect on EIB suggests populations of responders and nonresponders similar to those shown for the LT effects on overall asthma control (Fig E1).<sup>E331-E333</sup>

A second group of agents that affects the LT pathway by inhibiting synthesis are the lipoxygenase inhibitors. Lipoxygenase inhibitors have been shown to attenuate EIB when given orally,<sup>E60,E160,E334,E335</sup> but the duration of inhibition of these compounds is relatively short,<sup>E60,E160</sup> and they are not currently recommended for this indication (Table E1). Early-stage studies have demonstrated that inhibition of the LT pathway by 5-lipoxygenase activating protein inhibitors can inhibit EIB.<sup>E336</sup>

### Mast cell stabilizers

*Summary Statement 22:* Consider prescribing inhaled cromolyn sodium and nedocromil sodium (currently not available in the United States as a metered-dose inhaler or dry powder inhaler) shortly before exercise; this attenuates EIB but can have a short duration of action. There is no bronchodilator activity. They might be effective alone or as added therapy with other drugs for EIB. [Strength of Recommendation: Strong; Evidence: A]<sup>E2</sup>

Cromolyn sodium and nedocromil sodium are 2 structurally unrelated compounds that have no bronchodilator activity but have similar bronchoprotective activity against EIB when inhaled.<sup>E57,E259,E337,E338</sup> Several mechanisms have been proposed for these agents, including interference with mast cell mediator release of PGD<sub>2</sub>.<sup>E57,E337,E339</sup> The bronchoprotective effect is rapid<sup>E340</sup> but of short duration (1-2 hours; Fig E1 and Table E1).<sup>E164,E341</sup> These agents can be effective when taken alone or when inhaled shortly before, and perhaps simultaneously with, exercise and might increase overall inhibition of EIB when combined with other drugs used to diminish EIB.<sup>E259,E275,E341,E342</sup> Significant intersubject and

between-study variability has been observed in the ability of these agents to attenuate EIB. Some studies found few or no subjects protected, whereas other studies showed complete protection.<sup>E343,E344</sup> The effectiveness of cromolyn might be dose related.<sup>E344-E346</sup> Long-term use of either drug is not accompanied by tolerance. For this reason and because of their excellent safety profiles and rapidity of action, these agents can be used repeatedly to attenuate EIB in responsive subjects.<sup>E259,E347</sup>

### ICSs

*Summary Statement 23:* Consider prescribing ICSs in combination with other therapies because ICSs can decrease the frequency and severity of EIB but not necessarily eliminate it. [Strength of Recommendation: Strong; Evidence: A]<sup>E2</sup>

EIB in otherwise symptomatic asthmatic patients is best controlled by maintenance anti-inflammatory treatment alone<sup>E148,E348,E349</sup> or in combination with other short-term preventive treatment.<sup>E252,E318,E350</sup> ICSs improve overall asthma control in most patients with chronic persistent asthma. Use of ICSs is associated with attenuation of hyperresponsiveness to direct and indirect stimuli, including exercise.<sup>E351,E352</sup> The dose-dependent effect of ICSs has been observed shortly after the initial (3-4) weeks of treatment<sup>E148,E353</sup>; however, the effects of ICSs are also time dependent with longer duration (12 weeks) of treatment, demonstrating no difference between different doses inhibiting EIB.<sup>E349</sup> The relationship between control of persistent asthma and bronchoprotection, however, is imperfect (Fig E5).<sup>E89,E354</sup> Nevertheless, the degree of EIB is considered a reflection of asthma control (or lack of control), and in particular, moderate-to-severe EIB strongly suggests the need for reassessment of therapy or another diagnosis.<sup>E348</sup>

Some bronchoprotective effect against EIB with high-dose ICSs has been recorded as early as 4 hours after the first dose in adults.<sup>E56,E355,E356</sup> However, it has also been demonstrated that lower doses consistent with the daily treatment of asthma can have a bronchoprotective effect on EIB in children.<sup>E169</sup> After 1 week of therapy, efficacy begins to plateau<sup>E40,E148,E353</sup>; however, bronchoprotection can increase further over weeks or even months until it reaches its final plateau.<sup>E147,E348,E357</sup> This final plateau can come in the form of complete inhibition of EIB.<sup>E349</sup> Bronchoprotection has been shown to occur in 30% to 60% of asthmatic patients with EIB, with marked individual variability ranging from "complete" protection to little or no evidence of protection.<sup>E147</sup> In the absence of definitive dose-ranging and repetitive studies in individual patients, it is not clear whether this reflects distinct subpopulations of responders and nonresponders (eg, a reflection of genetic differences) or whether this is related to EIB severity.

Allergic rhinitis is a common finding in atopic asthmatic patients, and there is some evidence that effective treatment of nasal congestion and obstruction by nasal ICSs is associated with at least mild reduction in EIB.<sup>E357,E358,E359</sup> To some extent, these findings validate the concept of the unified airway theory, which states that allergic rhinitis of the nose and atopic airway inflammation in asthmatic patients are manifestations of similar pathologic processes in the upper and lower respiratory airways, respectively.<sup>E360</sup> It is unclear whether treating EIB with both intranasal corticosteroids and ICSs leads to more effective treatment of EIB in allergic asthmatic patients compared with ICSs alone.



ICSs do not necessarily obviate the need for acute bronchoprotection against EIB (Fig E1).  $\beta_2$ -Adrenergic agonists can be added intermittently, if necessary, for short-term prevention of EIB.<sup>E260,E261</sup> When maintenance ICSs are not sufficiently effective, LTRAs can be used to obtain added protection with low- and medium-dose ICSs<sup>E318,E361</sup> compared with high-dose ICSs<sup>E319</sup> while also administering  $\beta_2$ -agonists for acute bronchoprotection, if necessary.<sup>E121,E253,E267,E362</sup>

**Summary Statement 24:** Do not prescribe daily LABAs with ICS therapy to treat EIB unless needed to treat moderate-to-severe persistent asthma. The ICS might not prevent the occurrence of tolerance from daily  $\beta_2$ -agonist use. [Strength of Recommendation: Strong; Evidence: A]

The preponderance of evidence indicates little amelioration of tolerance to  $\beta_2$ -agonist bronchoprotection by ICSs<sup>E153,E284,E299,E316,E363</sup> and that a shortened degree of bronchoprotection remains when ICSs and LABAs are administered together. Nevertheless, one study that assessed the combination of an ICS and LABA (fluticasone and salmeterol) for maintenance therapy in adult patients indicated better bronchoprotection at 1 and 8.5 hours after dosing compared with the same dose of fluticasone alone during 4 weeks.<sup>E153</sup> In that study most patients receiving the combined therapy also exhibited greater complete (<10% decrease of FEV<sub>1</sub>) protection and overall asthma control. A somewhat similar study with the same agents in children and adolescents also indicated a small persistent effect of bronchoprotection when the combination was used compared with the ICS alone.<sup>E364</sup> LABAs in combination with ICSs when used on demand compared with a low dose of ICS daily can reduce EIB by a similar magnitude over 6 weeks.<sup>E365</sup>

### Anticholinergic agents

**Summary Statement 25:** Consider prescribing inhaled ipratropium bromide for patients who have not responded to other agents; however, its ability to attenuate EIB is inconsistent. [Strength of Recommendation: Weak; Evidence: A]<sup>E2</sup>

Anticholinergic agents have bronchodilator activity<sup>E314</sup> through blocking vagally mediated tone and have been used alone and in conjunction with SABAs with some success in treating acute asthma exacerbations.<sup>E366</sup> However, the efficacy of anticholinergic agents to prevent EIB<sup>E367</sup> has not been consistent in double-blind studies, especially in placebo-controlled trials (Table E1).<sup>E368</sup> Not all patients appear to respond to anticholinergic agents,<sup>E259,E342,E369-E371</sup> and responsiveness can be variable in the same patient.<sup>E370</sup> Studies should be performed to determine the characteristics of the responder population (perhaps based on increased cholinergic contributions to EIB in some patients).<sup>E314,E372</sup>

### Methylxanthines, antihistamines, and other agents

Theophylline and aminophylline are methylxanthines that have been used for long-term maintenance therapy to treat persistent asthma. In recent years, these agents have only been used as adjunct therapy to ICSs or similar maintenance therapy when further control of asthma is needed.<sup>E253,E350</sup> Methylxanthines are nonselective phosphodiesterase inhibitors of the cyclic AMP and cyclic guanine monophosphate pathways that play a role in the pathophysiology of asthma. Methylxanthines are mild bronchodilators and modify EIB in some patients, possibly in part due to their bronchodilator action.<sup>E373,E374</sup> However, there are studies that clearly show no benefit from

methylxanthines administered orally.<sup>E375</sup> Methylxanthines exhibit a relatively narrow therapeutic index with potentially serious adverse events, such as seizures. Selective phosphodiesterase inhibitors are safer and might have efficacy similar to that of the methylxanthines. One such agent, roflumilast, is a phosphodiesterase 4 inhibitor that has been reported to attenuate mild EIB.<sup>E376</sup>

Caffeine also belongs to this class of drug. When caffeine is ingested, it can attenuate EIB in a dose-response manner, with evidence of high doses of caffeine (6-10 mg/kg) inhibiting EIB.<sup>E170,E377,E378</sup> These studies have led to the recommendation of abstaining from caffeine before performing bronchial provocation testing to identify EIB (Table E1).

Some antihistamines have been reported to attenuate EIB,<sup>E61,E62,E320,E379-E382</sup> although with inconsistent results.<sup>E62,E162</sup> Other antihistamines appear ineffective.<sup>E383</sup> A possible explanation for this variability might relate to differences in the intensity and duration of the exercise stimulus, with greater intensity or more severe EIB required for participation of histamine in the pathogenesis of EIB.<sup>E54</sup> Furthermore, histamine is less potent than the other 2 main mediators (leukotriene and prostaglandins) that contribute to EIB (Fig E2).<sup>E384</sup> In addition, the antihistamine class has pharmacodynamic diversity. For example, antihistamines can inhibit mediator activation and release and act on end organs and other histamine receptors.<sup>E385</sup> Different routes of administration and dosages of antihistamines can also be confounding factors in previous studies.<sup>E386</sup>

However, other evidence suggests that antihistamines used in children with allergic rhinitis and EIB might not be bronchoprotective.<sup>E383</sup> The conclusion is that there is an absence of definitive studies that have determined the effectiveness of nasal or systemic antihistamines used to treat allergic rhinitis for an effect on EIB. It is likely to remain common practice to use antihistamines to treat allergic rhinitis in the hope that there will be some effect on EIB. Definitive studies are still needed to confirm the utility of this practice.

Numerous other compounds have been examined for activity against EIB and might have moderate effectiveness in some situations.<sup>E269</sup> These include calcium channel blockers,<sup>E387,E388</sup> inhaled furosemide,<sup>E389-E391</sup> some  $\alpha$ -adrenergic receptor antagonists (oral and inhaled),<sup>E367,E392</sup> inhaled heparin,<sup>E393</sup> and hyaluronic acid.<sup>E394</sup> These agents do not always produce consistent results in preventing EIB. The effectiveness of some of these agents does not necessarily apply to other members of the same drug class, suggesting various mechanisms of action not necessarily related to the obvious mechanisms attributed to each class of drug. The effectiveness of diverse kinds of drugs suggests that there are multiple mechanisms underlying EIB.<sup>E395</sup> Although some of these drugs are not recommended for clinical use against EIB, these and other agents might be useful as probes in studying the possible mechanisms underpinning EIB. In addition, it is important to recognize that these agents can interfere with clinical protocols that seek to examine the effects of other experimental drugs on EIB.

### Nonpharmacologic therapy

**Summary Statement 26:** Use pre-exercise warm-up for EIB since it may be helpful in reducing the severity of EIB. [Strength of Recommendation: Strong; Evidence: A]<sup>E2</sup>



Warm-up before exercise was studied on postexercise bronchoconstriction in athletes with EIB. Continuous warm-up before exercise was shown to cause a significant decrease in postexercise bronchoconstriction in some athletes (Fig E1).<sup>E396</sup> This has importance in patient education, and health care professionals should tell patients that pre-exercise warm-up should be done at 60% to 80% HRmax to provide partial attenuation of EIB; this refractory period can last typically from 1 to 3 hours and occasionally for 4 hours.<sup>E79,E397,E398</sup> However, this does not alleviate the need for medications. Albuterol plus a warm-up provides better protection than the warm-up or albuterol alone.<sup>E77,E399</sup>

The mechanisms for this approximately 50% reduction in airway responsiveness in 50% of patients with EIB with repeated exercise after an initial exercise stimulus is not well understood. Initially, the inhibition or refractory period was considered to be due to the effects of depleting bronchoconstrictive mediator from mast cells, although later this was considered to be due to the release of protective prostaglandins.<sup>E400</sup> However, using inhaled mannitol as a model for EIB, which also demonstrates a refractory period after an initial challenge test, has suggested the protective effect might be due to transient tachyphylaxis at the level of the airway smooth muscle to the mediators of bronchoconstriction, rather than mediator depletion.<sup>E81</sup>

**Summary Statement 27:** Consider with caution the recommendation of reduction of sodium intake and ingestion of fish oil and ascorbic acid supplementation; results are questionable in reducing the severity of EIB. [Strength of Recommendation: Weak; Evidence: A]<sup>E2</sup>

Dietary supplementation as a treatment for EIB has generally seen evidence of significant yet incomplete inhibition of the percentage decrease in FEV<sub>1</sub> after exercise with low-salt diets, omega-3 fatty acids, and ascorbic acid (vitamin C) with up to 3 weeks of supplementation.<sup>E46,E401-E406</sup> Although many of these studies have been performed in small numbers of subjects, they are generally sufficiently powered. However, many studies require validation because of the use of exercise protocols that might provide suboptimal dehydrating stimuli to the airways or unnecessarily prolong the duration of exercise beyond the recommendation of 6 to 8 minutes so that the severity of EIB is reduced because of refractoriness (eg, progressive exercise challenge until volitional exhaustion). This might be the reason why many of these studies have demonstrated inhibition to mild airway responses to exercise, which makes it difficult to extrapolate these results to persons with moderate-to-severe EIB and, for this reason, can only be given a weak recommendation. Thus it is strongly recommended that further studies assessing dietary supplementation validate these studies outlined below, and this task should be approached by using the recommended protocols for the diagnosis of EIB (also see the Diagnosis section),<sup>E2,E145</sup> which should also include recommendations for the assessment of pharmacotherapies for the treatment of EIB by regulatory agencies.<sup>E256</sup> If dietary supplements are to be prescribed, they should not be seen as an alternative to established pharmacotherapies.

In a series of studies in subjects with mild EIB, one group of investigators has comprehensively shown that a low-salt diet for 2 to 5 weeks inhibits EIB, whereas high-salt diets worsen EIB.<sup>E401,E404</sup> The same investigators have demonstrated that in elite athletes and asthmatic patients with EIB, a 3-week course of high-dose omega-3 fatty acid supplements inhibits the decrease in FEV<sub>1</sub> with exercise or EVH challenge and reduces associated

inflammatory markers.<sup>E46,E403,E405,E406</sup> These findings challenged the initial study assessing omega-3 supplements, which did not demonstrate inhibition on EIB with a similar high daily dose used for 3 weeks.<sup>E407</sup> This negative finding has been reproduced with inhaled mannitol<sup>E408</sup> and EVH<sup>E409</sup> by using similar dose and duration of treatment, leaving the role of omega-3 fatty acids in patients with EIB uncertain, and as such this is currently supportive of a weak recommendation. There also is weak evidence for vitamin C supplementation, leading to attenuation of EIB either acutely or after days or a few weeks of supplementation.<sup>E402,E410,E411</sup>

### Competitive and elite athletes

**Summary Statement 28:** Treat athletes with EIB alone in a similar manner to those with EIB and asthma by using the recommended general treatments for asthma. This might require additional consideration in athletes in whom some governing bodies might have requirements for obtaining permission to receive pharmaceutical agents for competition. [Strength of Recommendation: Strong; Evidence: A]

(<http://www.usada.org/substances/prohibited-list/athlete-guide/>)

Management of EIB in elite athletes is similar to that for recreational athletes and should include reducing relevant environmental exposures as much as possible; treatment of associated comorbid conditions; appropriate pharmacotherapy for control of symptoms, prophylaxis, and rescue; and patient education.<sup>E121,E412</sup> An individualized exercise prescription considering the athlete's venue might need to be designed by the athlete and the specialist to provide adequate control of EIB or EIB with asthma (eg, swimmers).

Similar to observations in asthmatic patients with EIB, as extensively outlined in this document, studies in athletes with EIB alone have shown the same results in the form of the acute protective effect of a  $\beta_2$ -agonist, the mast cell stabilizer cromoglycate, the leukotriene antagonist montelukast, and an inhibitory effect of high-dose ICSs when given acutely.<sup>E56,E57,E337,E413,E414</sup>

When considering treatment to control EIB, it is recommended that controller pharmacotherapy for athletes who have EIB with asthma should include daily ICSs.<sup>E2,E147</sup> Again, it should be noted that the combination of ICSs plus LABAs is not recommended because of the potential for tolerance to develop with daily use of  $\beta_2$ -agonists. This tolerance reduces the duration of bronchoprotection in exercise afforded by the  $\beta_2$ -agonist and prolongs recovery time with rescue bronchodilator.<sup>E309</sup> In some patients with concomitant moderate-to-severe persistent asthma, however, combination therapy can have added utility. The athlete's performance results should be monitored carefully because spirometry and symptoms alone might not be reliable end points to monitor asthma control. Because objective evidence of EIB is recommended by using indirect tests, it can be useful to repeat these tests after weeks to months on regular ICSs or to objectively assess the acute effect of a treatment to inhibit EIB and demonstrate this effect to the athlete, as has been demonstrated in airway hyperresponsiveness in patients with clinical asthma.<sup>E149,E352</sup>

Athletes engaged in either winter or summer sports with high ventilation rates (eg, swimming, mountain biking, rowing, biathlon, cross country skiing, and skating events) can have respiratory symptoms compatible with those with EIB alone, with or without demonstrating a positive challenge test result indicative of EIB or asthma. It has been proposed that the

repetitive epithelial injury repair cycle in response to breathing high volumes of unconditioned air over long periods can result in changes in the contractile properties of BSM as a result of exposure to plasma-derived products from exudation.<sup>E117,E415,E416</sup> This might be representative of an “airway injury” resulting in a form of overuse syndrome in contrast to EIB, which results from airway smooth muscle constriction from the osmotic release of bronchoconstrictive mediators from resident inflammatory cells (eg, mast cells and eosinophils). In the case of the winter athlete, it is common to see high prevalence of BHR to direct challenge tests, such as methacholine, and low prevalence of BHR to indirect tests.<sup>E417,E418</sup> If airway injury is suspected in an athlete, treatment recommendations can include the limitation of activity rather than introduction of the pharmacologic agents used in the treatment of asthma and EIB.<sup>E419,E420</sup> However, in the summer athlete with allergic sensitization, the conditioning of large volumes of air, which might contain higher levels of seasonal airborne allergen, could lead to airway inflammatory cell recruitment, as well the abovementioned consequences of plasma exudation, leading to passive sensitization of the BSM.<sup>E415</sup> In contrast to the winter athlete, summer athletes generally demonstrate higher rates of BHR to indirect tests and lower rates of BHR to direct tests,<sup>E137,E421</sup> which has led to suggestions that elite-level exercise in these environments in susceptible subjects can promote EIB.<sup>E422</sup>

## REFERENCES

- E1. Weiler JM, Anderson SD, Randolph C, Bonini S, Craig TJ, Pearlman DS, et al. Pathogenesis, prevalence, diagnosis, and management of exercise-induced bronchoconstriction: a practice parameter. *Ann Allergy Asthma Immunol* 2010;105(suppl):S1-47. (IV).
- E2. Parsons JP, Hallstrand TS, Mastronarde JG, Kaminsky DA, Rundell KW, Hull JH, et al. An official American Thoracic Society clinical practice guideline: exercise-induced bronchoconstriction. *Am J Respir Crit Care Med* 2013;187:1016-27. (IV).
- E3. Hallstrand TS, Lai Y, Henderson WR Jr, Altemeier WA, Gelb MH. Epithelial regulation of eicosanoid production in asthma. *Pulm Pharmacol Ther* 2012;25:432-7. (IV).
- E4. McFadden ER, Lenner KA, Strohl KP. Postexercise airway rewarming and thermally induced asthma. *J Clin Invest* 1986;78:18-25. (IIa).
- E5. McFadden ER, Pichurko BM. Intraairway thermal profiles during exercise and hyperventilation in normal man. *J Clin Invest* 1985;76:1007-10. (IIb).
- E6. Freed AN, Omori C, Schofield BH. The effect of bronchial blood flow on hyperpnea-induced airway obstruction and injury. *J Clin Invest* 1995;96:1221-9. (LB).
- E7. Anderson SD, Daviskas E. The airway microvasculature and exercise-induced asthma. *Thorax* 1992;47:748-52. (IV).
- E8. Anderson SD. Asthma provoked by exercise, hyperventilation, and the inhalation of non-isotonic aerosols. In: Barnes PJ, Rodger IW, Thomson NC, editors. *Asthma: basic mechanisms and clinical management*. 2nd ed. London: Academic Press; 1992. pp. 473-90. (IV).
- E9. Freed AN, Davis MS. Hyperventilation with dry air increases airway surface fluid osmolality in canine peripheral airways. *Am J Respir Crit Care Med* 1999;159:1101-7. (LB).
- E10. Belcher NG, Rees PJ, Clark TJ, Lee TH. A comparison of the refractory periods induced by hypertonic airway challenge and exercise in bronchial asthma. *Am Rev Respir Dis* 1987;135:822-5. (IIb).
- E11. Argyros GJ, Phillips YY, Rayburn DB, Rosenthal RR, Jaeger JJ. Water loss without heat flux in exercise-induced bronchospasm. *Am Rev Respir Dis* 1993;147:1419-24. (IIa).
- E12. Freed AN, Omori C, Hubbard WC, Adkinson NF. Dry air- and hypertonic aerosol-induced bronchoconstriction and cellular responses in the canine lung periphery. *Eur Respir J* 1994;7:1308-16. (Ib).
- E13. Brannan JD, Gulliksson M, Anderson SD, Chew N, Kumlin M. Evidence of mast cell activation and leukotriene release after mannitol inhalation. *Eur Respir J* 2003;22:491-6. (IIa).
- E14. Daviskas E, Gonda I, Anderson SD. Local airway heat and water vapour losses. *Respir Physiol* 1991;84:115-32. (IV).
- E15. Davis MS, Daviskas E, Anderson SD, Kotaru C, Hejal RB, Finigan JH, et al. Airway surface fluid desiccation during isocapnic hyperpnea. *J Appl Physiol* 2003;94:2545-7. (LB).
- E16. Tarran R. Regulation of airway surface liquid volume and mucus transport by active ion transport. *Proc Am Thorac Soc* 2004;1:42-6. (IV).
- E17. Daviskas E, Anderson SD, Gonda I, Chan HK, Cook P, Fulton R. Changes in mucociliary clearance during and after isocapnic hyperventilation in asthmatic and healthy subjects. *Eur Respir J* 1995;8:742-51. (IIb).
- E18. Gulliksson M, Palmberg L, Nilsson G, Ahlstedt S, Kumlin M. Release of prostaglandin D2 and leukotriene C in response to hyperosmolar stimulation of mast cells. *Allergy* 2006;61:1473-9. (LB).
- E19. Eggleston PA, Kagey-Sobotka A, Lichtenstein LM. A comparison of the osmotic activation of basophils and human lung mast cells. *Am Rev Respir Dis* 1987;135:1043-8. (LB).
- E20. Moloney ED, Griffin S, Burke CM, Poulter LW, O'Sullivan S. Release of inflammatory mediators from eosinophils following a hyperosmolar stimulus. *Respir Med* 2003;97:1-5. (LB).
- E21. Aitken ML, Marini JJ. Effect of heat delivery and extraction on airway conductance in normal and in asthmatic subjects. *Am Rev Respir Dis* 1985;131:357-61. (IIb).
- E22. Eschenbacher WL, Sheppard D. Respiratory heat loss is not the sole stimulus for bronchoconstriction induced by isocapnic hyperpnea with dry air. *Am Rev Respir Dis* 1985;131:894-901. (IIb).
- E23. Tabka Z, Ben Jebria A, Vergeret J, Guenard H. Effect of dry warm air on respiratory water loss in children with exercise-induced asthma. *Chest* 1988;94:81-6. (IIa).
- E24. Eliasson AH, Phillips YY, Rajagopal KR, Howard RS. Sensitivity and specificity of bronchial provocation testing. An evaluation of four techniques in exercise-induced bronchospasm. *Chest* 1992;102:347-55. (IIa).
- E25. Phillips YY, Jaeger JJ, Laube BL, Rosenthal RR. Eucapnic voluntary hyperventilation of compressed gas mixture. A simple system for bronchial challenge by respiratory heat loss. *Am Rev Respir Dis* 1985;131:31-5. (IIa).
- E26. Brummel NE, Mastronarde JG, Rittinger D, Philips G, Parsons JP. The clinical utility of eucapnic voluntary hyperventilation testing for the diagnosis of exercise-induced bronchospasm. *J Asthma* 2009;46:683-6. (III).
- E27. Dickinson J. Screening elite winter athletes for exercise induced asthma: a comparison of three challenge methods. *Br J Sports Med* 2006;40:179-82. (III).
- E28. Parsons JP, Kaeding C, Phillips GD, Jarjoura D, Wadley G, Mastronarde JG. Prevalence of exercise-induced bronchospasm in a cohort of varsity college athletes. *Med Sci Sports Exerc* 2007;39:1487-92. (III).
- E29. Stadelmann K, Stensrud T, Carlsen KH. Respiratory symptoms and bronchial responsiveness in competitive swimmers. *Med Sci Sports Exerc* 2011;43:375-81. (III).
- E30. Argyros GJ, Roach JM, Hurwitz KM, Eliasson AH, Phillips YY. Eucapnic voluntary hyperventilation as a bronchoprovocation technique. Development of a standardized dosing schedule in asthmatics. *Chest* 1996;109:1520-4. (III).
- E31. Carlsen KH, Anderson SD, Bjermer L, Bonini S, Brusasco V, Canonica W, et al. Exercise-induced asthma, respiratory and allergic disorders in elite athletes: epidemiology, mechanisms and diagnosis: part I of the report from the Joint Task Force of the European Respiratory Society (ERS) and the European Academy of Allergy and Clinical Immunology (EAACI) in cooperation with GA2-LEN. *Allergy* 2008;63:387-403. (IV).
- E32. Scollo M, Zanconato S, Ongaro R, Zaramella C, Zacchello F, Baraldi E. Exhaled nitric oxide and exercise-induced bronchoconstriction in asthmatic children. *Am J Respir Crit Care Med* 2000;161:1047-50. (III).
- E33. Malmberg LP, Pelkonen AS, Mattila PS, Hammaren-Malmi S, Makela MJ. Exhaled nitric oxide and exercise-induced bronchoconstriction in young wheezy children—interactions with atopy. *Pediatr Allergy Immunol* 2009;20:673-8. (III).
- E34. Hallstrand TS, Moody MW, Aitken ML, Henderson WR Jr. Airway immunopathology of asthma with exercise-induced bronchoconstriction. *J Allergy Clin Immunol* 2005;116:586-93. (IIa).
- E35. Carraro S, Corradi M, Zanconato S, Alinovi R, Pasquale MF, Zacchello F, et al. Exhaled breath condensate cysteinyl leukotrienes are increased in children with exercise-induced bronchoconstriction. *J Allergy Clin Immunol* 2005;115(4):764-70. (IIb).
- E36. Tahan F, Saraymen R, Gumus H. The role of lipoxin A4 in exercise-induced bronchoconstriction in asthma. *J Asthma* 2008;45:161-4. (IIb).
- E37. Melillo E, Woolley KL, Manning PJ, Watson RM, O'Byrne PM. Effect of inhaled PGE<sub>2</sub> on exercise-induced bronchoconstriction in asthmatic subjects. *Am J Respir Crit Care Med* 1994;149:1138-41. (Ib).
- E38. Hallstrand TS, Wurfel MM, Lai Y, Ni Z, Gelb MH, Altemeier WA, et al. Transglutaminase 2, a novel regulator of eicosanoid production in asthma revealed by

- genome-wide expression profiling of distinct asthma phenotypes. *PLoS One* 2010;5:e8583. (IIa).
- E39. Cai Y, Bjermer L, Halstensen TS. Bronchial mast cells are the dominating LTC4S-expressing cells in aspirin-tolerant asthma. *Am J Respir Cell Mol Biol* 2003;29:683-93. (IIa).
- E40. Duong M, Subbarao P, Adelroth E, Obminski G, Strinich T, Inman M, et al. Sputum eosinophils and the response of exercise-induced bronchoconstriction to corticosteroid in asthma. *Chest* 2008;133:404-11. (III).
- E41. Lai Y, Oslund RC, Bollinger JG, Henderson WR Jr, Santana LF, Altemeier WA, et al. Eosinophil cysteinyl leukotriene synthesis mediated by exogenous secreted phospholipase A2 group X. *J Biol Chem* 2010;285:41491-500. (LB).
- E42. Dougherty RH, Sidhu SS, Raman K, Solon M, Solberg OD, Caughey GH, et al. Accumulation of intraepithelial mast cells with a unique protease phenotype in T(H)2-high asthma. *J Allergy Clin Immunol* 2010;125:1046-53.e8. (IIa).
- E43. Woodruff PG, Boushey HA, Dolganov GM, Barker CS, Yang YH, Donnelly S, et al. Genome-wide profiling identifies epithelial cell genes associated with asthma and with treatment response to corticosteroids. *Proc Natl Acad Sci U S A* 2007;104:15858-63. (IIa).
- E44. Lai Y, Altemeier WA, Vandree J, Piliponsky AM, Johnson B, Appel CL, et al. Increased density of intraepithelial mast cells in patients with exercise-induced bronchoconstriction regulated through epithelially derived thymic stromal lymphopoietin and IL-33. *J Allergy Clin Immunol* 2014;133:448-55. (IIa).
- E45. Hallstrand TS, Moody MW, Wurfel MM, Schwartz LB, Henderson WR, Aitken ML. Inflammatory basis of exercise-induced bronchoconstriction. *Am J Respir Crit Care Med* 2005;172:679-86. (Ib).
- E46. Mickleborough TD, Murray RL, Ionescu AA, Lindley MR. Fish oil supplementation reduces severity of exercise-induced bronchoconstriction in elite athletes. *Am J Respir Crit Care Med* 2003;168:1181-9. (Ib).
- E47. Bikov A, Gajdosi R, Huszar E, Szili B, Lazar Z, Antus B, et al. Exercise increases exhaled breath condensate cysteinyl leukotriene concentration in asthmatic patients. *J Asthma* 2010;47:1057-62. (IIb).
- E48. O'Sullivan S, Roquet A, Dahlén B, Larsen F, Eklund A, Kumlin M, et al. Evidence for mast cell activation during exercise-induced bronchoconstriction. *Eur Respir J* 1998;12:345-50. (IIb).
- E49. Reiss TF, Hill JB, Harman E, Zhang J, Tanaka WK, Bronsky E, et al. Increased urinary excretion of LTE<sub>4</sub> after exercise and attenuation of exercise-induced bronchospasm by montelukast, a cysteinyl leukotriene receptor antagonist. *Thorax* 1997;52:1030-5. (Ib).
- E50. Hallstrand TS, Henderson WR Jr. An update on the role of leukotrienes in asthma. *Curr Opin Allergy Clin Immunol* 2010;10:60-6. (IV).
- E51. Hartley JPR, Charles TJ, Monie RDG, Seaton A, Taylor WH, Westood A, et al. Arterial plasma histamine after exercise in normal individuals and in patients with exercise induced asthma. *Clin Sci* 1981;61:151-7. (IIa).
- E52. Anderson SD, Bye PTP, Schoeffel RE, Seale JP, Taylor KM, Ferris L. Arterial plasma histamine levels at rest, during and after exercise in patients with asthma: effects of terbutaline aerosol. *Thorax* 1981;36:259-67. (IIa).
- E53. Haverkamp HC, Dempsey JA, Pegelow DF, Miller JD, Romer LM, Santana M, et al. Treatment of airway inflammation improves exercise pulmonary gas exchange and performance in asthmatic subjects. *J Allergy Clin Immunol* 2007;120:39-47. (Ib).
- E54. Anderson SD, Brannan JD. Exercise induced asthma: is there still a case for histamine? *J Allergy Clin Immunol* 2002;109:771-3. (IV).
- E55. Brannan JD, Anderson SD, Gomes K, King GG, Chan H-K, Seale JP. Fexofenadine decreases sensitivity to and montelukast improves recovery from inhaled mannitol. *Am J Respir Crit Care Med* 2001;163:1420-5. (Ib).
- E56. Kippelen P, Larsson J, Anderson SD, Brannan JD, Delin I, Dahlen B, et al. Acute effects of beclomethasone on hyperpnea-induced bronchoconstriction. *Med Sci Sports Exerc* 2010;42:273-80. (IIa).
- E57. Kippelen P, Larsson J, Anderson SD, Brannan JD, Dahlén B, Dahlén SE. Effect of sodium cromoglycate on mast cell mediators during hyperpnea in athletes. *Med Sci Sports Exerc* 2010;42:1853-60. (IIa).
- E58. Leff JA, Busse WW, Pearlman D, Bronsky EA, Kemp J, Hendeles L, et al. Montelukast, a leukotriene-receptor antagonist, for the treatment of mild asthma and exercise-induced bronchoconstriction. *N Engl J Med* 1998;339:147-52. (Ib).
- E59. Pearlman DS, van Adelsberg J, Philip G, Tilles SA, Busse W, Hendeles L, et al. Onset and duration of protection against exercise-induced bronchoconstriction by a single oral dose of montelukast. *Ann Allergy Asthma Immunol* 2006;97:98-104. (Ib).
- E60. Meltzer SS, Hasday JD, Cohn J, Bleeker ER. Inhibition of exercise-induced bronchospasm by zileuton: a 5-lipoxygenase inhibitor. *Am J Respir Crit Care Med* 1996;153:931-5. (Ib).
- E61. Baki A, Orhan F. The effect of loratadine in exercise-induced asthma. *Arch Dis Child* 2002;86:38-9. (Ib).
- E62. Dahlén B, Roquet A, Inman MD, Karlsson Ö, Naya I, Anstrén G, et al. Influence of zafirlukast and loratadine on exercise-induced bronchoconstriction. *J Allergy Clin Immunol* 2002;109:789-93. (Ib).
- E63. Patel KR. Terfenadine in exercise-induced asthma. *BMJ* 1984;85:1496-7. (Ib).
- E64. Van Rensen ELJ, Hiemstra PS, Rabe KF, Sterk PJ. Assessment of microvascular leakage via sputum induction. *Am J Respir Crit Care Med* 2002;165:1275-9. (IIb).
- E65. Kanazawa H, Asai K, Hirata K, Yoshikawa J. Vascular involvement in exercise-induced airway narrowing in patients with bronchial asthma. *Chest* 2002;122:166-70. (IIa).
- E66. Kanazawa H, Tochino Y, Asai K. Angiotensin-2 as a contributing factor of exercise-induced bronchoconstriction in asthmatic patients receiving inhaled corticosteroid therapy. *J Allergy Clin Immunol* 2008;121:390-5. (III).
- E67. Romberg K, LBJermer, Tufvesson E. Exercise but not mannitol provocation increases urinary Clara cell protein (CC16) in elite swimmers. *Respir Med* 2011;105:31.e6. (IIb).
- E68. Carbonnelle S, Francaux M, Doyle I, Dumont X, de Burbure C, Morel G, et al. Changes in serum pneumoproteins caused by short-term exposures to nitrogen trichloride in indoor chlorinated swimming pools. *Biomarkers* 2002;7:464e78. (IIb).
- E69. Kippelen P, Tufvesson E, Ali L, Bjermer L, Anderson SD. Urinary CC16 after challenge with dry air hyperpnea and mannitol in recreational summer athletes. *Respir Med* 2013;107:1837-44. (IIb).
- E70. Taylor-Clark TE, Nassenstein C, Udem BJ. Leukotriene D4 increases the excitability of capsaicin-sensitive nasal sensory nerves to electrical and chemical stimuli. *Br J Pharmacol* 2008;154:1359-68. (LB).
- E71. Freed AN, McCulloch S, Meyers T, Suzuki R. Neurokinins modulate hyperventilation-induced bronchoconstriction in canine peripheral airways. *Am J Respir Crit Care Med* 2003;167:1102-8. (LB).
- E72. Lai YL, Lee SP. Mediators in hyperpnea-induced bronchoconstriction of guinea pigs. *Naunyn Schmiedebergs Arch Pharmacol* 1999;360:597-602. (LB).
- E73. Fahy JV, Wong HH, Geppetti P, Reis JM, Harris SC, Maclean DB, et al. Effect of an NK1 receptor antagonist (CP-99,994) on hypertonic saline-induced bronchoconstriction and cough in male asthmatic subjects. *Am J Respir Crit Care Med* 1995;152:879-84. (Ib).
- E74. Ichinose M, Miura M, Yamauchi H, Kageyama N, Tomaki M, Oyake T, et al. A neurokinin 1-receptor antagonist improves exercise-induced airway narrowing in asthmatic patients. *Am J Respir Crit Care Med* 1996;153:936-41. (Ib).
- E75. Naline E, Devillier P, Drapeau G, Toty L, Bakdach H, Regoli D, et al. Characterization of neurokinin effects and receptor selectivity in human isolated bronchi. *Am Rev Respir Dis* 1989;140:679-86. (Ib).
- E76. Hallstrand TS, Debley JS, Farin FM, Henderson WR Jr. Role of MUC5AC in the pathogenesis of exercise-induced bronchoconstriction. *J Allergy Clin Immunol* 2007;119:1092-8. (IIa).
- E77. Mickleborough TD, Lindley MR, Turner LA. Comparative effects of a high-intensity interval warm-up and salbutamol on the bronchoconstrictor response to exercise in asthmatic athletes. *Int J Sports Med* 2007;28:456-62. (Ib).
- E78. Haverkamp HC, Dempsey JA, Miller JD, Romer LM, Pegelow DF, Lovering AT, et al. Repeat exercise normalizes the gas-exchange impairment induced by a previous exercise bout in asthmatic subjects. *J Appl Physiol* 2005;99:1843-52. (IIb).
- E79. Edmunds A, Tooley M, Godfrey S. The refractory period after exercise-induced asthma: its duration and relation to the severity of exercise. *Am Rev Respir Dis* 1978;117:247-54. (IIb).
- E80. Suh DI, Lee JK, Kim JT, Koh YY. Airway refractoriness to inhaled mannitol after repeated challenge. *Pediatr Pulmonol* 2011;46:1007-14. (IIb).
- E81. Larsson J, Perry CP, Anderson SD, Brannan JD, Dahlen SE, Dahlen B. The occurrence of refractoriness and mast cell mediator release following mannitol-induced bronchoconstriction. *J Appl Physiol* (1985) 2011;110:1029-35. (IIa).
- E82. Manning PJ, Watson RM, O'Byrne PM. Exercise-induced refractoriness in asthmatic subjects involves leukotriene and prostaglandin interdependent mechanisms. *Am Rev Respir Dis* 1993;148:950-4. (Ib).
- E83. Wilson BA, Bar-Or O, O'Byrne PM. The effects of indomethacin on refractoriness following exercise both with and without bronchoconstriction. *Eur Respir J* 1994;12:2174-8. (II).
- E84. Rundell KW, Im J, Mayers LB, Wilber RL, Szmedra L, Schmitz HR. Self-reported symptoms and exercise-induced asthma in the elite athlete. *Med Sci Sports Exerc* 2001;33:208-13. (III).
- E85. Weiler JM, Bonini S, Coifman R, Craig T, Delgado L, Capao-Filipe M, et al. American Academy of Allergy, Asthma & Immunology Work Group report: exercise-induced asthma. *J Allergy Clin Immunol* 2007;119:1349-58. (IV).



- E86. Hallstrand TS, Curtis JR, Koepsell TD, Martin DP, Schoene RB, Sullivan SD, et al. Effectiveness of screening examinations to detect unrecognized exercise-induced bronchoconstriction. *J Pediatr* 2002;141:343-9. (IV).
- E87. Mountjoy M, Fitch K, Boulet LP, Bougault V, van Mechelen W, Verhagen E. Prevalence and characteristics of asthma in the aquatic disciplines. *J Allergy Clin Immunol* 2015;136:588-94. (III).
- E88. Rundell KW, Anderson SD, Sue-Chu M, Bougault V, Boulet LP. Air quality and temperature effects on exercise-induced bronchoconstriction. *Compr Physiol* 2015;5:579-610. (IV).
- E89. EPR-3. Guidelines for the diagnosis and management of asthma—summary report 2007. *J Allergy Clin Immunol* 2007;120(suppl):S94-138. (IV).
- E90. Sano F, Sole D, Naspitz CK. Prevalence and characteristics of exercise-induced asthma in children. *Pediatr Allergy Immunol* 1998;9:181-5. (IIa).
- E91. Cabral ALB, Conceição GM, Fonseca-Guedes CHF, Martins MA. Exercise-induced bronchospasm in children. *Am J Respir Crit Care Med* 1999;159:1819-23. (IIb).
- E92. Randolph C. Pediatric exercise-induced bronchoconstriction: contemporary developments in epidemiology, pathogenesis, presentation, diagnosis, and therapy. *Curr Allergy Asthma Rep* 2013;13:662-71. (IV).
- E93. De Baets F, Bodart E, Dramaix-Wilmet M, Van Daele S, de Bildering G, Masset S, et al. Exercise-induced respiratory symptoms are poor predictors of bronchoconstriction. *Pediatr Pulmonol* 2005;39:301-5. (IIb).
- E94. Mannix ET, Roberts M, Fagin DP, Reid B, Farber MO. The prevalence of airways hyperresponsiveness in members of an exercise training facility. *J Asthma* 2003;40:349-55. (IIb).
- E95. Molphy J, Dickinson J, Hu J, Chester N, Whyte G. Prevalence of bronchoconstriction induced by eucapnic voluntary hyperpnea in recreationally active individuals. *J Asthma* 2014;51:44-50. (IV).
- E96. Godfrey S, König P. Exercise-induced bronchial lability in wheezy children and their families. *Pediatrics* 1975;56(Suppl):851-5. (IIa).
- E97. Helenius IJ, Tikkanen HO, Haahtela T. Occurrence of exercise induced bronchospasm in elite runners: dependence on atopy and exposure to cold air and pollen. *Br J Sports Med* 1998;32:125-9. (III).
- E98. Sallaoui R, Chamari K, Mossa A, Tabka Z, Chtara M, Feki Y, et al. Exercise-induced bronchoconstriction and atopy in Tunisian athletes. *BMC Pulm Med* 2009;9:8. (IIa).
- E99. Brutsche M, Britschgi D, Dayer E, Tschopp JM. Exercise-induced bronchospasm (EIB) in relation to seasonal and perennial specific IgE in young adults. *Allergy* 1995;50:905-9. (IIa).
- E100. Tilles SA. Vocal cord dysfunction in children and adolescents. *Curr Allergy Asthma Rep* 2003;3:467-72. (IV).
- E101. Schatz M, Clark S, Emond JA, Schreiber D, Camargo CA Jr. Sex differences among children 2-13 years of age presenting at the emergency department with acute asthma. *Pediatr Pulmonol* 2004;37:523-9. (III).
- E102. Lin RY, Lee GB. The gender disparity in adult asthma hospitalizations dynamically relates to age. *J Asthma* 2008;45:931-5. (III).
- E103. Bardagi S, Agudo A, Gonzalez CA, Romero PV. Prevalence of exercise-induced airway narrowing in schoolchildren from a Mediterranean town. *Am Rev Respir Dis* 1993;147:1112-5. (IIb).
- E104. Wilber RL, Rundell L, Szmedra L, Jenkinson DM, Im J, Drake SD. Incidence of exercise-induced bronchospasm in Olympic Winter Sport athletes. *Med Sci Sports Exerc* 2000;32:732-7. (IIb).
- E105. Couillard S, Bougault V, Turmel J, Boulet LP. Perception of bronchoconstriction following methacholine and eucapnic voluntary hyperpnea challenges in elite athletes. *Chest* 2014;145:794-802. (III).
- E106. Langdeau JB, Day A, Turcotte H, Boulet LP. Gender differences in the prevalence of airway hyperresponsiveness and asthma in athletes. *Respir Med* 2009;103:401-6. (III).
- E107. Kukafka DS, Lang DM, Porter S, Rogers J, Ciccolella D, Polansky M, et al. Exercise-induced bronchospasm in high school athletes via a free running test: incidence and epidemiology. *Chest* 1998;114:1613-22. (IIa).
- E108. Jones CO, Qureshi S, Rona RJ, Chinn S. Exercise-induced bronchoconstriction by ethnicity and presence of asthma in British nine year olds. *Thorax* 1996;51:1134-6. (III).
- E109. Weiler JM, Ryan EJ 3rd. Asthma in United States Olympic athletes who participated in the 1998 Olympic Winter Games. *J Allergy Clin Immunol* 2000;106:267-71. (III).
- E110. Weiler JM, Layton T, Hunt M. Asthma in United States Olympic athletes who participated in the 1996 Summer Games. *J Allergy Clin Immunol* 1998;102:722-6. (III).
- E111. Parsons JP, Mastrorade JG. Exercise-induced bronchoconstriction in athletes. *Chest* 2005;128:3966-74. (IV).
- E112. Mannix ET, Farber MO, Palange P, Galassetti P, Manfredi F. Exercise-induced asthma in figure skaters. *Chest* 1996;109:312-5. (IIb).
- E113. Rundell KW. High levels of airborne ultrafine and fine particulate matter in indoor ice arenas. *Inhal Toxicol* 2003;15:237-50. (III).
- E114. Rundell KW, Spiering BA, Evans TM, Baumann JM. Baseline lung function, exercise-induced bronchoconstriction, and asthma-like symptoms in elite women ice hockey players. *Med Sci Sports Exerc* 2004;36:405-10. (III).
- E115. Fitch KD, Morton AR. Specificity of exercise in exercise-induced asthma. *BMJ* 1971;4:577-81. (IIa).
- E116. Rundell KW, Wilber RL, Szmedra L, Jenkinson DM, Mayers LB, Im J. Exercise-induced asthma screening of elite athletes: field vs laboratory exercise challenge. *Med Sci Sports Exerc* 2000;32:309-16. (III).
- E117. Sue-Chu M, Larsson L, Moen T, Rennard SI, Bjermer L. Bronchoscopy and bronchoalveolar lavage findings in cross-country skiers with and without "ski asthma." *Eur Respir J* 1999;13:626-32. (IIa).
- E118. Sue-Chu M, Henriksen AH, Bjermer L. Non-invasive evaluation of lower airway inflammation in hyper-responsive elite cross-country skiers and asthmatics. *Respir Med* 1999;93:719-25.
- E119. Pohjantahti H, Laitinen J, Parkkari J. Exercise-induced bronchospasm among healthy elite cross country skiers and non-athletic students. *Scand J Med Sci Sports* 2005;15:324-8. (IIa).
- E120. Anderson SD, Fitch K, Perry CP, Sue-Chu M, Crapo R, McKenzie D, et al. Responses to bronchial challenge submitted for approval to use inhaled beta2 agonists prior to an event at the 2002 Winter Olympics. *J Allergy Clin Immunol* 2003;111:44-9. (III).
- E121. Fitch KD, Sue-Chu M, Anderson SD, Boulet LP, Hancox RJ, McKenzie DC, et al. Asthma and the elite athlete: summary of the International Olympic Committee's consensus conference, Lausanne, Switzerland, January 22-24, 2008. *J Allergy Clin Immunol* 2008;122:254-60. e1-7 (IV).
- E122. Randolph CC, Dreyfus D, Rundell KW, Bangladore D, Fraser B. Prevalence of allergy and asthma symptoms in recreational roadrunners. *Med Sci Sports Exerc* 2006;38:2053-7. (III).
- E123. Rundell KW, Spiering BA, Judelson DA, Wilson MH. Bronchoconstriction during cross-country skiing: is there really a refractory period? *Med Sci Sports Exerc* 2003;35:18-26. (IV).
- E124. Larsson K, Ohlsén P, Malmberg P, Rydström P-O, Ulriksen H. High prevalence of asthma in cross country skiers. *BMJ* 1993;307:1326-9. (IIa).
- E125. Helenius IJ, Tikkanen HO, Haahtela T. Association between type of training and risk of asthma in elite athletes. *Thorax* 1997;52:157-60. (III).
- E126. Helenius I, Haahtela T. Allergy and asthma in elite summer sport athletes. *J Allergy Clin Immunol* 2000;106:444-52. (IV).
- E127. Bernard A, Nickmilder M, Voisin C, Sardella A. Impact of chlorinated swimming pool attendance on the respiratory health of adolescents. *Pediatrics* 2009;124:1110-8. (IIa).
- E128. Helenius I, Ryttilä P, Sarna S, Lumme A, Helenius M, Remes V, et al. Effect of continuing or finishing high-level sports on airway inflammation, bronchial hyperresponsiveness, and asthma: a 5-year prospective follow-up study of 42 highly trained swimmers. *J Allergy Clin Immunol* 2002;109:962-8. (IIb).
- E129. Rundell KW, Caviston R. Ultrafine and fine particulate matter inhalation decreases exercise performance in healthy subjects. *J Strength Cond Res* 2008;22:2-5. (IIa).
- E130. Rundell KW, Hoffman JR, Caviston R, Bulbulian R, Hollenbach AM. Inhalation of ultrafine and fine particulate matter disrupts systemic vascular function. *Inhal Toxicol* 2007;19:133-40. (IIa).
- E131. Rundell KW, Caviston R, Hollenbach AM, Murphy K. Vehicular air pollution, playgrounds, and youth athletic fields. *Inhal Toxicol* 2006;18:541-7. (IIb).
- E132. McCreanor J, Cullinan P, Nieuwenhuijsen MJ, Stewart-Evans J, Malliarou E, Jarup L, et al. Respiratory effects of exposure to diesel traffic in persons with asthma. *N Engl J Med* 2007;357:2348-58. (IIb).
- E133. Choi IS, Ki WJ, Kim TO, Han ER, Seo IK. Seasonal factors influencing exercise-induced asthma. *Allergy Asthma Immunol Res* 2012;4:192-8. (III).
- E134. Goldberg S, Mimouni F, Joseph L, Izbicki G, Picard E. Seasonal effect on exercise challenge tests for the diagnosis of exercise-induced bronchoconstriction. *Allergy Asthma Proc* 2012;33:416-20. (III).
- E135. Ucock K, Dane S, Gokbel H, Akar S. Prevalence of exercise-induced bronchospasm in long distance runners trained in cold weather. *Lung* 2004;182:265-70. (IIa).
- E136. Moorman JE, Akinbami LJ, Bailey CM, Zahran HS, King ME, Johnson CA, et al. National surveillance of asthma: United States, 2001-2010. *Vital Health Stat* 3 2012(35):1-67. (IV).
- E137. Holzer K, Anderson SD, Douglass J. Exercise in elite summer athletes: challenges for diagnosis. *J Allergy Clin Immunol* 2002;110:374-80. (IV).
- E138. Weiler JM, Hallstrand TS, Parsons JP, Randolph C, Silvers WS, Storms WW, et al. Improving screening and diagnosis of exercise-induced bronchoconstriction: a call to action. *J Allergy Clin Immunol Pract* 2014;2:275-80.e7. (IV).



- E139. Carlsen KH, Engh G, Mørk M. Exercise induced bronchoconstriction depends on exercise load. *Respir Med* 2000;94:750-5. (III).
- E140. Weinberger M, Abu-Hasan M. Perceptions and pathophysiology of dyspnea and exercise intolerance. *Pediatr Clin North Am* 2009;56:33-48. ix, (IV).
- E141. Anderson SD, Pearlman DS, Rundell KW, Perry CP, Boushey H, Sorkness CA, et al. Reproducibility of the airway response to an exercise protocol standardized for intensity, duration, and inspired air conditions, in subjects with symptoms suggestive of asthma. *Respir Res* 2010;11:120. (Ib).
- E142. van Leeuwen JC, Driessen JM, Kersten ET, Thio BJ. Assessment of exercise-induced bronchoconstriction in adolescents and young children. *Immunol Allergy Clin North Am* 2013;33:381-94. viii-ix (IV).
- E143. Simpson AJ, Romer LM, Kippelen P. Self-reported symptoms after induced and inhibited bronchoconstriction in athletes. *Med Sci Sports Exerc* 2015;47:2005-13. (Ib).
- E144. Rundell KW, Slee JB. Exercise and other indirect challenges to demonstrate asthma or exercise-induced bronchoconstriction in athletes. *J Allergy Clin Immunol* 2008;122:238-48. (IV).
- E145. Crapo RO, Casaburi R, Coates AL, Enright PL, Hankinson JL, Irvin CG, et al. Guidelines for methacholine and exercise challenge testing—1999. *Am J Respir Crit Care Med* 2000;161:309-29. (IV).
- E146. Cockcroft D, Davis B. Direct and indirect challenges in the clinical assessment of asthma. *Ann Allergy Asthma Immunol* 2009;103:363-72. 400 (IV).
- E147. Koh MS, Tee A, Lasserson TJ, Irving LB. Inhaled corticosteroids compared to placebo for prevention of exercise induced bronchoconstriction. *Cochrane Database Syst Rev* 2007(3):CD002739. (Ia).
- E148. Subbarao P, Duong M, Adelroth E, Otis J, Obminski G, Inman M, et al. Effect of ciclesonide dose and duration of therapy on exercise-induced bronchoconstriction in patients with asthma. *J Allergy Clin Immunol* 2006;117:1008-13. (Ib).
- E149. Lipworth BJ, Short PM, Williamson PA, Clearie KL, Fardon TC, Jackson CM. A randomized primary care trial of steroid titration against mannitol in persistent asthma: STAMINA trial. *Chest* 2012;141:607-15. (Ib).
- E150. Anderson SD, Charlton B, Weiler JM, Nichols S, Spector SL, Pearlman DS. Comparison of mannitol and methacholine to predict exercise-induced bronchoconstriction and a clinical diagnosis of asthma. *Respir Res* 2009;10:4. (Ib).
- E151. Holley AB, Cohee B, Walter RJ, Shah AA, King CS, Roop S. Eucapnic voluntary hyperventilation is superior to methacholine challenge testing for detecting airway hyperreactivity in nonathletes. *J Asthma* 2012;49:614-9. (III).
- E152. Anderson SD, Lambert S, Brannan JD, Wood RJ, Koskela H, Morton AR, et al. Laboratory protocol for exercise asthma to evaluate salbutamol given by two devices. *Med Sci Sports Exerc* 2001;33:893-900. (Ib).
- E153. Weiler JM, Nathan RA, Rupp NT, Kalberg CJ, Emmett A, Dorinsky PM. Effect of fluticasone/salmeterol administered via a single device on exercise-induced bronchospasm in patients with persistent asthma. *Ann Allergy Asthma Immunol* 2005;94:65-72. (Ib).
- E154. Anderson SD, Kippelen P. Assessment of EIB: What you need to know to optimize test results. *Immunol Allergy Clin North Am* 2013;33:363-80. viii. (IV).
- E155. Tanaka H, Monahan KD, Seals DR. Age-predicted maximal heart rate revisited. *J Am Coll Cardiol* 2001;37:153-6.
- E156. Price OJ, Ansley L, Hull JH. Diagnosing exercise-induced bronchoconstriction with eucapnic voluntary hyperpnea: is one test enough? *J Allergy Clin Immunol Pract* 2015;3:243-9. (III).
- E157. Chateaubriand do Nascimento Silva Filho MJ, Goncalves AV, Tavares Viana M, Peixoto DM, Cavalcanti Sarinho ES, Rizzo JA. Exercise-induced bronchoconstriction diagnosis in asthmatic children: comparison of treadmill running and eucapnic voluntary hyperventilation challenges. *Ann Allergy Asthma Immunol* 2015;115:277-81. (III).
- E158. Newnham DM, Ingram CG, Earnshaw J, Palmer JBD, Dhillon DP. Salmeterol provides prolonged protection against exercise-induced bronchoconstriction in a majority of subjects with mild, stable asthma. *Respir Med* 1993;87:439-44. (Ib).
- E159. Gröneröd TA, Von Berg A, Schwabe G, Soliman S. Formoterol via Turbuhaler® gave better protection than terbutaline against repeated exercise challenge for up to 12 hours in children and adolescents. *Respir Med* 2000;94:661-7. (Ib).
- E160. Coreno A, Skowronski M, Kotaur C, McFadden ER. Comparative effects of long-acting  $\beta_2$ -agonists, leukotriene antagonists, and a 5-lipoxygenase inhibitor on exercise-induced asthma. *J Allergy Clin Immunol* 2000;106:500-6. (Ib).
- E161. Pearlman D, Milgrom H, Till D, Ziehm B. Effect of formoterol fumarate treatment on exercise-induced bronchoconstriction in children. *Ann Allergy Asthma Immunol* 2006;97:382-8. (Ib).
- E162. Peroni DG, Piacentini GL, Pietrobelli A, Loiacono A, De Gasperi W, Sabbion A, et al. The combination of single-dose montelukast and loratadine on exercise-induced bronchospasm in children. *Eur Respir J* 2002;20:104-7. (Ib).
- E163. Smith CM, Anderson SD, Seale JP. The duration of action of the combination of fenoterol hydrobromide and ipratropium bromide in protecting against asthma provoked by hyperpnea. *Chest* 1988;94:709-17. (Ib).
- E164. Woolley M, Anderson SD, Quigley B. Duration of protective effect of terbutaline sulphate and cromolyn sodium alone and in combination on exercise-induced asthma. *Chest* 1990;97:39-45. (Ib).
- E165. König P, Hordvik NL, Kreutz C. The preventative effect and duration of action of nedocromil sodium and cromolyn sodium on exercise-induced asthma (EIA) in adults. *J Allergy Clin Immunol* 1987;79:64-8. (Ib).
- E166. Haney S, Hancox RJ. Recovery from bronchoconstriction and bronchodilator tolerance. *Clin Rev Allergy Immunol* 2006;31:181-96. (IV).
- E167. Davis BE, Reid JK, Cockcroft DW. Formoterol thrice weekly does not result in the development of tolerance to bronchoprotection. *Can Respir J* 2003;10:23-6. (Ib).
- E168. O'Byrne PM, van der Linde J, Cockcroft DW, Gauvreau GM, Brannan JD, Fitzgerald M, et al. Prolonged bronchoprotection against inhaled methacholine by inhaled BI 1744, a long-acting beta(2)-agonist, in patients with mild asthma. *J Allergy Clin Immunol* 2009;124:1217-21. (Ib).
- E169. Visser R, Wind M, de Graaf B, de Jongh FH, van der Palen J, Thio BJ. Protective effect of a low single dose inhaled steroid against exercise induced bronchoconstriction. *Pediatr Pulmonol* 2015;50:1178-83. (IIa).
- E170. VanHaitisma TA, Mickleborough T, Stager JM, Kocaja DM, Lindley MR, Chapman R. Comparative effects of caffeine and albuterol on the bronchoconstrictor response to exercise in asthmatic athletes. *Int J Sports Med* 2010;31:231-6. (Ib).
- E171. Nielsen EW, Hull JH, Backer V. High prevalence of exercise-induced laryngeal obstruction in athletes. *Med Sci Sports Exerc* 2013;45:2030-5. (III).
- E172. Rundell KW, Weiss P. Exercise-induced bronchoconstriction and vocal cord dysfunction: two sides of the same coin? *Curr Sports Med Rep* 2013;12:41-6. (IV).
- E173. Christopher KL, Wood RP 2nd, Eckert RC, Blager FB, Raney RA, Souhrada JF. Vocal-cord dysfunction presenting as asthma. *N Engl J Med* 1983;308:1566-70. (Ib).
- E174. Björnsdóttir US, Gudmundsson K, Hjartarson H, Brøndbo K, Magnusson B, Juliusson S. Exercise-induced laryngomalacia: an imitator of exercise-induced bronchospasm. *Ann Allergy Asthma Immunol* 2000;85:387-91. (III).
- E175. Chemery L, Le Clech G, Delaval P, Carre F, Gogibu J, Dassonville J. Exercise-induced laryngomalacia. *Rev Mal Respir* 2002;19:641-3. (III).
- E176. Bittleman DB, Smith RJ, Weiler JM. Abnormal movement of the arytenoid region during exercise presenting as exercise-induced asthma in an adolescent athlete. *Chest* 1994;106:615-6. (III).
- E177. Rundell K, Spiering BA. Inspiratory stridor in elite athletes. *Chest* 2003;123:468-74. (III).
- E178. Hanks CD, Parsons J, Benninger C, Kaeding C, Best TM, Phillips G, et al. Etiology of dyspnea in elite and recreational athletes. *Phys Sportsmed* 2012;40:28-33. (III).
- E179. Tilles SA, Ayars AG, Picciano JF, Altman K. Exercise-induced vocal cord dysfunction and exercise-induced laryngomalacia in children and adolescents: the same clinical syndrome? *Ann Allergy Asthma Immunol* 2013;111:342-6.e1. (III).
- E180. McFadden ER, Zawadski DK. Vocal cord dysfunction masquerading as exercise-induced asthma. A physiologic cause for "choking" during athletic activities. *Am J Respir Crit Care Med* 1996;153:942-7. (Ib).
- E181. Morris MJ, Deal LE, Bean DR, Grbach VX, Morgan JA. Vocal cord dysfunction in patients with exertional dyspnea. *Chest* 1999;116:1676-82. (IIa).
- E182. Perkins PJ, Morris MJ. Vocal cord dysfunction induced by methacholine challenge testing. *Chest* 2002;122:1988-93. (IIa).
- E183. Davis RS, Brugman SM, Larsen GL. Use of videography in the diagnosis of exercise-induced vocal cord dysfunction: a case report with video clips. *J Allergy Clin Immunol* 2007;119:1329-31. (III).
- E184. Heimdal JH, Roksund OD, Halvorsen T, Skadberg BT, Olofsson J. Continuous laryngoscopy exercise test: a method for visualizing laryngeal dysfunction during exercise. *Laryngoscope* 2006;116:52-7. (III).
- E185. Fahey JT, Bryant NJ, Karas D, Goldberg B, Destefano R, Gracco LC. Exercise-induced stridor due to abnormal movement of the arytenoid area: videoendoscopic diagnosis and characterization of the "at risk" group. *Pediatr Pulmonol* 2005;39:51-5. (III).
- E186. Richter GT, Rutter MJ, deAlarcon A, Orvidas LJ, Thompson DM. Late-onset laryngomalacia: a variant of disease. *Arch Otolaryngol Head Neck Surg* 2008;134:75-80. (Ib).

- E187. Bent JP 3rd, Miller DA, Kim JW, Bauman NM, Wilson JS, Smith RJ. Pediatric exercise-induced laryngomalacia. *Ann Otol Rhinol Laryngol* 1996; 105:169-75. (III).
- E188. Lakin RC, Metzger WJ, Haughey BH. Upper airway obstruction presenting as exercise-induced asthma. *Chest* 1984;86:499-501. (III).
- E189. Mandell DL, Arjmand EM. Laryngomalacia induced by exercise in a pediatric patient. *Int J Pediatr Otorhinolaryngol* 2003;67:999-1003. (III).
- E190. Smith RJ, Bauman NM, Bent JP, Kramer M, Smits WL, Ahrens RC. Exercise-induced laryngomalacia. *Ann Otol Rhinol Laryngol* 1995;104: 537-41. (IV).
- E191. Gessler EM, Simko EJ, Greinwald JH Jr. Adult laryngomalacia: an uncommon clinical entity. *Am J Otolaryngol* 2002;23:386-9. (III).
- E192. Patel NJ, Jorgensen C, Kuhn J, Merati AL. Concurrent laryngeal abnormalities in patients with paradoxical vocal fold dysfunction. *Otolaryngol Head Neck Surg* 2004;130:686-9. (IIb).
- E193. Powell DM, Karanfilov BI, Beechler KB, Treole K, Trudeau MD, Forrest LA. Paradoxical vocal cord dysfunction in juveniles. *Arch Otolaryngol Head Neck Surg* 2000;126:29-34. (IIb).
- E194. Ford CN. Evaluation and management of laryngopharyngeal reflux. *JAMA* 2005;294:1534-40. (IV).
- E195. Balkissoon R, Kenn K. Asthma: vocal cord dysfunction (VCD) and other dysfunctional breathing disorders. *Semin Respir Crit Care Med* 2012;33: 595-605. (IV).
- E196. Tilles SA. Exercise-induced respiratory symptoms: an epidemic among adolescents. *Ann Allergy Asthma Immunol* 2010;104:361-70. 412 (IV).
- E197. Alshati M, Cockcroft DW, Fenton ME. Exercise-induced hyperventilation: more common than appreciated. *Ann Allergy Asthma Immunol* 2012;109: 282-4. (IV).
- E198. Buckner K. Cardiac asthma. *Immunol Allergy Clin North Am* 2013;33: 35-44. (IV).
- E199. Abu-Hasan M, Tannous B, Weinberger M. Exercise-induced dyspnea in children and adolescents: if not asthma then what? *Ann Allergy Asthma Immunol* 2005;94:366-71. (IIb).
- E200. Hammo AH, Weinberger MM. Exercise-induced hyperventilation: a pseudoasthma syndrome. *Ann Allergy Asthma Immunol* 1999;82:574-8. (III).
- E201. Weinberger MM. Etiology of exercise-induced dyspnea: not just exercise-induced asthma or vocal cord dysfunction. *J Allergy Clin Immunol* 2008;121: 269. (IV).
- E202. Weinberger M, Abu-Hasan M. Pseudo-asthma: when cough, wheezing, and dyspnea are not asthma. *Pediatrics* 2007;120:855-64. (IV).
- E203. Weinberger M. Exercise induced dyspnoea: if not asthma, then what? *Arch Dis Child* 2006;91:543-4. (IV).
- E204. Meyer T, Faude O, Scharhag J, Urhausen A, Kindermann W. Is lactic acidosis a cause of exercise induced hyperventilation at the respiratory compensation point? *Br J Sports Med* 2004;38:622-5. (III).
- E205. White KM, Quinn JM, Hagan LL, Johnson TL 2nd. Exercise-induced hyperventilation. *Ann Allergy Asthma Immunol* 2008;100:171-2. (III).
- E206. Jack S, Rossiter HB, Pearson MG, Ward SA, Warburton CJ, Whipp BJ. Ventilatory responses to inhaled carbon dioxide, hypoxia, and exercise in idiopathic hyperventilation. *Am J Respir Crit Care Med* 2004;170:118-25. (IIa).
- E207. Dempsey JA, McKenzie DC, Haverkamp HC, Eldridge MW. Update in the understanding of respiratory limitations to exercise performance in fit, active adults. *Chest* 2008;134:613-22. (IV).
- E208. Babb TG, Ranasinghe KG, Comeau LA, Semon TL, Schwartz B. Dyspnea on exertion in obese women: association with an increased oxygen cost of breathing. *Am J Respir Crit Care Med* 2008;178:116-23. (IIb).
- E209. Davies CT, Godfrey S, Light M, Sargeant AJ, Zeidifard E. Cardiopulmonary responses to exercise in obese girls and young women. *J Appl Physiol* 1975;38: 373-6. (III).
- E210. Lopes WA, Radominski RB, Rosario Filho NA, Leite N. Exercise-induced bronchospasm in obese adolescents. *Allergol Immunopathol (Madr)* 2009;37: 175-9. (IIa).
- E211. Koumbourlis AC, Stolar CJ. Lung growth and function in children and adolescents with idiopathic pectus excavatum. *Pediatr Pulmonol* 2004;38: 339-43. (III).
- E212. Fonkalsrud EW, DeUgarte D, Choi E. Repair of pectus excavatum and carinatum deformities in 116 adults. *Ann Surg* 2002;236:304-14. (III).
- E213. Smyth RJ, Chapman KR, Wright TA, Crawford JS, Rebeck AS. Ventilatory patterns during hypoxia, hypercapnia, and exercise in adolescents with mild scoliosis. *Pediatrics* 1986;77:692-7. (III).
- E214. Ben-Dov I, Kaminski N, Reichert N, Rosenman J, Shulimzon T. Diaphragmatic paralysis: a clinical imitator of cardiorespiratory diseases. *Isr Med Assoc J* 2008;10:579-83. (III).
- E215. American Thoracic Society/European Respiratory Society. ATS/ERS Statement on respiratory muscle testing. *Am J Respir Crit Care Med* 2002; 166:518-624. (IV).
- E216. Marciniuk DD, Sridhar G, Clemens RE, Zintel TA, Gallagher CG. Lung volumes and expiratory flow limitation during exercise in interstitial lung disease. *J Appl Physiol* (1985) 1994;77:963-73. (III).
- E217. Harris-Eze AO, Sridhar G, Clemens RE, Zintel TA, Gallagher CG, Marciniuk DD. Role of hypoxemia and pulmonary mechanics in exercise limitation in interstitial lung disease. *Am J Respir Crit Care Med* 1996;154: 994-1001. (III).
- E218. Sheffer AL, Austen KF. Exercise-induced anaphylaxis. *J Allergy Clin Immunol* 1984;73:699-703. (IV).
- E219. Maulitz RM, Pratt DS, Schocket AL. Exercise-induced anaphylactic reaction to shellfish. *J Allergy Clin Immunol* 1979;63:433-4. (III).
- E220. Kidd JM 3rd, Cohen SH, Sosman AJ, Fink JN. Food-dependent exercise-induced anaphylaxis. *J Allergy Clin Immunol* 1983;71:407-11. (III).
- E221. Kivity S, Sneh E, Greif J, Topilsky M, Mekori YA. The effect of food and exercise on the skin response to compound 48/80 in patients with food-associated exercise-induced urticaria-angioedema. *J Allergy Clin Immunol* 1988;81: 1155-8. (III).
- E222. Schwartz HJ. Elevated serum tryptase in exercise-induced anaphylaxis. *J Allergy Clin Immunol* 1995;95:917-9. (III).
- E223. Du Toit G. Food-dependent exercise-induced anaphylaxis in childhood. *Pediatr Allergy Immunol* 2007;18:455-63. (IV).
- E224. Palosuo K, Alenius H, Varjonen E, Koivuluhta M, Mikkola J, Keskinen H, et al. A novel wheat gliadin as a cause of exercise-induced anaphylaxis. *J Allergy Clin Immunol* 1999;103:912-7. (IIb).
- E225. Palosuo K, Varjonen E, Nurkkala J, Kalkkinen N, Harvima R, Reunala T, et al. Transglutaminase-mediated cross-linking of a peptic fraction of omega-5 gliadin enhances IgE reactivity in wheat-dependent, exercise-induced anaphylaxis. *J Allergy Clin Immunol* 2003;111:1386-92. (IIb).
- E226. Matsuo H, Kohno K, Niihara H, Morita E. Specific IgE determination to epitope peptides of omega-5 gliadin and high molecular weight glutenin subunit is a useful tool for diagnosis of wheat-dependent exercise-induced anaphylaxis. *J Immunol* 2005;175:8116-22. (IIb).
- E227. Matsuo H, Dahlstrom J, Tanaka A, Kohno K, Takahashi H, Furumura M, et al. Sensitivity and specificity of recombinant omega-5 gliadin-specific IgE measurement for the diagnosis of wheat-dependent exercise-induced anaphylaxis. *Allergy* 2008;63:233-6. (IIa).
- E228. Brockow K, Kneissl D, Valentini L, Zelger O, Grosber M, Kugler C, et al. Using a gluten oral food challenge protocol to improve diagnosis of wheat-dependent exercise-induced anaphylaxis. *J Allergy Clin Immunol* 2015;135: 977-84.e4 (III).
- E229. Oyefara BI, Bahna SL. Delayed food-dependent, exercise-induced anaphylaxis. *Allergy Asthma Proc* 2007;28:64-6. (III).
- E230. Meyer FJ, Ewert R, Hoepfer MM, Olschewski H, Behr J, Winkler J, et al. Peripheral airway obstruction in primary pulmonary hypertension. *Thorax* 2002;57: 473-6. (IIa).
- E231. D'Alonzo GE, Gianotti LA, Pohil RL, Reagle RR, DuRee SL, Fuentes F, et al. Comparison of progressive exercise performance of normal subjects and patients with primary pulmonary hypertension. *Chest* 1987;92:57-62. (IIa).
- E232. Rastogi D, Ngai P, Barst RJ, Koumbourlis AC. Lower airway obstruction, bronchial hyperresponsiveness, and primary pulmonary hypertension in children. *Pediatr Pulmonol* 2004;37:50-5. (IIb).
- E233. Achouh L, Montani D, Garcia G, Jais X, Hamid AM, Mercier O, et al. Pulmonary arterial hypertension masquerading as severe refractory asthma. *Eur Respir J* 2008;32:513-6. (III).
- E234. Reindl I, Kleber FX. Exertional hyperpnea in patients with chronic heart failure is a reversible cause of exercise intolerance. *Basic Res Cardiol* 1996;91(suppl 1):37-43. (IIb).
- E235. Maron BJ. Hypertrophic cardiomyopathy: a systematic review. *JAMA* 2002; 287:1308-20. (IV).
- E236. Frank MJ, Abdulla AM, Watkins LO, Prisant L, Stefadouros MA. Long-term medical management of hypertrophic cardiomyopathy: usefulness of propranolol. *Eur Heart J* 1983;4(suppl F):155-64. (IIb).
- E237. Besley DC, McWilliams GJ, Moodie DS, Castle LW. Long-term follow-up of young adults following permanent pacemaker placement for complete heart block. *Am Heart J* 1982;103:332-7. (III).
- E238. Parker JM, Cary-Freitas B, Berg BW. Symptomatic vascular rings in adulthood: an uncommon mimic of asthma. *J Asthma* 2000;37:275-80. (III).
- E239. Chuang ML, Chang HC, Lim KE, Vintch JR. Gas exchange detection of right-to-left shunt in dyspneic patients: report of three cases. *Int J Cardiol* 2006;108: 117-9. (III).

- E240. Gossage JR, Kanj G. Pulmonary arteriovenous malformations. A state of the art review. *Am J Respir Crit Care Med* 1998;158:643-61. (IV).
- E241. Ofir D, Laveneziana P, Webb KA, Lam YM, O'Donnell DE. Mechanisms of dyspnea during cycle exercise in symptomatic patients with GOLD stage I chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 2008;177:622-9. (IIa).
- E242. Ferrari M, Bonella F, Benini L, Ferrari P, De Iorio F, Testi R, et al. Acid reflux into the oesophagus does not influence exercise-induced airway narrowing in bronchial asthma. *Br J Sports Med* 2008;42:845-50. (IIb).
- E243. Weiner P, Konson N, Sternberg A, Zamir D, Fireman Z. Is gastro-oesophageal reflux a factor in exercise-induced asthma? *Respir Med* 1998;92:1071-5. (IIa).
- E244. Wright RA, Sagatelian MA, Simons ME, McClave SA, Roy TM. Exercise-induced asthma. Is gastroesophageal reflux a factor? *Dig Dis Sci* 1996;41:921-5. (IIb).
- E245. Peterson KA, Samuelson WM, Ruyjin DT, Young DC, Thomas KL, Hilden K, et al. The role of gastroesophageal reflux in exercise-triggered asthma: a randomized controlled trial. *Dig Dis Sci* 2009;54:564-71. (Ib).
- E246. Peters HP, De Kort AF, Van Krevelen H, Akkermans LM, Van Berge Henegouwen GP, Bol E, et al. The effect of omeprazole on gastro-oesophageal reflux and symptoms during strenuous exercise. *Aliment Pharmacol Ther* 1999;13:1015-22. (Ib).
- E247. Haller RG, Lewis SF, Estabrook RW, DiMauro S, Servidei S, Foster DW. Exercise intolerance, lactic acidosis, and abnormal cardiopulmonary regulation in exercise associated with adult skeletal muscle cytochrome c oxidase deficiency. *J Clin Invest* 1989;84:155-61. (III).
- E248. Hooper RG, Thomas AR, Kearl RA. Mitochondrial enzyme deficiency causing exercise limitation in normal-appearing adults. *Chest* 1995;107:317-22. (III).
- E249. Flaherty KR, Wald J, Weisman IM, Zeballos RJ, Schork MA, Blaivas M, et al. Unexplained exertional limitation: characterization of patients with a mitochondrial myopathy. *Am J Respir Crit Care Med* 2001;164:425-32. (IIa).
- E250. Haden JR, Khan DA. Psychiatric syndromes that mimic asthma. *Adv Psychosom Med* 2003;24:72-85. (IV).
- E251. Ringsberg KC, Akerlind I. Presence of hyperventilation in patients with asthma-like symptoms but negative asthma test responses: provocation with voluntary hyperventilation and mental stress. *J Allergy Clin Immunol* 1999;103:601-8. (IIa).
- E252. National Asthma Education and Prevention Program. Expert Panel Report 3 (EPR-3): guidelines for the diagnosis and management of asthma—summary report 2007. *J Allergy Clin Immunol* 2007;120(suppl):S94-138. (IV).
- E253. Global Initiative for Asthma. NHLBI/WHO workshop report. Bethesda: Medical Communication Resources; 2007. (IV).
- E254. Rossing TH, Weiss JW, Breslin FJ, Ingram RH Jr, McFadden ER Jr. Effects of inhaled sympathomimetics on obstructive response to respiratory heat loss. *J Appl Physiol* 1982;52:1119-23. (Ib).
- E255. Latimer KM, O'Byrne PM, Morris MM, Roberts R, Hargreave FE. Bronchoconstriction stimulated by airway cooling. Better protection with combined inhalation of terbutaline sulphate and cromolyn sodium than with either alone. *Am Rev Respir Dis* 1983;128:440-3. (Ib).
- E256. Guidance for Industry. Exercise-induced Bronchospasm (EIB) development of Drugs to Prevent EIB. Draft Guidance. U.S. Department of Health and Human Services, Food and Drug Administration, Centre for Drug Evaluation and Research. 2002 (IV) [http://www.fda.gov/OHRMS/DOCKETS/98fr/02d-0003\\_gdl0001.pdf](http://www.fda.gov/OHRMS/DOCKETS/98fr/02d-0003_gdl0001.pdf).
- E257. Kemp JP, Dockhorn RJ, Shapiro GG, Nguyen HH, Reiss TF, Seidenberg BC, et al. Montelukast once daily inhibits exercise-induced bronchoconstriction in 6- to 14-year-old children with asthma. *J Pediatr* 1998;133(3):424-8. (IV).
- E258. Raissy HH, Harkins M, Kelly F, Kelly HW. Pretreatment with albuterol versus montelukast for exercise-induced bronchospasm in children. *Pharmacotherapy* 2008;28:287-94. (Ib).
- E259. Spooner C, Spooner G, Rowe B. Mast-cell stabilising agents to prevent exercise-induced bronchoconstriction. *Cochrane Database Syst Rev* 2003(4):CD002307. (Ia).
- E260. Anderson SD, Seale JP, Ferris L, Schoeffel RE, Lindsay DA. An evaluation of pharmacotherapy for exercise-induced asthma. *J Allergy Clin Immunol* 1979;64:612-24. (IV).
- E261. Godfrey S, König P. Suppression of exercise-induced asthma by salbutamol, theophylline, atropine, cromolyn, and placebo in a group of asthmatic children. *Pediatrics* 1975;56:930-4. (Ib).
- E262. Godfrey S, König P. Inhibition of exercise-induced asthma by different pharmacological pathways. *Thorax* 1976;31:137-43. (Ib).
- E263. Hofstra WB, Sont JK, Sterk PJ, Neijens HJ, Kuethe MC, Duiverman EJ. Sample size estimation in studies monitoring exercise-induced bronchoconstriction in asthmatic children. *Thorax* 1997;52:739-41. (IIa).
- E264. Godfrey S, Springer C, Bar-Yishay E, Avital A. Cut-off points defining normal and asthmatic bronchial reactivity to exercise and inhalation challenges in children and young adults. *Eur Respir J* 1999;14:659-68. (Ia).
- E265. Haby MM, Peat JK, Mellis CM, Anderson SD, Woolcock AJ. An exercise challenge for epidemiological studies of childhood asthma: validity and repeatability. *Eur Respir J* 1995;8:729-36. (IIa).
- E266. Clinical exercise testing with reference to lung diseases: indications, standardization and interpretation strategies. ERS Task Force on Standardization of Clinical Exercise Testing. European Respiratory Society. *Eur Respir J* 1997;10:2662-89. (IV).
- E267. Grzelewski T, Stelmach I. Exercise-induced bronchoconstriction in asthmatic children: a comparative systematic review of the available treatment options. *Drugs* 2009;69:1533-53. (IV).
- E268. Kattan M, Thomas CM, Keens TG, Mellis CM, Levison H. The response to exercise in normal and asthmatic children. *J Pediatr* 1978;92:718-21. (IIa).
- E269. Hendrickson CD, Lynch JM, Gleeson K. Exercise induced asthma: a clinical perspective. *Lung* 1994;172:1-14. (IV).
- E270. Anderson SD, Rodwell LT, Du Toit J, Young IH. Duration of protection by inhaled salmeterol in exercise-induced asthma. *Chest* 1991;100:1254-60. (Ib).
- E271. Ferrari M, Balestreri F, Baratieri S, Biasin C, Oldani V, Lo Cascio V. Evidence of the rapid protective effect of formoterol dry-powder inhalation against exercise-induced bronchospasm in athletes with asthma. *Clin Invest* 2000;67:510-3. (Ib).
- E272. Ferrari M, Segattini C, Zanon R, Bertaiola M, Balestreri F, Brotto E, et al. Comparison of the protective effect of salmeterol against exercise-induced bronchospasm when given immediately before a cycloergometric test. *Respiration* 2002;69:509-12. (Ib).
- E273. Bisgaard H. Long-acting beta<sub>2</sub>-agonists in management of childhood asthma: a critical review of the literature. *Pediatr Pulmonol* 2000;29:221-34. (IV).
- E274. Anderson SD, Seale JP, Rozea P, Bandler L, Theobald G, Lindsay DA. Inhaled and oral salbutamol in exercise-induced asthma. *Am Rev Respir Dis* 1976;114:493-500. (IIa).
- E275. McFadden ER, Gilbert IA. Exercise-induced asthma. *N Engl J Med* 1994;330:1362-7. (IV).
- E276. Tan RA, Spector SL. Exercise-induced asthma: diagnosis and management. *Ann Allergy* 2002;89:226-36. (IV).
- E277. Kemp JP, Dockhorn RJ, Busse WW, Bleecker ER. Prolonged effect of inhaled salmeterol against exercise-induced bronchospasm. *Am J Respir Crit Care Med* 1994;150:1612-5. (Ib).
- E278. Nelson JA, Strauss L, Skowronski M, Ciuffo R, Novak R, McFadden ER. Effect of long-term salmeterol treatment on exercise-induced asthma. *N Engl J Med* 1998;339:141-6. (Ib).
- E279. Carlsen KH, Roksund O, Olsholt K, Nija F, Leegard J, Bratten G. Overnight protection by inhaled salmeterol on exercise-induced asthma in children. *Eur Respir J* 1995;8:1852-5. (Ib).
- E280. Boner AL, Spezia E, Piovesan P, Chiocca E, Maiocchi G. Inhaled formoterol in the prevention of exercise-induced bronchoconstriction in asthmatic children. *Am J Respir Crit Care Med* 1994;149:935-8. (Ib).
- E281. Vilsvik J, Ankerst J, Palmqvist M, Persson G, Schaanning J, Schwabe G, et al. Protection against cold air and exercise-induced bronchoconstriction while on regular treatment with Oxis<sup>®</sup>. *Respir Med* 2001;95:484-90. (Ib).
- E282. Bronsky EA, Yegen Ü, Yeh CM, Larsen LV, Della Cioppa G. Formoterol provides long-lasting protection against exercise-induced bronchospasm. *Ann Allergy Asthma Immunol* 2002;89:407-12. (Ib).
- E283. Ramage L, Lipworth BJ, Ingram CG, Cree IA, Dhillon DP. Reduced protection against exercise induced bronchoconstriction after chronic dosing with salmeterol. *Respir Med* 1994;88:363-8. (Ib).
- E284. Simons FE, Gerstner TV, Cheang MS. Tolerance to the bronchoprotective effect of salmeterol in adolescents with exercise-induced asthma using concurrent inhaled glucocorticoid treatment. *Pediatrics* 1997;99:655-9. (Ib).
- E285. Hanev S, Hancox RJ. Rapid onset of tolerance to beta-agonist bronchodilation. *Respir Med* 2005;99:566-71. (IIa).
- E286. Villaran C, O'Neill J, Helbling A, van Noord JA, Lee TH, Chuchalin AG, et al. Montelukast versus salmeterol in patients with asthma and exercise-induced bronchoconstriction. *J Allergy Clin Immunol* 1999;104:547-53. (Ib).
- E287. Edelman JM, Turpin JA, Bronsky EA. Oral Montelukast compared with inhaled salmeterol to prevent exercise-induced bronchoconstriction. *Ann Intern Med* 2000;132:97-104. (Ib).
- E288. Hancox RJ, Subbarao P, Kamada D, Watson RM, Hargreave FE, Inman MD. Beta<sub>2</sub>-agonist tolerance and exercise-induced bronchospasm. *Am J Respir Crit Care Med* 2002;165:1068-70. (Ib).
- E289. Inman MD, O'Byrne PM. The effect of regular inhaled albuterol on exercise-induced bronchoconstriction. *Am J Respir Crit Care Med* 1996;153:65-9. (Ib).



- E290. Wraight JM, Hancox RJ, Herbison GP, Cowan JO, Flannery EM, Taylor DR. Bronchodilator tolerance: the impact of increasing bronchoconstriction. *Eur Respir J* 2003;21:810-5. (Ib).
- E291. Hancox RJ, Aldridge EE, Cowan JO, Flannery EM, Herbison GP, McLachlan CR, et al. Tolerance to beta-agonists during acute bronchoconstriction. *Eur Respir J* 1999;14:283-7. (Ib).
- E292. Hancox RJ, Cowan JO, Flannery EM, Herbison GP, McLachlan CR, Taylor DR. Bronchodilator tolerance and rebound bronchoconstriction during regular inhaled beta-agonist treatment. *Respir Med* 2000;94:767-71. (Ib).
- E293. Haney S, Hancox RJ. Overcoming beta-agonist tolerance: high dose salbutamol and ipratropium bromide. Two randomised controlled trials. *Respir Res* 2007;8:19. (Ib).
- E294. Bonini M, Permaul P, Kulkarni T, Kazani S, Segal A, Sorkness CA, et al. Loss of salmeterol bronchoprotection against exercise in relation to ADRB2 Arg16-Gly polymorphism and exhaled nitric oxide. *Am J Respir Crit Care Med* 2013;188:1407-12. (Ib).
- E295. Aziz I, Tan KS, Hall IP, Devlin MM, Lipworth BJ. Subsensitivity to bronchoprotection against adenosine monophosphate challenge following regular once-daily formoterol. *Eur Respir J* 1998;12:580-4. (Ib).
- E296. Drotar DE, Davis EE, Cockcroft DW. Tolerance to the bronchoprotective effect of salmeterol 12 hours after starting twice daily treatment. *Ann Allergy Asthma Immunol* 1998;80:31-4. (Ib).
- E297. Bhagat R, Kalra S, Swystun VA, Cockcroft DW. Rapid onset of tolerance to the bronchoprotective effect of salmeterol. *Chest* 1995;108:1235-9. (Ib).
- E298. Giannini D, Carlett A, Dente FL, Bacci E, DiFranco A, Vagaggini B, et al. Tolerance to the protective effect of salmeterol on allergen challenge. *Chest* 1996;110:1452-7. (Ib).
- E299. Storms W, Chervinsky P, Ghannam AF, Bird S, Hustad CM, Edelman JM. A comparison of the effects of oral montelukast and inhaled salmeterol on response to rescue bronchodilation after challenge. *Respir Med* 2004;98:1051-62. (Ib).
- E300. Garcia R, Guerra P, Feo F, Galindo PA, Gomez E, Borja J, et al. Tachyphylaxis following regular use of formoterol in exercise-induced bronchospasm. *J Investig Allergol Clin Immunol* 2001;11:176-82.
- E301. Johnson M. Molecular mechanisms of  $\beta_2$  adrenergic receptor function, response and regulation. *J Allergy Clin Immunol* 2006;117:18-24. (IV).
- E302. Hayes MJ, Qing F, Rhodes CG, Rahman SU, Ind PW, Sriskandan S, et al. In vivo quantification of human pulmonary beta-adrenoceptors: effect of beta-agonist therapy. *Am J Respir Crit Care Med* 1996;154:1277-83. (III).
- E303. McGraw DW, Liggett SB. Heterogeneity of beta adrenergic receptor kinase expression in the lung accounts for cell-specific desensitisation of the beta adrenergic receptor. *J Biol Chem* 1997;272:7338-44. (IIa).
- E304. Scola AM, Chong LK, Suvarna SK, Chess-Williams R, Peachell PT. Desensitisation of mast cell  $\beta_2$ -adrenoceptor-mediated responses by salmeterol and formoterol. *Br J Pharmacol* 2004;141:163-71. (LB).
- E305. Chong LK, Suvarna K, Chess-Williams R, Peachell PT. Desensitization of  $\beta_2$ -adrenoceptor-mediated responses by short-acting  $\beta_2$ -adrenoceptor agonists in human lung mast cells. *Br J Pharmacol* 2003;138:512-20. (LB).
- E306. O'Connor BJ, Aikman S, Barnes PJ. Tolerance to the non-bronchodilator effects of inhaled beta-agonists in asthma. *N Engl J Med* 1992;327:1204-8. (Ib).
- E307. Peachell P. Regulation of mast cells by  $\beta_2$ -agonists. *Clin Rev Allergy Immunol* 2006;31:131-42. (IV).
- E308. Swystun VA, Gordon JR, Davis EB, Zhand X, Cockcroft DW. Mast cell tryptase release and asthmatic responses to allergen increase with regular use of salbutamol. *J Allergy Clin Immunol* 2000;106:57-64. (Ib).
- E309. Anderson SD, Caillaud C, Brannan JD.  $\beta_2$ -Agonists and exercise-induced asthma. *Clin Rev Allergy Immunol* 2006;31:163-80. (IV).
- E310. Mak JCW, Roffel F, Katsunuma T, Elzinga CRS, Zaagsma J, Barnes PJ. Up-regulation of airway smooth muscle histamine H1 receptor mRNA, protein, and function by beta 2-adrenoceptor activation. *Mol Pharmacol* 2000;57:857-64. (LB).
- E311. McGraw DW, Elwing JM, Fogel KM, Wang WCH, Glinka CB, Muhihbachler KA, et al. Cross talk between Gi and Gq/Gs pathways in airway smooth muscle regulates bronchial contractility and relaxation. *J Clin Invest* 2007;117:1391-8. (LB).
- E312. Anderson GP. Current issues with  $\beta_2$ -adrenoceptor agonists: pharmacology and molecular and cellular mechanisms. *Clin Rev Allergy Immunol* 2006;31:119-30. (IV).
- E313. Finnerty JP, Holgate ST. The contribution of histamine release and vagal reflexes, alone and in combination, to exercise-induced asthma. *Eur Respir J* 1993;6:1132-7. (Ib).
- E314. Knopfli BH, Bar-Or O, Araujo CG. Effect of ipratropium bromide on EIB in children depends on vagal activity. *Med Sci Sports Exerc* 2005;37:354-9. (IIa).
- E315. Sarria B, Naline E, Zhang Y, Cortijo J, Molimard M, Moreau J, et al. Muscarinic M2 receptors in acetylcholine-isoproterenol functional antagonism in human isolated bronchus. *Am J Physiol Lung Cell Mol Physiol* 2002;283:L1125-32. (IIa).
- E316. Kalra S, Swystun VA, Bhagat R, Cockcroft DW. Inhaled corticosteroids do not prevent the development of tolerance to the bronchoprotective effect of salmeterol. *Chest* 1996;109:953-6. (Ib).
- E317. FDA drug safety communication: new safety requirements for long-acting inhaled asthma medications called long-acting beta-agonists (LABAs). Washington (DC): US Food and Drug Association; 2010. (IV).
- E318. Stelmach I, Grzelewski T, Majak P, Jerzynska J, Stelmach W, Kuna P. Effect of different antiasthmatic treatments on exercise-induced bronchoconstriction in children with asthma. *J Allergy Clin Immunol* 2008;121:383-9. (Ib).
- E319. Vidal C, Fernández-Ovide E, Piñero J, Nuñez R, González-Quintela A. Budesonide or montelukast prevents exercise-induced bronchoconstriction. *Ann Allergy Asthma Immunol* 2001;86:655-8. (IIa).
- E320. Finnerty JP, Holgate ST. Evidence for the roles of histamine and prostaglandins as mediators in exercise-induced asthma: the inhibitory effect of terfenadine and flurbiprofen alone and in combination. *Eur Respir J* 1990;3:540-7. (Ib).
- E321. O'Byrne PM. Leukotriene bronchoconstriction induced by allergen and exercise. *Am J Respir Crit Care Med* 2000;161(suppl):S68-72. (IV).
- E322. Pearlman DS, Ostrom NK, Bronsky EA, Bonuccelli CM, Hanby LA. The leukotriene D<sub>4</sub>-receptor antagonist zafirlukast attenuates exercise-induced bronchoconstriction in children. *J Pediatr* 1999;134:273-9. (Ib).
- E323. Manning PJ, Watson RM, Margolskee DJ, Williams VC, Schwartz JI, O'Byrne PM. Inhibition of exercise-induced bronchoconstriction by MK-571, a potent leukotriene D<sub>4</sub>-receptor antagonist. *N Engl J Med* 1990;323:1736-9. (Ib).
- E324. Finnerty JP, Wood-Baker R, Thomson H, Holgate S. Role of leukotrienes in exercise-induced asthma. Inhibitory effect of ICI 204219, a potent leukotriene D<sub>4</sub> receptor antagonist. *Am Rev Respir Dis* 1992;145:746-9. (Ib).
- E325. Philip G, Villaran C, Pearlman DS, Loeys T, Dass SB, Reiss TF. Protection against exercise-induced bronchoconstriction two hours after a single oral dose of montelukast. *J Asthma* 2007;44:213-7. (Ib).
- E326. Wasfi YS, Kemp JP, Villaran C, Massaad R, Xin W, Smugar SS, et al. Onset and duration of attenuation of exercise-induced bronchoconstriction in children by single-dose of montelukast. *Allergy Asthma Proc* 2011;32:453-9. (Ib).
- E327. Philip G, Pearlman DS, Villaran C, Legrand C, Loeys T, Langdon RB, et al. Single-dose montelukast or salmeterol as protection against exercise-induced bronchoconstriction. *Chest* 2007;132:875-83. (Ib).
- E328. Bronsky EA, Kemp JP, Zhand J, Guerreiro D, Reiss TF. Dose-related protection of exercise bronchoconstriction by montelukast, a cysteinyl leukotriene-receptor antagonist, at the end of a once-daily dosing interval. *Clin Pharmacol Ther* 1997;62:556-61. (Ib).
- E329. Peroni DG, Piacentini GL, Ressa M, Bodini A, Loiacono A, Aralla R, et al. Time efficacy of a single dose of montelukast on exercise-induced asthma in children. *Pediatr Allergy Immunol* 2002;13:434-7. (Ib).
- E330. de Benedictis FM, del Giudice MM, Forenza N, Decimo F, de Benedictis D, Capristo A. Lack of tolerance to the protective effect of montelukast in exercise-induced bronchoconstriction in children. *Eur Respir J* 2006;28:291-5. (Ib).
- E331. Drazen JM, Silverman EK, Lee TH. Heterogeneity of therapeutic responses in asthma. *Br Med Bull* 2000;56:1054-70. (IV).
- E332. Kang MJ, Lee SY, Kim HB, Yu J, Kim BJ, Choi WA, et al. Association of IL-13 polymorphisms with leukotriene receptor antagonist drug responsiveness in Korean children with exercise-induced bronchoconstriction. *Pharmacogenomics* 2008;18:551-8. (III).
- E333. Kim JH, Lee SY, HBKim, Jin HS, Yu JH, Kim BJ, et al. TBXA2R gene polymorphism and responsiveness to leukotriene receptor antagonist in children with asthma. *Clin Exp Allergy* 2008;38:51-9. (IIa).
- E334. Lehnigk B, Rabe KF, Dent G, Herst RS, Carpentier PJ, Magnussen H. Effects of a 5-lipoxygenase inhibitor, ABT-761, on exercise-induced bronchoconstriction and urinary LTE<sub>4</sub> in asthmatic patients. *Eur Respir J* 1998;11:617-23. (Ib).
- E335. van Schoor J, Joos GF, Kips JC, Drajesk JF, Carpentier PJ, Pauwels RA. The effect of ABT-761, a novel 5-lipoxygenase inhibitor, on exercise- and adenosine-induced bronchoconstriction in asthmatic subjects. *Am J Respir Crit Care Med* 1997;155:875-80. (Ib).
- E336. Kent SE, Bentley JH, Miller D, Sterling R, Menendez R, Tarpay M, et al. The effect of GSK2190915, a 5-lipoxygenase-activating protein inhibitor, on exercise-induced bronchoconstriction. *Allergy Asthma Proc* 2014;35:126-33. (Ib).
- E337. Anderson SD, Kippelen P. Assessment and prevention of exercise-induced bronchoconstriction. *Br J Sports Med* 2012;46:391-6.
- E338. Kelly KD, Spooner CH, Rowe BH. Nedocromil sodium *versus* sodium cromoglycate in treatment of exercise-induced bronchoconstriction: a systematic review. *Eur Respir J* 2001;17:39-45. (Ia).



- E339. Brannan JD, Gulliksson M, Anderson SD, Chew N, Seale JP, Kumlin M. Inhibition of mast cell PGD<sub>2</sub> release protects against mannitol-induced airway narrowing. *Eur Respir J* 2006;27:944-50. (IIa).
- E340. Silverman M, Andrea T. Time course of effect of disodium cromoglycate on exercise-induced asthma. *Arch Dis Child* 1972;47:419-22. (Ib).
- E341. Comis A, Valletta EA, Sette L, Andreoli A, Boner AL. Comparison of nedocromil sodium and sodium cromoglycate administered by pressurized aerosol, with and without a spacer device in exercise-induced asthma in children. *Eur Respir J* 1993;6:523-6. (Ib).
- E342. de Benedictis FM, Tuteri G, Pazzelli P, Solinas LF, Niccoli A, Parente C. Combination drug therapy for the prevention of exercise-induced bronchoconstriction in children. *Ann Allergy Asthma Immunol* 1998;80:352-6. (Ib).
- E343. Tullett WM, Tan KM, Wall RT, Patel KR. Dose-response effect of sodium cromoglycate pressurized aerosol in exercise induced asthma. *Thorax* 1985;40:41-4. (Ib).
- E344. Patel KR, Wall RT. Dose-duration effect of sodium cromoglycate aerosol in exercise-induced asthma. *Eur J Respir Dis* 1986;69:256-60. (Ib).
- E345. Schoeffel RE, Anderson SD, Lindsay DA. Sodium Cromoglycate as a pressurized aerosol (Vicrom) in exercise-induced asthma. *Aust NZ J Med* 1983;13:157-61. (IIa).
- E346. Patel KR, Tullett WM, Neale MG, Wall RT, Tan KM. Plasma concentrations of sodium cromoglycate given by nebulisation and metered dose inhalers in patients with exercise-induced asthma: relationship to protective effect. *Br J Clin Pharmacol* 1986;21:231-3. (Ib).
- E347. Kuzemko JA. Twenty years of sodium cromoglycate treatment: a short review. *Respir Med* 1989;83:11-6. (IV).
- E348. Hofstra WB, Neijens HJ, Duiverman EJ, Kouwenberg JM, Mulder PG, Kueth MC, et al. Dose-response over time to inhaled fluticasone propionate: treatment of exercise- and methacholine-induced bronchoconstriction in children with asthma. *Pediatr Pulmonol* 2000;29:415-23. (Ib).
- E349. Jonasson G, Carlsen KH, Hultquist C. Low-dose budesonide improves exercise-induced bronchospasm in schoolchildren. *Pediatr Allergy Immunol* 2000;11:120-5. (Ib).
- E350. National Institutes of Health; National Heart, Lung, and Blood Institute. Expert panel report 3 (EPR-3): guidelines for the diagnosis and management of asthma—summary report 2007. *J Allergy Clin Immunol* 2007;120(suppl):S94-138.
- E351. Anderson SD, Holzer K. Exercise-induced asthma: Is it the right diagnosis in elite athletes? *J Allergy Clin Immunol* 2000;106:419-28. (IV).
- E352. Brannan JD. Bronchial hyperresponsiveness in the assessment of asthma control: airway hyperresponsiveness in asthma: its measurement and clinical significance. *Chest* 2010;138(suppl):11S-7S. (IV).
- E353. Pedersen S, Hansen OR. Budesonide treatment of moderate and severe asthma in children: a dose-response study. *J Allergy Clin Immunol* 1995;95:29-33. (Ib).
- E354. Madhuban AA, Driessen JM, Brusse-Keizer MG, van Aalderen WM, de Jongh FH, Thio BJ. Association of the asthma control questionnaire with exercise-induced bronchoconstriction. *J Asthma* 2011;48:275-8. (Ib).
- E355. Thio BJ, Slingerland GLM, Nagelkerke AF, Roord JJ, Mulder PGH, Dankert-Roelse JE. Effects of single-dose fluticasone on exercise-induced asthma in asthmatic children: a pilot study. *Pediatr Pulmonol* 2001;32:115-21. (IIa).
- E356. Driessen JM, Nieland H, van der Palen JA, van Aalderen WM, Thio BJ, de Jongh FH. Effects of a single dose inhaled corticosteroid on the dynamics of airway obstruction after exercise. *Pediatr Pulmonol* 2011;46:849-56. (Ib).
- E357. Henriksen JM, Wenzel A. Effect of an intranasally administered corticosteroid (budesonide) on nasal obstruction, mouth breathing, and asthma. *Am Rev Respir Dis* 1984;130:1014-8. (Ib).
- E358. Kersten ET, van Leeuwen JC, Brand PL, Duiverman EJ, de Jongh FH, Thio BJ, et al. Effect of an intranasal corticosteroid on exercise induced bronchoconstriction in asthmatic children. *Pediatr Pulmonol* 2012;47:27-35. (Ib).
- E359. Shturman-Ellstein R, Zeballos RJ, Buckley JM, Souhrada JF. The beneficial effect of nasal breathing on exercise-induced bronchoconstriction. *Am Rev Respir Dis* 1978;118:65-73. (Ib).
- E360. Brozek JL, Bousquet J, Baena-Cagnani CE, Bonini S, Canonica GW, Casale TB, et al. Allergic Rhinitis and its Impact on Asthma (ARIA) guidelines: 2010 revision. *J Allergy Clin Immunol* 2010;126:466-76. (IV).
- E361. Duong M, Amin R, Baatjes AJ, Kritzing F, Qi Y, Meghji Z, et al. The effect of montelukast, budesonide alone, and in combination on exercise-induced bronchoconstriction. *J Allergy Clin Immunol* 2012;130:535-9.e3. (Ib).
- E362. Carlsen KH, Anderson SD, Bjermer L, Bonini S, Brusasco V, Canonica W, et al. Treatment of exercise-induced asthma, respiratory and allergic disorders in sports and the relationship to doping: part II of the report from the Joint Task Force of European Respiratory Society (ERS) and European Academy of Allergy and Clinical Immunology (EAACI) in cooperation with GA(2)LEN. *Allergy* 2008;63:492-505. (IV).
- E363. Yates DH, Kharitonov S, Barnes PJ. An inhaled glucocorticoid does not prevent tolerance to the protective effect of a long-acting inhaled beta 2-agonist. *Am J Respir Crit Care Med* 1996;154:1603-7. (Ib).
- E364. Pearlman D, Qaundah P, Matz J, Yancey SW, Stempel DA, Ortega HG. Fluticasone propionate/salmeterol and exercise-induced asthma in children with persistent asthma. *Pediatr Pulmonol* 2009;44:429-35. (Ib).
- E365. Lazarinis N, Jorgensen L, Ekstrom T, Bjermer L, Dahlen B, Pullerits T, et al. Combination of budesonide/formoterol on demand improves asthma control by reducing exercise-induced bronchoconstriction. *Thorax* 2014;69:130-6. (Ib).
- E366. Blake K. Review of guidelines and the literature in the treatment of acute bronchospasm in asthma. *Pharmacotherapy* 2006;26(suppl):148S-55S. (IV).
- E367. Beil M, de Kock MA. Role of alpha-adrenergic receptors in exercise-induced bronchoconstriction. *Respiration* 1978;35:78-86. (IIa).
- E368. Boulet L-P, Turcotte H, Tennina S. Comparative efficacy of salbutamol, ipratropium and cromoglycate in the prevention of bronchospasm induced by exercise and hyperosmolar challenges. *J Allergy Clin Immunol* 1989;83:882-7. (Ib).
- E369. Poppius H, Sovijarvi ARA, Tammilehto L. Lack of protective effect of high-dose ipratropium on bronchoconstriction following exercise with cold air breathing in patients with mild asthma. *Eur J Respir Dis* 1986;68:319-25. (Ib).
- E370. Magnussen H, Nowak D, Wiebicke W. Effect of inhaled ipratropium bromide on the airway response to methacholine, histamine, and exercise in patients with mild bronchial asthma. *Respiration* 1992;59:42-7. (Ib).
- E371. Boner AL, Vallone G, De Stefano G. Effect of inhaled ipratropium bromide on methacholine and exercise provocation in asthmatic children. *Pediatr Pulmonol* 1989;6:81-5. (Ib).
- E372. Hallstrand TS, Kippelen P, Larsson J, Bougault V, van Leeuwen JC, Driessen JM, et al. Where to from here for exercise-induced bronchoconstriction: the unanswered questions. *Immunol Allergy Clin North Am* 2013;33:423-42. ix. (IV).
- E373. Ellis EF. Inhibition of exercise-induced asthma by theophylline. *J Allergy Clin Immunol* 1984;73:690-2. (IV).
- E374. Iikura Y, Hashimoto K, Akasawa A, Katsunuma T, Ebisawa M, Saito H, et al. Serum theophylline concentration levels and preventative effects on exercise-induced asthma. *Clin Exp Allergy* 1996;26(suppl 2):38-41. (IIa).
- E375. Seale JP, Anderson SD, Lindsay DA. A comparison of oral theophylline and oral salbutamol in exercise-induced asthma. *Aust NZ J Med* 1977;7:270-4. (IIa).
- E376. Timmer W, Lecher V, Birraux G, Neuhäuser M, Hatzelmann A, Bethke T, et al. The phosphodiesterase 4 inhibitor roflumilast is efficacious in exercise-induced asthma and leads to suppression of LPS-stimulated TNF- $\alpha$  ex vivo. *J Clin Pharmacol* 2002;42:297-303. (Ib).
- E377. Duffy P, Phillips YY. Caffeine consumption decreases the response to bronchoprovocation challenge with dry gas hyperventilation. *Chest* 1991;99:1374-7. (Ib).
- E378. Kivity S, Ben Aharon Y, Man A, Topilsky M. The effect of caffeine on exercise-induced bronchoconstriction. *Chest* 1990;97:1083-5. (Ib).
- E379. Clee MD, Ingram CG, Reid PC, Robertson AS. The effect of astemizole on exercise-induced asthma. *Br J Dis Chest* 1984;78:180-3. (IIa).
- E380. Magnussen H, Reuss G, Jörres R, Aurich R. The effect of azelastine on exercise-induced asthma. *Chest* 1988;93:937-40. (Ib).
- E381. Wiebicke W, Poynter A, Montgomery M, Chernick V, Pasterkamp H. Effect of terfenadine on the response to exercise and cold air in asthma. *Pediatr Pulmonol* 1988;4:225-9. (Ib).
- E382. Zielinski J, Chodosowska E. Exercise-induced bronchoconstriction in patients with bronchial asthma. Its prevention with an antihistaminic agent. *Respiration* 1977;34:31-5. (IIa).
- E383. Manjra AI, Nel H, Maharaj B. Effect of desloratadine on patients with allergic rhinitis and exercise-induced bronchoconstriction: a placebo controlled study. *J Asthma* 2009;46:156-9. (Ib).
- E384. O'Byrne PM. Leukotrienes in the pathogenesis of asthma. *Chest* 1997;111(suppl 2):27S-34S. (IV).
- E385. Passalacqua G, Canonica GW, Bousquet J. Structure and classification of H1-antihistamines and overview of their activities. *Clin Allergy Immunol* 2002;17:65-100. (IV).
- E386. Ghosh SK, De Vos C, McIlroy I, Patel KR. Effect of cetirizine on exercise induced asthma. *Thorax* 1991;46:242-4. (Ib).
- E387. Barnes PJ, Wilson NM, Brown MJ. A calcium antagonist, nifedipine, modifies exercise-induced asthma. *Thorax* 1981;36:726-30. (Ib).
- E388. Corris PA, Nariman S, Gibson GJ. Nifedipine in the prevention of asthma induced by exercise and histamine. *Am Rev Respir Dis* 1983;128:991-2. (Ib).
- E389. Lockhart A, Slutsky AS. Furosemide and loop diuretics in human asthma. *Chest* 1994;106:244-9. (IV).

- E390. Prandota J. Furosemide: progress in understanding its diuretic, anti-inflammatory, and bronchodilating mechanism of action, and use in the treatment of respiratory tract diseases. *Am J Ther* 2002;9:317-28. (IV).
- E391. Bianco S, Vaghi A, Robuschi M, Pasargiklian M. Prevention of exercise-induced bronchoconstriction by inhaled frusemide. *Lancet* 1988;2:252-5. (Ib).
- E392. Barnes PJ, Wilson NM, Vickers H. Prazosin, an alpha 1-adrenoceptor antagonist, partially inhibits exercise-induced asthma. *J Allergy Clin Immunol* 1981; 68:411-5. (Ib).
- E393. Ahmed T, Gonzalez BJ, Danta I. Prevention of exercise-induced bronchoconstriction by inhaled low-molecular-weight heparin. *Am J Respir Crit Care Med* 1999;160:576-81. (Ib).
- E394. Kunz LI, van Rensen EL, Sterk PJ. Inhaled hyaluronic acid against exercise-induced bronchoconstriction in asthma. *Pulm Pharmacol Ther* 2006;19: 286-91. (Ib).
- E395. Anderson SD, Daviskas E. The mechanism of exercise-induced asthma is .... *J Allergy Clin Immunol* 2000;106:453-9. (IV).
- E396. Stickland MK, Rowe BH, Spooner CH, Vandermeer B, Dryden DM. Effect of warm-up exercise on exercise-induced bronchoconstriction. *Med Sci Sports Exerc* 2012;44:383-91. (Ia).
- E397. Schoeffel RE, Anderson SD, Gillam I, Lindsay DA. Multiple exercise and histamine challenge in asthmatic patients. *Thorax* 1980;35:164-70. (IIb).
- E398. Anderson SD, Schoeffel RE. Respiratory heat and water loss during exercise in patients with asthma: effect of repeated exercise challenge. *Eur J Respir Dis* 1982;63:472-80. (IIb).
- E399. McKenzie DC, McLuckie SL, Stirling DR. The protective effects of continuous and interval exercise in athletes with exercise-induced asthma. *Med Sci Sports Exerc* 1994;26:951-6. (Ib).
- E400. O'Byrne PM, Jones GL. The effect of indomethacin on exercise-induced bronchoconstriction and refractoriness after exercise. *Am Rev Respir Dis* 1986;134: 69-72. (IIa).
- E401. Mickleborough TD, Lindley MR, Ray S. Dietary salt, airway inflammation, and diffusing capacity in exercise-induced asthma. *Med Sci Sports Exerc* 2005;37: 904-14. (Ib).
- E402. Tecklenburg SL, Mickleborough TD, Fly AD, Bai Y, Stager JM. Ascorbic acid supplementation attenuates exercise-induced bronchoconstriction in patients with asthma. *Respir Med* 2007;101:1770-8. (Ib).
- E403. Mickleborough TD, Lindley MR, Ionescu AA, Fly AD. Protective effect of fish oil supplementation on exercise-induced bronchoconstriction in asthma. *Chest* 2006;129:39-49. (Ib).
- E404. Gotshall RW, Mickleborough TD, Cordain L. Dietary salt restriction improves pulmonary function in exercise-induced asthma. *Med Sci Sports Exerc* 2000;32: 1815-9. (Ib).
- E405. Tecklenburg-Lund S, Mickleborough TD, Turner LA, Fly AD, Stager JM, Montgomery GS. Randomized controlled trial of fish oil and montelukast and their combination on airway inflammation and hyperpnea-induced bronchoconstriction. *PLoS One* 2010;5:e13487. (Ib).
- E406. Mickleborough TD, Vaughn CL, Shei RJ, Davis EM, Wilhite DP. Marine lipid fraction PCSO-524 (lyprinol/omega XL) of the New Zealand green lipped mussel attenuates hyperpnea-induced bronchoconstriction in asthma. *Respir Med* 2013;107:1152-63. (Ib).
- E407. Arm JP, Horton CE, Mencia-Huerta JM, House F, Eiser NM, Clark TJ, et al. Effect of dietary supplementation with fish oil lipids on mild asthma. *Thorax* 1988;43:84-92. (Ib).
- E408. Brannan JD, Bood J, Alkhabaz A, Balgoma D, Otis J, Delin I, et al. The effect of omega-3 fatty acids on bronchial hyperresponsiveness, sputum eosinophilia, and mast cell mediators in asthma. *Chest* 2015;147:397-405. (Ib).
- E409. Price OJ, Hull JH, Howatson G, Robson-Ansley P, Ansley L. Vitamin D and omega-3 polyunsaturated fatty acid supplementation in athletes with exercise-induced bronchoconstriction: a pilot study. *Expert Rev Respir Med* 2015;9: 369-78. (IIa).
- E410. Cohen HA, Neuman I, Nahum H. Blocking effect of vitamin C in exercise-induced asthma. *Arch Pediatr Adolesc Med* 1997;151:367-70. (Ib).
- E411. Schachter EN, Schlesinger A. The attenuation of exercise-induced bronchospasm by ascorbic acid. *Ann Allergy* 1982;49:146-51. (Ib).
- E412. Boulet LP, O'Byrne PM. Asthma and Exercise-induced bronchoconstriction in athletes. *N Engl J Med* 2015;372:613-8. (IV).
- E413. Simpson AJ, Tufvesson E, Anderson SD, Romer LM, Bjermer L, Kippelen P. Effect of terbutaline on hyperpnoea-induced bronchoconstriction and urinary club cell protein 16 in athletes. *J Appl Physiol* (1985) 2013;115: 1450-6. (Ib).
- E414. Rundell K, Spiering BA, Baumann JM, Evans TM. Effects of montelukast on airway narrowing from eucapnic voluntary hyperventilation and cold air exercise. *Br J Sports Med* 2005;39:232-6. (Ib).
- E415. Anderson SD, Kippelen P. Airway injury as a mechanism for exercise-induced bronchoconstriction in elite athletes. *J Allergy Clin Immunol* 2008;122:225-35. (IV).
- E416. Karjalainen E-M, Laitinen A, Sue-Chu M, Altraja A, Bjermer L, Laitinen LA. Evidence of airway inflammation and remodeling in ski athletes with and without bronchial hyperresponsiveness to methacholine. *Am J Respir Crit Care Med* 2000;161:2086-91. (IIa).
- E417. Sue-Chu M, Brannan JD, Anderson SD, Chew N, Bjermer L. Airway responsiveness to methacholine (Mch), adenosine 5-monophosphate (AMP), mannitol (Man), eucapnic voluntary hyperpnea (EVH) and sport specific field exercise challenge (Ex) in cross country ski athletes. *Eur Respir J* 2002;20(suppl 38): 410s. (IIb).
- E418. Stensrud T, Mykland KV, Gabrielsen K, Carlsen KH. Bronchial hyperresponsiveness in skiers: field test versus methacholine provocation? *Med Sci Sports Exerc* 2007;39:1681-6. (IIb).
- E419. Bougault V, Turmel J, Boulet LP. Bronchial challenges and respiratory symptoms in elite swimmers and winter sport athletes: airway hyperresponsiveness in asthma: its measurement and clinical significance. *Chest* 2010;138(suppl): 31S-7S. (IIb).
- E420. Hull JH, Hull PJ, Parsons JP, Dickinson JW, Ansley L. Approach to the diagnosis and management of suspected exercise-induced bronchoconstriction by primary care physicians. *BMC Pulm Med* 2009;9:29. (IV).
- E421. Pedersen L, Winther S, Backer V, Anderson SD, Larsen KR. Airway responses to eucapnic hyperpnea, exercise and methacholine in elite swimmers. *Med Sci Sports Exerc* 2008;40:1567-72. (IIb).
- E422. Kippelen P, Anderson SD. Pathogenesis of exercise-induced bronchoconstriction. *Immunol Allergy Clin North Am* 2013;33:299-312. vii. (IV).

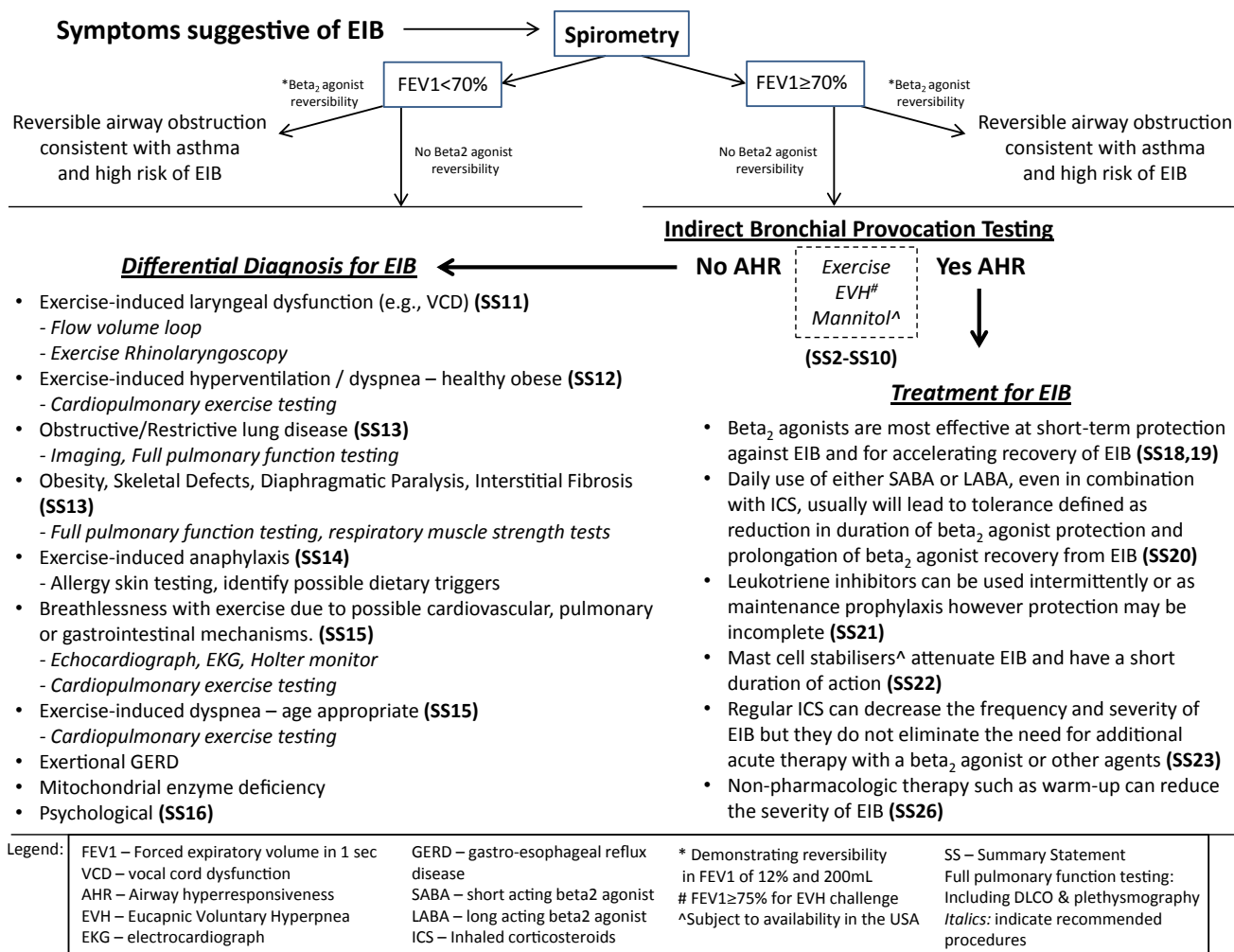
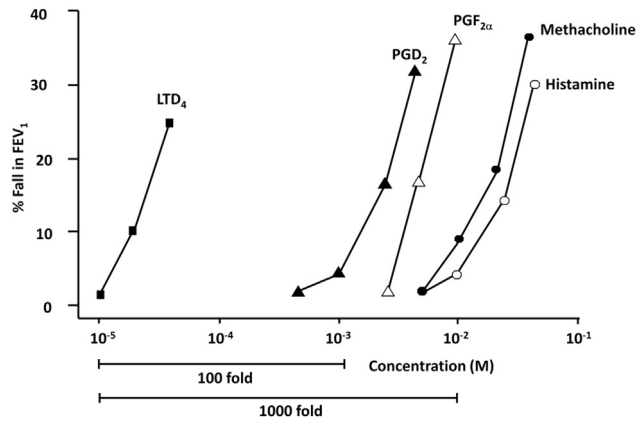
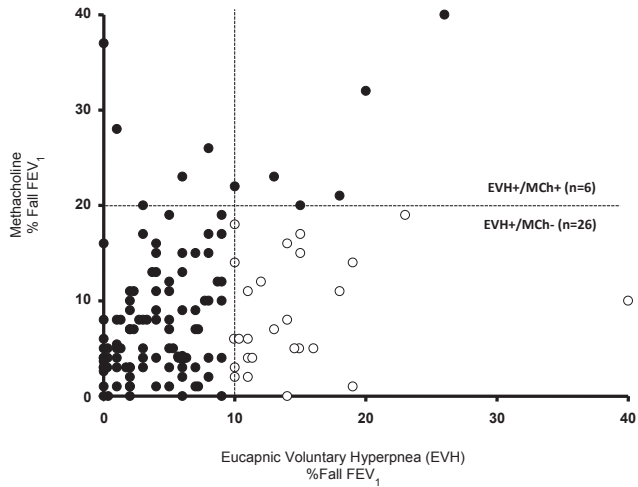


FIG E1. Algorithm for the diagnosis, treatment, and differential diagnosis of EIB.

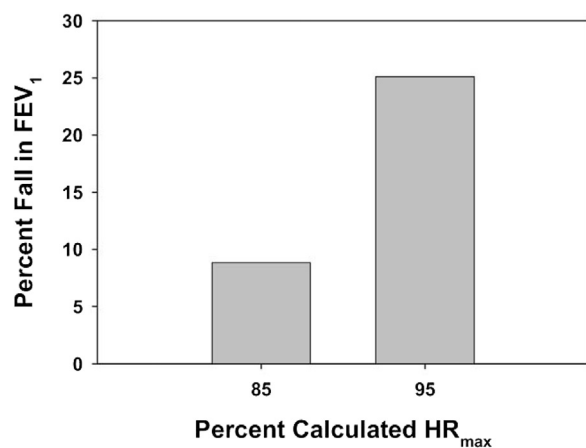


**FIG E2.** Airway smooth muscle sensitivity to mediators of bronchoconstriction. The potency of mediators of bronchoconstriction that are released in the presence of dry air hyperpnea or osmotic stimuli and then act on specific receptors on airway smooth muscle to cause airway narrowing is shown. These mediators that are endogenously released in the airway, such as leukotrienes and prostaglandins, are significantly more potent than mediators that are exogenously administered to assess airway hyper-responsiveness, such as methacholine. Taken from O'Byrne.<sup>E384</sup>

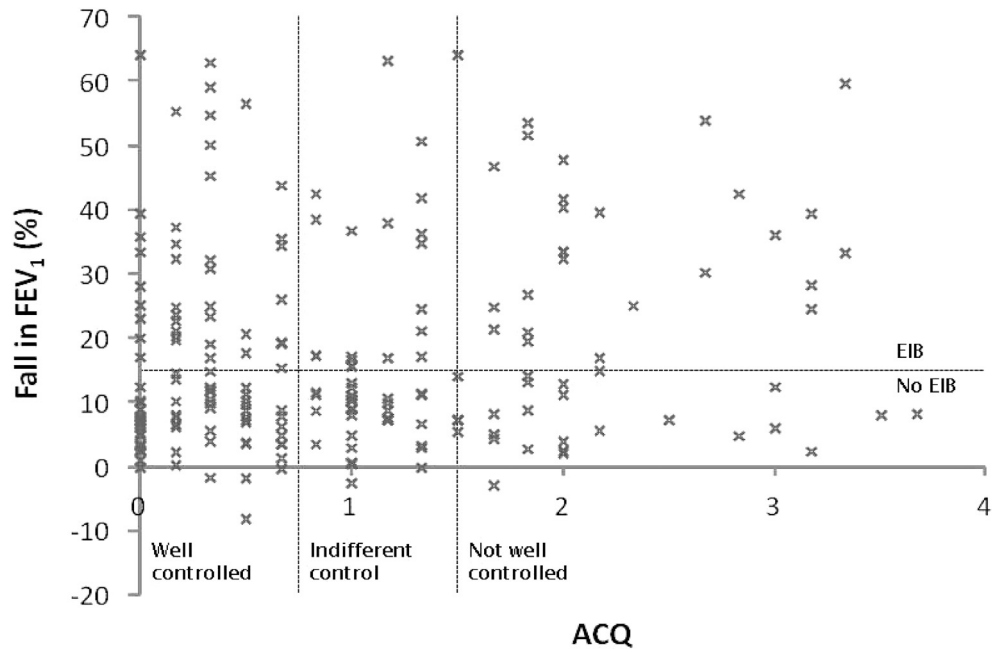




**FIG E3.** Relationship of airway sensitivity to EVH versus methacholine. The percentage decrease in FEV<sub>1</sub> to a methacholine challenge in relation to the percentage decrease in FEV<sub>1</sub> to EVH challenge in 131 adults reporting symptoms with exercise and no previous diagnosis of asthma. A positive response to EVH documented as a 10% decrease in FEV<sub>1</sub> identified BHR suggestive of EIB more frequently than a 20% decrease in FEV<sub>1</sub> to methacholine. *White dots* highlight those with a positive EVH challenge result and a negative methacholine challenge result. Adapted from Holley et al.<sup>E151</sup>



**FIG E4.** Importance of optimal exercise intensity: highlighting the importance of optimal exercise intensity when assessing EIB. Twenty asthmatic adolescent patients were exercised at 85% of estimated calculated HR<sub>max</sub>, resulting in only 9 of 20 having positive results for EIB (mean decrease FEV<sub>1</sub> = 8.84%); however, all 20 asthmatic patients had positive test results after exercise at 95% of HR<sub>max</sub> (mean decrease in FEV<sub>1</sub> = 25.11%). Adapted from Carlsen et al.<sup>E139</sup>



**FIG E5.** The poor relationship between asthma control measured by using the Asthma Control Questionnaire (ACQ) and the percentage decrease in FEV<sub>1</sub> after exercise challenge in 200 adolescents who demonstrated EIB when a 15% decrease in FEV<sub>1</sub> was documented. The cutoff values for asthma control are less than 0.75, representing controlled asthma, and greater than 1.5, representing uncontrolled asthma. Taken from Madhuban et al.<sup>E354</sup>

**TABLE E1.** Medication withdrawal schedule

Medication/activity/food	Recommended time to withhold before challenge testing	Evidence of maximum duration of protection on EIB	Reference
SABA (albuterol, turbutaline)	8 h	<6 h	E158, E159
LABA (salmeterol, eformoterol)	24 h*	12 h	E158, E160, E161
LABA in combination with an ICS (salmeterol/fluticasone, formoterol/budesonide)	24 h*	NA†	E153
Ultra-LABAs (indacaterol, olodaterol, vilanterol)	≥72 h‡	NA	
ICS (budesonide, fluticasone propionate, beclomethasone)	6 h	NA§	
Long-acting ICS (fluticasone furoate)	24 h	NA	
Leukotriene receptor antagonists (montelukast, zafirlukast)	4 d	24 h¶	E59, E62
Leukotriene synthesis inhibitors (zileuton/slow-release zileuton)	12 h/16 h	4 h	E160
Antihistamines (loratadine, cetirzine, fexofenadine)	72 h	<2 h#	E162
Short-acting muscarinic acetylcholine antagonist (ipratropium bromide)	12 h	<0.5 h	E163††
Long-acting muscarinic acetylcholine antagonist (tiotropium bromide, aclidinium bromide, glycopyrronium)	≥72 h‡	NA	
Cromones (sodium cromoglycate, nedocromil sodium)	4 h	2 h	E164, E165
Xanthines (theophylline)	24 h	NA**	
Caffeine	24 h‡‡	NA**	
Vigorous exercise	>4 h	<4 h	E79

NA, Not available.

\*A longer duration of withdrawal of up to 48 hours might be warranted in subjects who take regular LABAs to permit recovery of tolerance and prevent delay of recovery to a standard dose of rescue  $\beta_2$ -agonist.<sup>E166,E167</sup>

†Study demonstrates similar efficacy for a LABA acutely when included in combination with an ICS.

‡No evidence exists for the protective effects of ultra-LABAs or long-acting muscarinic acetylcholine antagonists on indirect tests. Thus we recommend testing after complete washout of drug, and for some ultra-long-acting drugs, washout can be up to 10 days. There is evidence of 36 hours of protection of an ultra-LABA on methacholine challenge.<sup>E168</sup>

§Evidence exists for high-dose ICSs to attenuate airway responses to indirect challenge tests in adults (eg, 1500  $\mu\text{g}/\text{d}$ ),<sup>E56</sup> with evidence for lower standard doses in children.<sup>E169</sup> Thus we recommend testing after complete washout of drug.

||Based on longer absorption half-life compared with fluticasone propionate.

¶Significant inhibition of EIB is observed at 24 hours after dosing. Thus we recommend testing after complete washout of drug.

#H<sub>1</sub>-antagonists demonstrate an overall weak inhibitory effect on EIB<sup>E54</sup>; however, they might have a modifying effect on the airway sensitivity (PD<sub>15</sub>) to mannitol.<sup>E55</sup> Thus we recommend testing after complete washout of drug.

\*\*In the absence of direct evidence, we recommend testing after complete washout of drug.

††Available data supporting withholding times are from studies assessing the duration of action of drugs on exercise challenge, unless otherwise indicated for EVH.

‡‡High doses of caffeine consumption (>3 cups of coffee) are most effective.<sup>E170</sup>